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Article

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Revisional notes on the monkey-moth genus *Stenoglene* Felder & Felder, 1874 and its allies, with descriptions of several new genera and species (Lepidoptera: Bombycoidea: Eupterotidae: Janinae)

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Abstract

The monkey-moth genus *Stenoglene* Felder & Felder, 1874 and its allies, *Epijana* Holland, 1893, *Drepanojana* Aurivillius, 1893, *Urojana* Gaede, 1915 and *Pterocerota* Hampson, 1905, are fully reviewed and redefined based on a synthesis of morphological and barcode data. *Stenoglene* is found to be polyphyletic and in order to maintain monophyly, ten new genera are established: *Artemisana* **gen. n.**, *Atopojana* **gen. n.**, *Cryptojana* **gen. n.**, *Hecateana* **gen. n.**, *Lokiana* **gen. n.**, *Microjana* **gen. n.**, *Neojana* **gen. n.**, *Nepenthojana* **gen. n.**, *Smithiana* **gen. n.** and *Streptojana* **gen. n.** Eighteen new species are described in ten genera: *Cryptojana volynkini* **sp. n.**, *Drepanojana hogani* **sp. n.**, *Hecateana dargei* **sp. n.**, *Hecateana giustii* **sp. n.**, *Hecateana kitchingi* **sp. n.**, *Lokiana lydiae* **sp. n.**, *Microjana bouyeri* **sp. n.**, *Microjana hikaru* **sp. n.**, *Microjana tabithae* **sp. n.**, *Pterocerota anima* **sp. n.**, *Smithiana smithi* **sp. n.**, *Stenoglene lucindae* **sp. n.**, *Stenoglene sarahae* **sp. n.**, *Stenoglene susanae* **sp. n.**, *Streptojana forbesi* **sp. n.**, *Streptojana laurenae* **sp. n.**, *Streptojana molliae* **sp. n.** and *Urojana basquini* **sp. n.** Seven species are revived from synonymy and forty-four new combinations are proposed: *Artemisana thelda* (Druce, 1887) **comb. n.**, *Atopojana bouyeri* (Basquin, 2013) **comb. n.**, *Cryptojana cinerea* (Holland, 1893) **comb. n.**, *Cryptojana clucki* (Bouyer, 2012) **comb. n.**, *Cryptojana decellei* (Dall'Asta & Poncin, 1980) **comb. n.**, *Cryptojana grandiplaga* (Holland, 1920) **stat. rev.**, **comb. n.**, *Cryptojana insignifica* (Rothschild, 1917) **comb. n.**, *Cryptojana maculifera* (Strand, 1912) **comb. n.**, *Cryptojana noellae* (Bouyer, 2012) **comb. n.**, *Cryptojana plagiata* (Aurivillius, 1911) **comb. n.**, *Cryptojana preussi* (Aurivillius, 1893) **comb. n.**, *Cryptojana shabae* (Dall'Asta & Poncin, 1980) **comb. n.**, *Drepanojana maeonia* (Druce, 1896) **stat. rev.**, *Epijana gigantea* (Rothschild, 1917) **comb. n.**, *Hecateana paulisi* (Dall'Asta & Poncin, 1980) **comb. n.**, *Lokiana sigyna* (Aurivillius, 1893) **comb. n.**, *Microjana annae* (Giusti & László, 2025) **comb. n.**, *Microjana brunneofasciata* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana citrina* (Druce, 1886) **comb. n.**, *Microjana citrinoides* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana dallastai* (Bouyer, 2012) **comb. n.**, *Microjana dehanica* (Strand, 1911) **comb. n.**, *Microjana dufranei* (Bouyer, 2012) **comb. n.**, *Microjana fontainei* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana fouassini* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana latimaculata* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana morettoi* (Bouyer, 2012) **comb. n.**, *Microjana novempunctata* (Bethune-Baker, 1927) **stat. rev.**, **comb. n.**, *Microjana opalina* (Druce, 1910) **comb. n.**, *Microjana parvula* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana pellucida* (Joicey & Talbot, 1924) **comb. n.**, *Microjana pujoli* (Dall'Asta & Poncin, 1980) **comb. n.**, *Microjana sinjaevi* (Bouyer, 2012) **comb. n.**, *Microjana smithi* (László & Giusti, 2025) **comb. n.**, *Microjana subiridescens* (Holland, 1893) **comb. n.**, *Microjana sulphureoides* (Kühne, 2007) **comb. n.**, *Microjana sulphureotincta* (Strand, 1912) **comb. n.**, *Microjana uelei* (Dall'Asta & Poncin, 1980) **comb. n.**, *Neojana pseudosygina* (Bouyer & Basquin, 2012) **comb. n.**, *Nepenthojana plenevauxae* (Bouyer, 2012) **comb. n.**, *Nepenthojana uniformis* (Dall'Asta & Poncin, 1980) **comb. n.**, *Pterocerota*

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perissinottoi (Bouyer, 2012) **comb. n.**, *Smithiana bipartita* (Rothschild, 1917) **comb. n.**, *Stenoglene elgonae* (Bethune-Baker, 1927) **stat. rev.**, *Stenoglene gregorii* (Butler, 1894) **stat. rev.**, *Stenoglene labda* (Druce, 1887) **stat. rev.**, *Stenoglene similis* (Rothschild, 1917) **stat. rev.**, *Streptojana brunnea* (Bouyer, 2012) **comb. n.**, *Streptojana nivalis* (Rothschild, 1917) **comb. n.** Six synonymies are newly established or revived: *Phasicnecus livingstonensis* (Strand, 1909) **syn. n.** considered a synonym of *Stenoglene labda*, *Phasicnecus aequidistans* Strand, 1915 **syn. n.** considered a synonym of *Stenoglene pira* Druce, 1896, *Phasicnecus pulverulentus* Joicey & Talbot, 1924 **syn. n.** and *Phasicnecus similis flavidior* Rothschild, 1917 **syn. rev.** considered synonyms of *Stenoglene similis*, *Stenoglene decellei flava* Dall'Asta & Poncin, 1980 **syn. n.** considered a synonym of *Cryptojana decellei*, and *Hoplojana purpurata* Wichgraf, 1921 **syn. n.** considered a synonym of *Epijana gigantea*. The hitherto unknown female of *Urojana eborea* Gaede, 1915, is described and illustrated based on a recently-collected, barcode-confirmed specimen. Diagnostic morphological characters, namely the forewing venation and male genitalia are reviewed, and insights into the biogeography of this group of moths are briefly discussed in light of the taxonomic rearrangement and accurate delimitation of genera.

Key words Afrotropics, *Epijana*, *Drepanojana*, *Urojana*, *Pterocerota*, integrative taxonomy, new synonymy, revived synonymy, new combination.

Introduction

The monkey-moth genus *Stenoglene* Felder & Felder, 1874 comprises many beautiful and charismatic species, and is widely distributed throughout continental Africa. The genus currently contains 52 species and one subspecies (Basquin 2013; Kitching & Rougerie *et al*. 2018; Basquin & Faravel 2025; László *et al*. 2025;), and is the most speciose of the 16 Janinae genera (Nässig & Oberprieler 2008). Its constituents are a seemingly heterogenous mix of species in terms of size, wing shape and phenotype, and subdividing them into species-groups has proved difficult (e.g., Forbes 1955). Of the two species initially described in *Stenoglene*, *S. hilaris* Felder & Felder, 1874 from South Africa and *S. tristis* Felder & Felder, 1874 from Sri Lanka (*patria falsa*), the former was designated as the type species by Kirby (1892) who placed the genus in the Lasiocampidae. It was transferred to the Eupterotidae by Aurivillius (1893) but later in his revisional work on the family (Aurivillius 1901), he considered *Stenoglene* to be a *nomen nudum* due to the lack of a description and the type species being unknown to him. Here, Aurivillius (1901) tentatively synonymised the genus *Phasicnecus* Butler, 1894 with its type species *P. gregorii* Butler, 1894 under *Stenoglene* but continued to use the former generic name for the 13 taxa he placed in this group. Gaede (1927) in his review of the Eupterotidae, followed suit by referring only to *Phasicnecus* but omitted the name *Stenoglene* entirely. It was not until Forbes (1955) that *Stenoglene* was correctly treated as the senior name and *Phasicnecus* synonymised under it. The most comprehensive review of the genus was undertaken by Dall'Asta & Poncin (1980) who described 14 new species and one new subspecies, while illustrating all other species that had not been figured in previous works (e.g., Gaede 1927). More recently, a further 17 species have been described in four works (Kühne 2007; Bouyer 2012; Basquin 2013; László *et al*. 2025).

Despite the phenotypic disparity in *Stenoglene* and suggestions that the generic classification was unsatisfactory (Bouyer 2012), the monophyly of the genus has never been tested. Only a single *Stenoglene* taxon was included in the Bombycoidea phylogeny of Hamilton *et al*. (2019), while barcodes of two taxa were included in Naumann *et al.* (2020) although collapsed branches on their neighbour-joining tree rendered any results uninformative. Several other allied genera require consideration in this current body of work due to similarities in genital configurations, and this opportunity is taken to redescribe these genera, some of which have not been studied for a century. These include *Epijana* Holland, 1893, a genus which was considered an "immediate derivative" of *Stenoglene* by Forbes (1955: 95), *Pterocerota* Hampson, 1905 suggested as being closely related by Minet (1990, 1994), and *Drepanojana* Aurivillius, 1893 and *Urojana* Gaede, 1915, both considered to be near *Stenoglene* by Gaede (1927). Two other genera with affinities to *Stenoglene*, *Vianga* Fletcher, 1982 and *Gonojana* Aurivillius, 1893 have already been reviewed by the present author (Takano 2024a).

As part of investigations into the rich Eupterotidae material held in the collections of the African Natural History Research Trust (ANHRT), a large proportion of the Janinae were barcoded and numerous dissections undertaken which allowed for a broad overview of all genera. Based on the synthesis of morphological and molecular markers, the genus *Stenoglene sensu str.* is redefined with

implications on taxonomy and nomenclature. In order to maintain discrete monophyletic units, ten new genera are established and numerous new combinations are implemented as a result of the subsequent taxonomic rearrangements. Furthermore, genetic and morphological analyses have revealed a hidden diversity of species and the descriptions of 18 new species are provided herein.

Materials and methods

To test the monophyly of *Stenoglene*, DNA barcodes were obtained by removing tarsal segments from 133 adult specimens (122 *Stenoglene*, 6 *Epijana*, 3 *Drepanojana* and 2 *Urojana*, including type species of all genera) and submitting them to the Canadian Centre for DNA Barcoding (CCDB, Biodiversity Institute of Ontario, University of Guelph). *Acrojana* Aurivillius, 1901, which was recovered as sister to *Stenoglene* in the phylogeny of Hamilton *et al*. (2019), was selected as the outgroup taxon. Sequences were obtained using Single Molecule Real-Time sequencing through the Sequel II (PacBio) pipeline at CCDB (Hebert *et al*. 2018) and, together with 53 publicly available sequences from the BOLD (https://boldsystems.org) database, were aligned using MUSCLE in MEGA version X (Kumar *et al*. 2018). Genetic distances were calculated using Kimura's two-parameter model (Kimura 1980). The final dataset comprised 162 individuals of *Stenoglene*, 16 of *Epijana*, and four each of *Drepanojana* and *Urojana*. Phylogenetic tree searches were performed using Bayesian Inference (BI) and Maximum Likelihood (ML). BI analyses were performed using MrBayes version 3.2.7a (Ronquist *et al*. 2012). Metropolis-coupled Markov chain Monte Carlo (MCMC) analyses were run with four chains (one cold and three heated) for 10,000,000 generations sampling every 100 generations, discarding the first 25% as burn-in. The two runs converged with the standard deviation of split frequencies 0.007. ML analyses were performed using raxmlGUI 2.0 (Edler *et al*. 2021) with the model parameters optimised for the dataset using the Bayesian Information Criterion (HKY+GAMMA+I). Support for clades was evaluated for BI using posterior probabilities (PP) and ML using rapid bootstrapping (BS) with 1000 replicates. Trees were visualised and annotated in FigTree version 1.4.4 and Adobe Photoshop. All voucher specimens are dry-mounted and labelled with unique identifiers. Barcodes of type specimens generated as part of this study are tabulated in Table 1 and will be made publicly available.

Preparation of genitalia followed Lafontaine & Mikkola (1987), the phalluses were weakly stained with Eosin red and the dissected genitalia were mounted on microscope slides or stored in glycerol. Genitalia were photographed using a Canon EOS 700D camera mounted on a Leitz Diaplan compound microscope or a Wild M3Z stereo microscope. Adults were photographed using a Canon EOS 80D with a Canon 100 mm Macro lens. All images were edited in Adobe Photoshop.

Primary label data have been transcribed verbatim with "||" denoting a different label and "|" denoting a line break. All name-bearing type specimens have been examined unless stated otherwise (in parentheses). Taxa are listed alphabetically under each genus. Venation follows Forbes (1955: 86, fig. 1) and genital terminology follows Oberprieler *et al*. (2003) and Zwick (2009).

Institutional abbreviations:

AMNH	American Museum of Natural History, New York, U.S.A.
ANHRT	African Natural History Research Trust, Leominster, U.K.
CMNH	Carnegie Museum of Natural History, Pittsburgh, U.S.A.
MfN	Museum für Naturkunde, Berlin, Germany
MNHN	Museum national d'Histoire Naturelle, Paris, France
NHMUK	Natural History Museum, London, U.K.
NHMW	Naturhistorisches Museum, Vienna, Austria
OUMNH	Oxford University Museum of Natural History, Oxford, U.K.
RBINS	Royal Belgian Institute of Natural Sciences, Brussels, Belgium
RCMO	Research Collection of Michael Ochse, Weisenheim am Berg, Germany
RCPB	Research Collection of Patrick Basquin, Yvetot-Bocage, France
RMCA	Royal Museum for Central Africa, Tervuren, Belgium
ZIMG	Zoologische Institut und Museum, Greifswald, Germany
ZSM	Zoologische Staatssammlung, Munich, Germany

Table 1. List of type specimen barcodes generated as part of this current study. HT = holotype, PT = paratype, BIN = Barcode Index Number and bp = base pairs. All specimens deposited in ANHRT.

Taxon	Type	Sex	Process ID	Locality	BIN	bp
Stenoglene lucindae sp. n.	HT	♂	ANLMO6540-23	Tanzania, Biharamulo, 26.x.2008	BOLD:AFG3494	651
Stenoglene lucindae sp. n.	PT	♂	ANLMO6537-23	Tanzania, Samina, 1294 m, 20.x.2011	BOLD:AFG4210	653
Stenoglene lucindae sp. n.	PT	♂	ANLMO6538-23	Tanzania, Samina, 1294 m, 20.x.2011	BOLD:AFG4210	653
Stenoglene lucindae sp. n.	PT	♀	ANLMO6541-23	Tanzania, Samina, 1294 m, 20.x.2011	BOLD:AFG4210	653
Stenoglene sarahae sp. n.	HT	♂	ANLMO5606-23	Zambia, Kapishya Hot Springs, 1437 m, 22.xi–8.xii.2015	BOLD:AAO9401	653
Stenoglene sarahae sp. n.	PT	♂	ANLMO4382-23	Zambia, Kalungu, 1280 m, 22–24.xi.2016	BOLD:AAO9401	656
Stenoglene sarahae sp. n.	PT	♀	ANLMO4384-23	Zambia, Kalungu, 1280 m, 22–24.xi.2016	BOLD:AAO9401	653
Stenoglene sarahae sp. n.	PT	♀	ANLMO4386-23	Zambia, Kalungu, 1280 m, 22–24.xi.2016	BOLD:AAO9401	655
Stenoglene susanae sp. n.	HT	♂	ANLMO6386-23	Zambia, Kalene Hill, 1440 m, 27.xi–03.xii.2020	BOLD:AFI0443	648
Stenoglene susanae sp. n.	PT	♂	ANLMO6532-23	Zambia, Kambishi,1340 m, 18–19.x.2014	BOLD:AFI0443	651
Cryptojana volynkini sp. n.	HT	♂	ANLMO6559-23	Tanzania, Minziro Forest, 1176 m, 10.x.2012	BOLD:AAE1933	653
Microjana hikaru sp. n.	HT	♂	ANLMO4137-23	Congo, Nouabalé-Ndoki NP, 352 m, 29.ix–01.x.2022	BOLD:AAM8653	653
Microjana hikaru sp. n.	PT	♂	ANLMO4142-23	Congo, Nouabalé-Ndoki NP, 352 m, 29.ix–01.x.2022	BOLD:AAM8653	654
Microjana hikaru sp. n.	PT	♂	ANLMO4147-23	Congo, Nouabalé-Ndoki NP, 352 m, 29.ix–01.x.2022	BOLD:AAM8653	653
Microjana hikaru sp. n.	PT	♀	ANLMO4175-23	Congo, Nouabalé-Ndoki NP, 372 m, 01–10.x.2022	BOLD:AAM8653	654
Streptojana forbesi sp. n.	PT	♂	ANLMO5098-23	Liberia, Wologizi Mts, 883 m, 22–31.xi.2018	BOLD:AFF4367	654
Streptojana forbesi sp. n.	PT	♂	ANLMO5369-23	Liberia, Wonegizi NR, 551 m, 19–27.iii.2019	BOLD:AFF4367	652
Streptojana forbesi sp. n.	PT	♂	ANLMO6156-23	Liberia, Wologizi Mts, 585 m, 18.xi–1.xii.2018	BOLD:AFF4367	653
Streptojana forbesi sp. n.	HT	♂	ANLMO6197-23	Liberia, Wologizi Mts, 883 m, 22–31.xi.2018	BOLD:AFF4367	667
Streptojana laurenae sp. n.	HT	♂	ANLMO5587-23	D.R. Congo, Mamove, 950 m, ix.2015	BOLD:AFG4352	654
Smithiana smithi sp. n.	PT	♂	ANLMO5784-23	Liberia, Wologizi Mts, 611 m, 20.xi–01.xii.2017	BOLD:AAP9523	653
Smithiana smithi sp. n.	PT	♂	ANLMO6149-23	Liberia, Wologizi Mts, 883 m, 22–31.xi.2018	BOLD:AAP9523	654
Lokiana lydiae sp. n.	PT	♂	ANLMO4424-23	Côte d'Ivoire, Tai NP, 174 m, 25.iii–17.iv.2017	BOLD:AAO6229	654
Lokiana lydiae sp. n.	PT	♀	ANLMO5368-23	Liberia, Wonegizi NR, 551 m, 19–27.iii.2019	BOLD:AAO6229	654
Lokiana lydiae sp. n.	PT	♂	ANLMO6152-23	Liberia, Wologizi Mts, 585 m, 18.xi–01.xii.2018	BOLD:AAO6229	654
Lokiana lydiae sp. n.	PT	♂	ANLMO6192-23	Liberia, Wologizi Mts, 883 m, 22–30.xi.2018	BOLD:AAO6229	656
Hecateana kitchingi sp. n.	HT	♂	ANLMO4403-23	Côte d'Ivoire, Tai NP, 174 m, 25.iii–17.iv.2017	BOLD:AAO5978	653

Results

Synthesis of molecular and morphological data

Phylogenetic inferences based on BI and ML were largely congruent wherein *Stenoglene* was recovered as polyphyletic with regard to *Epijana*, *Drepanojana* and *Urojana* (ML tree figured in Fig. 1). In the BI analysis, many of the major lineages were recovered as polytomies, but in both analyses, each major lineage was recovered with strong support (PP >97). In order to maintain monophyly, the establishment of several new genera is required and each lineage corresponds to a new genus that can be diagnosed by morphological characters. The following three clades were recovered in both analyses: *Smithiana* **gen. n.** sister to *Lokiana* **gen. n.** + (*Epijana* + *Urojana*) (BS: 20, PP: 68), *Neojana* **gen. n.** sister to *Drepanojana* (BS: 29, PP: 56), and *Atopojana* **gen. n.** sister to *Hecateana* **gen. n.** (BS: 53, PP: 100), albeit most with weak support. This is perhaps unsurprising considering the disparate genital morphology and phenotypes of all these genera. One species of *Stenoglene*, *S. gigantea* (Rothschild, 1917) was recovered within the *Epijana* clade and is herein assigned to the latter (*Epijana gigantea* **comb. n.**) based on congruent morphological characteristics, most notably the forewing venation.

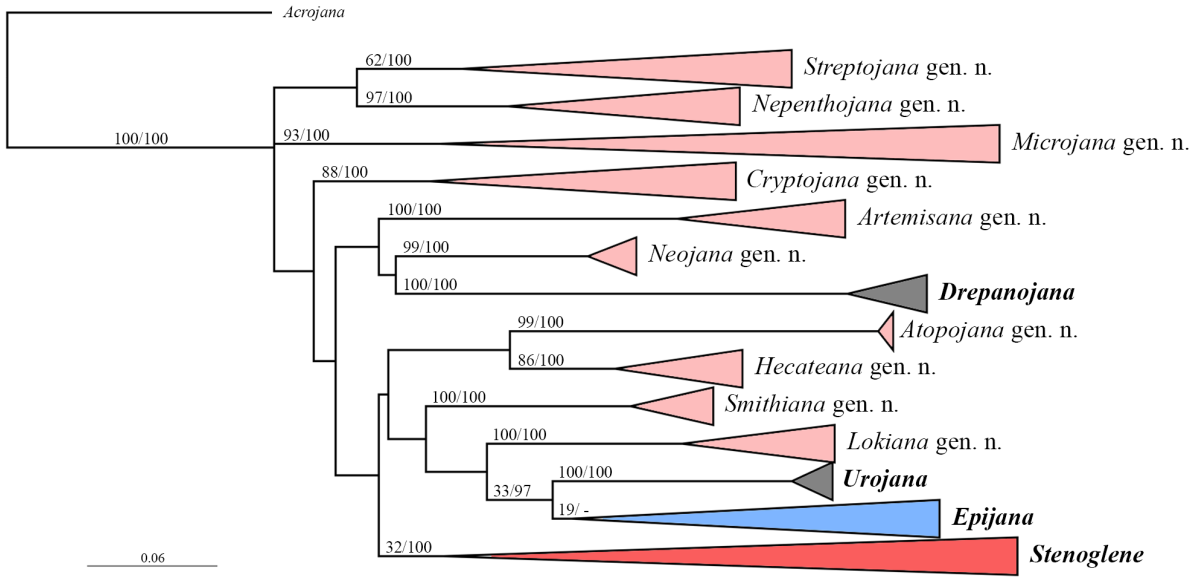


Figure 1. Maximum likelihood cladogram with collapsed branches of *Stenoglene* Felder & Felder, 1874 *sensu lato* and allied genera, with *Acrojana* as outgroup, based on DNA barcodes. *Stenoglene sensu str.* is highlighted in bold red, other taxa previously assigned to *Stenoglene* (and now new genera) highlighted in pale red, *Epijana* Holland, 1893 highlighted in blue, and *Drepanojana* Aurivillius, 1893 and *Urojana* Gaede, 1915 highlighted in grey. Numbers above branches indicate support values (bootstrap / posterior probabilities). Scale bar indicates substitution rates per site.

Forbes (1955) stated that the Janinae genera were distinguished based mainly on venation, although he concluded that it alone could not wholly discriminate between *Epijana* and *Stenoglene*, listing several species of the latter that displayed identical venation to the former. While venation was heavily utilised for higher-level classification of Lepidoptera in the early 20th century, its suitability was doubted by Mell (1930) and it has been shown that it is not always a reliable generic character in the Afrotropical Janinae (Takano 2025). For all of the genera reviewed below, some intraspecific variation was observed in the venation, most noticeable between males and females of the same species, and sometimes even at the individual level. However, the newly redefined *Stenoglene* and *Epijana* display constantly differing venation, at least in the males, which can be used to diagnose these genera. For the majority of the genera reviewed in this present work, including *Stenoglene*, forewing veins R_4 + R_5 are stalked, with the exception of *Epijana*, *Urojana* and *Pterocerota* where R_4 is placed beyond R_5 and are thus unstalked (Fig. 2). *Pterocerota* differs from *Epijana* and *Urojana* in the positioning of forewing vein M_1 which arises from the discocellular rather than the radial veins (Fig. 2), a configuration

inadvertently noticed by Bouyer (2012) in the original description of *Stenoglene perissinottoi* Bouyer, 2012, herein assigned to the genus *Pterocerota* (*P. perissinottoi* **comb. n.**). The radial and medial veins were incorrectly interpreted by László *et al*. (2025), their R_4 and R_5 referable instead to R_5 and M_1 respectively. Forbes (1955: Fig. 1) rather unfortunately selected *Epijana gigantea* as a typical *Stenoglene* to illustrate the venation in his work on the Eupterotidae, having grouped it with *Stenoglene gregorii* based on wing pattern, while also believing it could possibly be synonymous with *Stenoglene similis* (Rothschild, 1917). The venation in Forbes' (1955) figure is virtually identical with Holland's (1893: 514) illustration in the original description of *Epijana*, but perhaps the structural differences in the male genital capsule outweighed any observed similarities in vein configuration. The venation of a true *Stenoglene sensu str*. species, *S. rosea* (Druce, 1886), is illustrated here in Fig. 2 for comparison. Forbes (1955) also stated that *Stenoglene cinerea* displayed the same vein configuration as *Epijana* but examination of male specimens revealed $R_4 + R_5$ to be stalked, albeit with a very short stalk.

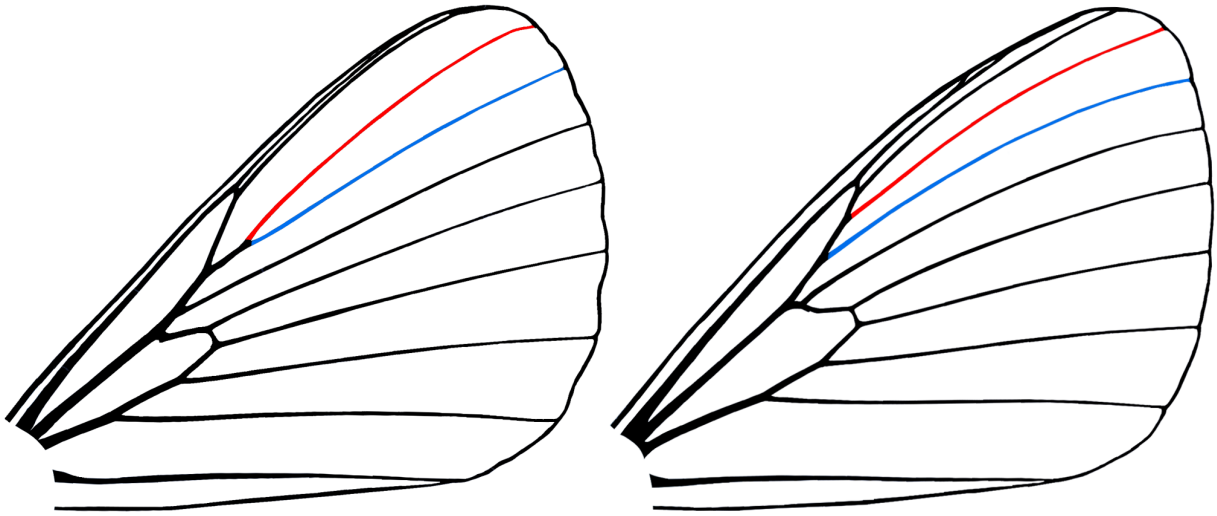


Figure 2. Forewing venation of *Stenoglene rosea* (Druce, 1886) (left) and *Pterocerota perissionottoi* (Bouyer, 2012) (right) showing the differing configurations of vein R_4 (in red) and R_5 (in blue).

Forbes (1955) was the first to classify the Eupterotidae based on characters of the genitalia, examining six *Stenoglene* and one *Epijana* species in the process and provided detailed descriptions and figures of *S. gregorii* and *E. cosima* (Plötz, 1880). Forbes (1955) referred to projections of the dorsal section of the genital capsule as processes of the tegumen in *Stenoglene* and an uncus in *Epijana*, whilst Dall'Asta & Poncin (1980) for the most part and Oberprieler *et al*. (2003) referred to the same structures as belonging entirely to the uncus. More recently, the reference to the eighth tergite for these characters by Bouyer (2012) is fundamentally erroneous, with the tegumen belonging to the ninth tergum and the uncus the tenth. In this present work, examination of the genitalia of over 50 taxa has revealed that all processes belong to the uncus, which in turn is firmly fused to the tegumen sometimes with very indiscernible fusion lines (cf. Oberprieler *et al*. 2003). Another character that requires clarification is the sensillae-bearing structure present between the valve and the juxta. Forbes (1955) referred to the "clavus" as being represented by a pair of "irregular pilose sclerites" in *S. gregorii* while it was reduced in *S. subiridescens* and that they were "not visibly connected to each other" or the juxta in the latter. In *Epijana*, he referred to a "pair of setose chitinous structures representing the sides of the transtilla…connected by the narrow chitinous transtilla". Dall'Asta & Poncin (1980: 458) referred to these structures as "lobes of the transtilla", but the transtilla in its strict sense (i.e. *sensu* Pierce 1914) is not present in Bombycoidea and likely refer instead to paired mesal extensions of the valva apodemes (Zwick 2009). More recently, László *et al*. (2025) erroneously referred to these as basal lobes of the tegumen, with which they are not articulated. However, for this present work, the term "mesal protrusions", first introduced for the Anthelidae by Zwick (2009: 156), is adopted, with these setose protuberances located distad (= mesally, towards the anellus) of the valve apodeme. These structures are treated as being homologous across all the genera outlined below, and can be heavily attenuated and reduced (e.g., in *Microjana* **gen. n.**) or form large, sclerotised and angled lobes fused medially (e.g., in

E. cosima and some *Streptojana* **gen. n.** species). The extent of sclerotisation of the mesal protrusions or their fusion are seemingly not generic synapomorphies, instead only being useful to distinguish species.

Taxonomy

***Stenoglene* Felder & Felder, 1874**

Stenoglene Felder & Felder 1874: pl. 95, figs 3, 4; Kirby 1892: 809; Aurivillius 1893: 206; 1901: 28; Forbes 1955: 95, 129; Dall'Asta & Poncin 1980: 466; Fletcher & Nye 1982: 154; Vári *et al*. 2002: 150; Nässig & Oberprieler 2008: 57, 71; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 137.

Type species: *Stenoglene hilaris* Felder & Felder, 1874, by subsequent designation (Kirby 1892: 809).

Phasicnecus Butler, 1894

Phasicnecus Butler 1894: 585; Aurivillius 1901: 21, 28; Gaede 1927: 307; Forbes 1955: 95, 129 (syn.); Pinhey 1975: 131.

Type species: *Phasicnecus gregorii* Butler, 1894, by monotypy.

Redescription. Adults. Small- to medium-sized moths with colouration varying from yellow and orange through to olive-brown. Antennae bipectinate, rami shorter in females. Forewing generally elongate with arcuate outer margin; $R_4 + R_5$ stalked. Discal marking always absent. Single or double postmedial and submarginal fasciae curved (the latter sometimes oblique) in shades of dark brown to red, as well as submarginal dark spots in spaces Cu_1 and Cu_2. Hindwing uniform lacking all fasciae except for dark spots along anal margin. Underside as on upperside but often uniform lacking markings. For those species where females are known, they display some sexual dimorphism mainly in terms of size and colour, although some differ in wing shape. Male genitalia. Uncus with medial or lateral projections, usually bifid or trifid. Tegumen broad. Gnathos ring-shaped, thin and thread-like. Mesal protrusions separated, well-developed, rounded or trapezoidal, densely covered in setae. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section triangular or ovoid, tapering to distal process, apically pointed and recurved in some species. Saccular section triangular, broad, tapering to long, apically pointed distal process, often as long as sacculus. Juxta semi-cylindrical, trapezoidal and bifurcate dorsally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded (curved dorsad in most species), carina heavily sclerotised and tapered; vesica with small patch of scobination.

Diagnosis. *Stenoglene sensu str.* are yellow, orange or brown in colouration and the hindwing is uniform lacking all fasciae. This character separates similarly coloured species in the genus *Microjana* **gen. n.** which are smaller in size and possess more rounded wings as well as well-defined arcuate postmedial and submarginal fasciae. Some species of *Epijana* also lack hindwing fasciae but are generally larger and forewing veins R_4 and R_5 are always unstalked. In the male genitalia, the uncus takes various forms but in general, the mesal protrusions are large and well-defined, and the valves are simple with an ensiform distal saccular process.

Distribution. Members of this genus are widespread in forest and forest-savanna mosaics throughout Africa, the majority of species found in more open woodland habitats.

Notes. The gender of *Stenoglene* is treated here as feminine (ICZN (1999), Article 30.1), the generic name deriving from the adjective *stenos* (στενός) meaning "narrow" and the feminine noun *glene* (γλήνη) meaning "pupil" presumably referring to the habitus of the two species originally placed in the genus. The fact that these two taxa (*S. hilaris* and *S. tristis*) derived their names from third-declension Latin adjectives where the nominative singular is the same for both genders may be one reason for recent works listing several names declined in the masculine (Dall'Asta & Poncin 1980; Varí *et al*. 2002; Bouyer 2012; Krüger 2020; László *et al*. 2025) and in accordance with Article 34.2 of the ICZN (1999) code, these species-group names are emended to agree in gender. Although many Lepidoptera databases and catalogues (e.g., Kitching & Rougerie *et al*. 2018) have ceased to follow this rule in favour of original orthography (van Nieukerken *et al*. 2019), the aforementioned articles are

adhered to strictly by the present author in accordance with the ICZN (1999) code until such time as consistent changes are implemented in a new version.

Species content: 16 species are assigned to this genus, three of which are new species.

***Stenoglene basquini* Bouyer, 2012**
Stenoglene basquini Bouyer 2012: 5, pl. 3, fig. 16; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 25.

Type locality: "Centrafrique, 4°05N, 16°38E [sic], route Mambele-Mbaéré".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Stenoglene bipunctata* (Aurivillius, 1909)**
Phasicnecus bipunctatus Aurivillius 1909: 9, fig. 42; Gaede 1927: 308.
Stenoglene bipunctata (Aurivillius, 1909): Forbes 1955: 129 (comb.).
Stenoglene bipunctatus (Aurivillius, 1909): Bouyer 2012: 4; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20; Basquin & Faravel 2025: pl. 28.
Stenoglene bipunctatus bipunctatus (Aurivillius, 1909): Dall'Asta & Poncin 1980: 482, photo 1, fig. 15.

Type locality: [D.R. Congo] "Bayenghé" [= Boyenge].
Name-bearing type material: holotype ♂ in RBINS (not examined).

***Stenoglene elgonae* (Bethune-Baker, 1927) stat. rev.**
Phasicnecus elgonae Bethune-Baker 1927: 327.
Stenoglene elgonae (Bethune-Baker, 1927): Forbes 1955: 129 (comb.).
Stenoglene roseus auct. (nec Druce): Dall'Asta & Poncin 1980: 485 (syn.); Vári *et al*. 2002: 151; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.
Stenoglene sp.?: Basquin 2024: 22.
Stenoglene rosea auct. (nec Druce): Basquin & Faravel 2025: pl. 38.

Type locality: "Mount Elgon, Uganda Protectorate".
Name-bearing type material: holotype ♀ in NHMUK.

Notes. Based on barcode divergences and differing male genital configurations, *S. elgonae* **stat. rev.** is revived from synonymy with *S. rosea*. This taxon is referrable to the population found in the Sudanian savannas from Senegal to Uganda, but further investigations are required to understand whether there is more than one taxon involved.

***Stenoglene gregorii* (Butler, 1894) stat. rev.**
Phasicnecus gregorii Butler 1894: 586, pl. 37, fig. 6; Aurivillius 1901: 29; Gaede 1927: 308, pl. 47.
Stenoglene gregorii (Butler, 1894): Forbes 1955: 95, 129, figs 11, 38, 39 (comb.).
Phasicnecus roseus auct. (nec Druce): Pinhey 1975: 131 (syn.).
Stenoglene roseus auct. (nec Druce): Dall'Asta & Poncin 1980: 485; Vári *et al*. 2002: 151; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.

Type locality: [Kenya] "Sabaki Valley".
Name-bearing type material: holotype ♂ (and not a ♀ as stated in the original description) in NHMUK.

Notes. Based on differing genital configurations, *S. gregorii* **stat. rev.** is revived from synonymy with *S. rosea*. This taxon is referrable to the population found in south-eastern Kenya and coastal northern Tanzania.

***Stenoglene hilaris* Felder & Felder, 1874** (Figs 3–4, 17)
Stenoglene hilaris Felder & Felder 1874: pl. 95, fig. 3; Kirby 1892: 809; Forbes 1955: 95, 129; Vári *et al*. 2002: 150; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138; Takano & László 2022: 13, figs 53, 54; Basquin 2024: 22.
Phasicnecus hilaris (Fleder & Felder, 1874): Aurivillius 1901: 30 (comb.); Janse 1917: 81; Gaede 1927: 309.

Type locality: [South Africa] "Port natal [sic]" [= Durban].
Name-bearing type material: holotype ♀ in NHMUK.

Chrysopoloma bithynia Druce, 1887
Chrysopoloma bithynia Druce 1887: 676.
Phasicnecus bithynia (Druce, 1887): Aurivillius 1901: 30 (comb.); Janse 1917: 81; Gaede 1927: 309, pl. 47.
Stenoglene bithynia (Druce, 1887): Kirby 1892: 810 (comb.); Forbes 1955: 95, 129; Dall'Asta & Poncin 1980: 485; Vári *et al*. 2002: 150 (syn.).

Type locality: [Mozambique] "Delagoa Bay" [= Maputo Bay].
Name-bearing type material: 3 ♂ and 5 ♀ syntypes in NHMUK.

***Stenoglene inversa* (Gaede, 1927)**
Phasicnecus inversus Gaede 1927: 328, pl. 47.
Stenoglene inversa (Gaede, 1927): Forbes 1955: 129 (comb.).
Stenoglene bipunctatus inversus (Gaede, 1927): Dall'Asta & Poncin 1980: 482 (stat.).
Stenoglene inversus (Gaede, 1927): Bouyer 2012: 4 (stat.); Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [Malawi] "Nyassa-Gebiet" [Zomba, according to label data].
Name-bearing type material: holotype ♂ in NHMUK.

***Stenoglene kenyensis* Bouyer, 2012**
Stenoglene kenyensis Bouyer 2012: 5, pl. 3, fig. 18; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Kenya Coast, East of Garsen, W. Witu".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Stenoglene labda* (Druce, 1887) stat. rev.**
Chrysopoloma labda Druce 1887: 677.
Phasicnecus labda (Druce, 1887): Aurivillius 1901: 30 (comb.); Gaede 1927: 308.
Stenoglene labda (Druce, 1887): Kirby 1892: 810 (comb.); Forbes 1955: 129.
Stenoglene obtusus auct. (nec Walker): Dall'Asta & Poncin 1980: 483 (syn.); Kitching & Rougerie *et al*. 2018: suppl. 1.
Stenoglene obtusa auct. (nec Walker): Vári *et al*. 2002: 150; Krüger 2020: 138.

Type locality: [Malawi] "Nyassa".
Name-bearing type material: holotype ♂ in NHMUK.

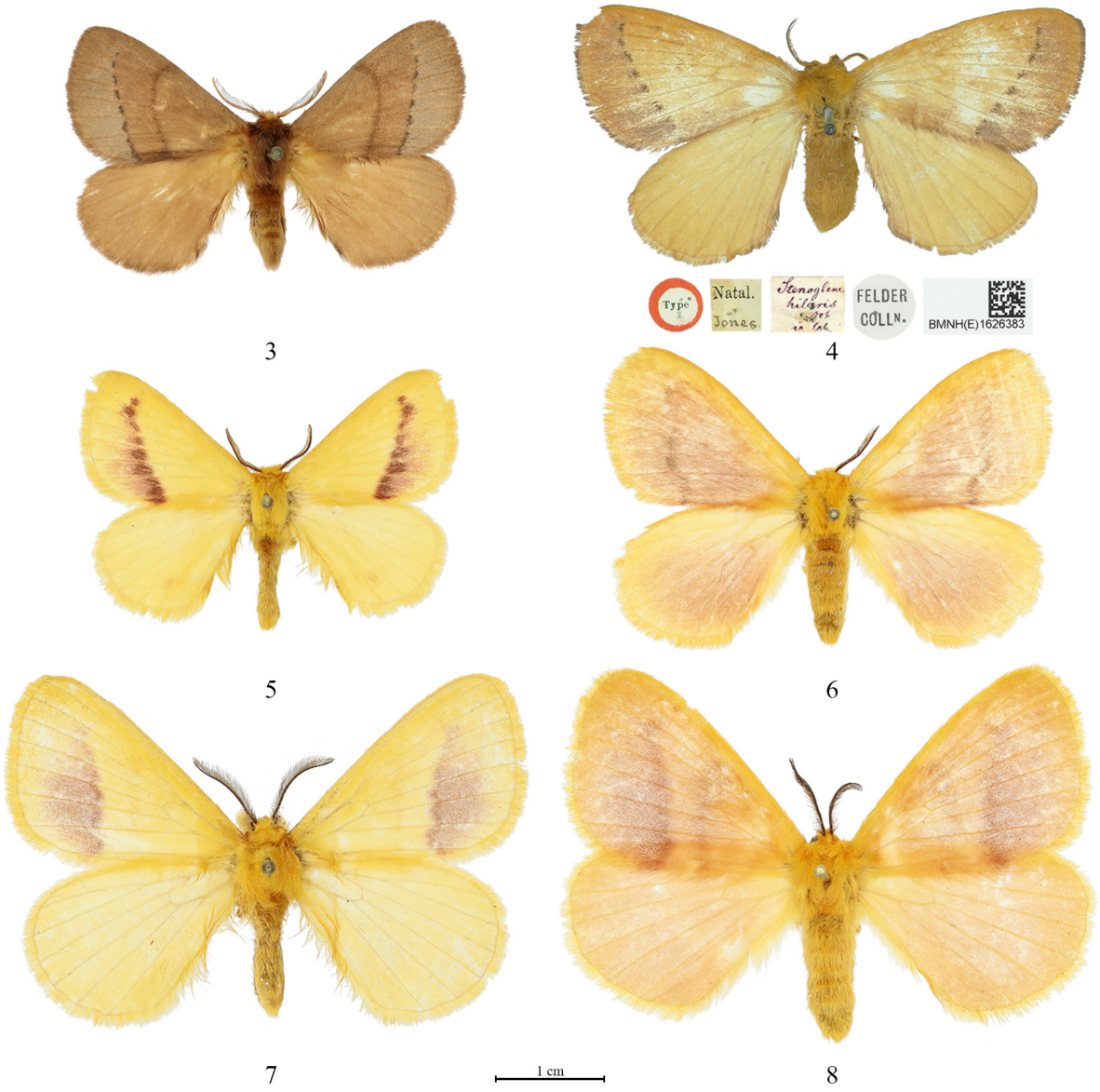
Notes. Based on barcode divergences of topotypical specimens and differing genital configurations, *S. labda* **stat. rev.** is revived from synonymy with *S. obtusa*. True *S. obtusa* is seemingly found only in South Africa and adjoining areas of coastal Mozambique, while *S. labda* is found broadly along the Zambezi Valley from Zambia, Zimbabwe and Mozambique into southern Malawi.

Phasicnecus livingstonensis (Strand, 1909) **syn. n.**
Phasicnecus livingstonensis Strand 1909: 379; Gaede 1927: 308, pl. 41.

Stenoglene livingstonensis (Strand, 1909): Forbes 1955: 129 (comb.); Vári *et al*. 2002: 150; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138.

Type locality: [Zambia] "Livingstone".
Name-bearing type material: holotype ♂ in MfN.

Notes. Considered to be a valid species by Varí *et al*. (2002) and later authors, examination of the holotype together with barcodes of topotypical specimens and genital dissections suggests *Phasicnecus livingstonensis* **syn. n.** is conspecific with *S. labda*.



Figures 3–8. *Stenoglene* Felder & Felder, 1874 species, habitus (in ANHRT unless otherwise stated). 3. *S. hilaris* Felder & Felder, 1874, ♂, Mozambique, Maputo Special Reserve. 4. *Id*., holotype ♀ (NHMUK). 5. *S. rosea* (Druce, 1886), ♂, Mozambique, Maputo Special Reserve. 6. *Id*., ♀. 7. *S. similis* (Rothschild, 1917), ♂, Zambia, Jiwundu Swamp. 8. *Id*., ♀, Zambia, Kambishi School.

Stenoglene leonardi Bouyer, 2012

Stenoglene leonardi Bouyer 2012: 5, pl. 3, fig. 17; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Guinée Conakry, forêt classée de Diéké".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Stenoglene obtusa* (Walker, 1865)** (Figs 9–10, 20)
Poloma obtusa Walker 1865: 357.
Phasicnecus obtusus (Walker, 1865): Aurivillius 1901: 30, fig. 19 (comb.); Janse 1917: 81; Gaede 1927: 308, pl. 47; Pinhey 1975: 131, pl. 28, fig. 575, pl. 30, figs 575a–c.
Stenoglene obtusa (Walker, 1865): Kirby 1892: 809 (comb.); Forbes 1955: 95, 129; Vári *et al*. 2002: 150; Krüger 2020: 138; Takano & László 2022: 13, figs 50–52; Basquin 2024: 22 (partim).
Stenoglene obtusus (Walker, 1865): Dall'Asta & Poncin 1980: 482; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [South Africa] "Natal".
Name-bearing type material: holotype ♂ in NHMUK.

Stenoglene tristis Felder & Felder, 1874 (Fig. 10)
Stenoglene tristis Felder & Felder 1874: pl. 95, fig. 4; Aurivillius 1901: 30 (syn.).
Messata tristis (Felder & Felder, 1874): Moore 1883: 139, pl. 134, fig. 2 (comb.); Cotes & Swinhoe 1887: 210; Kirby 1892: 809.

Type locality: "Ceylon" [= Sri Lanka, *patria falsa* (probably South Africa)].
Name-bearing type material: holotype ♀ in NHMUK.

Phasicnecus evanescens Distant, 1903
Phasicnecus evanescens Distant 1903: 84, fig. 7; Janse 1917: 82; Gaede 1927: 308; Pinhey 1975: 131 (syn.).
Stenoglene evanescens (Distant, 1903): Forbes 1955: 129 (comb.).

Type locality: [South Africa] "Transvaal, Lydenburg District".
Name-bearing type material: holotype ♀ (and not a ♂ as stated in the original description) in NHMUK.

Phasicnecus subcroceus Distant, 1903
Phasicnecus subcroceus Distant 1903: 84, pl. 7, fig. 12; Janse 1917: 82; Gaede 1927: 308; Pinhey 1975: 131 (syn.).
Stenoglene subcroceus (Distant, 1903): Forbes 1955: 129 (comb.).

Type locality: [South Africa] "Transvaal, Pretoria".
Name-bearing type material: holotype ♂ in NHMUK.

***Stenoglene pira* Druce, 1896**
Stenoglene pira Druce 1896: 354; Forbes 1955: 129; Vári *et al*. 2002: 151; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.
Phasicnecus pira (Druce, 1896): Aurivillius 1901: 30 (comb.); Gaede 1927: 309.

Type locality: [Tanzania] "Dar-es-Salaam".
Name-bearing type material: holotype ♀ in NHMUK.

Phasicnecus aequidistans Strand, 1915 **syn. n.**
Phasicnecus aequidistans Strand 1915: 130; Gaede 1927: 308, pl. 47.
Stenoglene aequidistans (Strand, 1915): Forbes 1955: 129 (comb.).
Stenoglene obtusus auct. (nec Walker): Dall'Asta & Poncin 1980: 482 (syn.); Kitching & Rougerie *et al*. 2018: suppl. 1.
Stenoglene obtusa auct. (nec Walker): Vári *et al*. 2002: 151; Krüger 2020: 138.

Type locality: [Tanzania] "Daressalam".
Name-bearing type material: holotype ♂ in MfN.

Notes. Considered to be synonymous with *S. obtusa* by Dall'Asta & Poncin (1980) and later authors, examination of the holotype of *P. aequidistans* **syn. n.** (incidentally from the same type locality as *S. pira*) has confirmed that it is synonymous with *S. pira*.

***Stenoglene rosea* (Druce, 1886)** (Figs 5–6, 18)
Chrysopoloma rosea Druce 1886: 410, pl. 38, fig. 3; Janse 1917: 135.
Stenoglene rosea (Druce, 1886): Kirby 1892: 810 (comb.); Forbes 1955: 129; Takano & László 2022: 13, figs 55, 56; Basquin 2024: 22.
Phasicnecus roseus (Druce, 1886): Aurivillius 1901: 29 (comb.); Janse 1917: 81; Gaede 1927: 307, pl. 47.
Phasicnecus roseus roseus (Druce, 1886): Pinhey 1975: 131, pl. 30, fig. 576a.
Stenoglene roseus (Druce, 1886): Dall'Asta & Poncin 1980: 485; Vári *et al*. 2002: 151; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.

Type locality: [Mozambique] "Delagoa Bay" [= Maputo Bay].
Name-bearing type material: holotype ♂ in NHMUK.

Phasicnecus monteironis Rothschild, 1917
Phasicnecus monteironis Rothschild 1917b: 491; Gaede 1927: 307;
Stenoglene monteironis (Rothschild, 1917): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 485 (syn.).

Type locality: [Mozambique] "Delagoa Bay" [= Maputo Bay].
Name-bearing type material: holotype ♀ in NHMUK.

***Stenoglene similis* (Rothschild, 1917) stat. rev.** (Figs 7–8, 19)
Phasicnecus similis similis Rothschild 1917b: 491; Gaede 1927: 307, pl. 47.
Stenoglene similis (Rothschild, 1917): Forbes 1955: 95, 129 (comb.).
Phasicnecus roseus similis Rothschild, 1917: Pinhey 1975: 131, pl. 30, fig. 576c (stat.).
Stenoglene similis similis (Rothschild, 1917): Dall'Asta & Poncin 1980: 485.
Stenoglene roseus auct. (nec Druce): Vári *et al*. 2002: 151 (syn.); Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.

Type locality: [Malawi] "Lilongwi, Angoniland" [= Lilongwe].
Name-bearing type material: 1 ♂ and 2 ♀ syntypes in NHMUK.

Notes. Dall'Asta & Poncin (1980) correctly revived *S. similis* from synonymy with *S. rosea* (as *roseus*) based on the different uncus shape, but this was ignored by Varí *et al*. (2002), Kitching & Rougerie *et al*. (2018) and Kruger (2020) who maintained the former as a junior synonym of the latter. Although the habitus of the two species are similar, *S. similis* is larger and the diffuse claret marking on the outer edge of the forewing submarginal fascia is always present and larger. Moreover, a curved postmedial fascia is weakly visible in the male and often well-defined in the female of *S. similis* whereas it is always absent in *S. rosea*.

Phasicnecus similis flavidior Rothschild, 1917 **syn. rev.**
Phasicnecus similis flavidior Rothschild 1917b: 491; Gaede 1927: 308.
Stenoglene similis flavidior (Rothschild, 1917): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 485 (syn.).
Stenoglene roseus auct. (nec Druce): Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [Malawi] "Kasangazi, nr. Bandawe, Lake Nyassa".
Name-bearing type material: syntype ♀ in NHMUK (♂ could not be traced).

Notes. The correct synonymy of this taxon with *S. similis* by Dall'Asta & Poncin (1980) is herein revived.

Phasicnecus pulverulentus Joicey & Talbot, 1924 **syn. n.**
[*Phasicnecus pulverulentus*]: Talbot 1923: pl. 81, figs 13, 15.
Phasicnecus pulverulentus Joicey & Talbot 1924: 559; Gaede 1927: 308.
Phasicnecus pulverulentis [sic] Joicey & Talbot, 1924: Dall'Asta & Poncin 1980: 485.
Stenoglene pulverulentus (Joicey & Talbot, 1924): Forbes 1955: 129 (comb.).
Stenoglene roseus auct. (nec Druce): Dall'Asta & Poncin 1980: 485 (syn.); Vári *et al*. 2002: 151; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 139.

Type locality: [D.R. Congo] "Lufira River, near Likasi Copper Mine"; "East Luvua Valley, five days north-east of Lake Mwaru".
Name-bearing type material: holotype ♂ in NHMUK. The original description lists three specimens, one male type, one female type and another male. The male type is considered the holotype based on Talbot's handwritten label designating it as such, as well as the fact that Joicey and Talbot were in the practice of assigning holotypes and allotypes when both sexes were to hand for a description. With regard to the third specimen, only a single female (not male) specimen matching the East Luvua Valley collecting data could be located and this specimen is considered a paratype.

Notes. Considered to be synonymous with *S. rosea* by Dall'Asta & Poncin (1980) and later authors, examination of the holotype has revealed *Phasicnecus pulverulentus* **syn. n.** to be conspecific with *S. similis* instead.

Description of new species

***Stenoglene lucindae* sp. n.** (Figs 11–12, 21)
https://www.zoobank.org/urn:lsid:zoobank.org:act:A37CA33A-7DB3-4AE2-9E9C-A5C38913F346

Holotype ♂ (Fig. 11):
"TANZANIE : Kagera | Region,, BIHARAMULO, | Old Boma, 26-X-2008 | (Ph. DARGE) || Ex. Coll. Ph. Darge | ANHRT:2019.20 || ANHRTUK | 00276391" (ANHRT).

Paratypes (3 ♂♂, 4 ♀♀):
TANZANIA: Geita Forest, Samina Village, 1294 m, 02°53.474′S, 32°08.075′E, 20.x.2011, leg. P. Darge (3 ♂♂, 1 ♀ ANHRT); Tabora, Ounyanyembé, leg. Père C. Hauttecoeur (2 ♀♀ NHMUK); Unjamwesi, 27.iv.1892, leg. Baron L. Fischer von Nagy-Szalatya (1 ♀ NHMW).

Notes. The single female specimen from "Unjamwesi" was collected by Baron Ludwig Fischer on his journey from Tabora to Lake Victoria in 1892, where he died of fever soon after his arrival at Nyegesi on the southern shore (Weidmann 1894). The specimens collected by him together with his diary were brought back by the other members of the expedition and an annotated map was published posthumously in volume 5 of *Mittheilungen von Forschungsreisenden und Gelehrten aus den deutschen Schutzgebieten* (1892). Cross-referencing the date of capture with the map, the specimen was collected in a region to the south-west of Old Shinyanga around 03°37′S, 33°19′E.

Description. Forewing lengths: male: 25–26 mm, female: 29–30 mm.

Male (Fig. 11). Upperside. Ground colour of body and wings golden yellow. Antennae brown, bipectinate. Forewing elongate triangular, outer margin curved. Postmedial and submarginal fasciae dark brown, curved inwards at vein R_3, the latter indicated by spots between the veins. Subterminal fascia dark brown, oblique, gently arcuate from apex, disappearing towards tornus. Area beyond subterminal fascia heavily irrorated with reddish-brown scales. Fringe yellowish-brown. Hindwing rounded, paler towards costa. Anal margin with dark brown spots almost conjoined, the spot nearest to tornus the largest. Fringe yellow. Underside. Ground colour of body rich orangey-yellow. Wings uniform golden-yellow with only forewing postmedial and submarginal fasciae weakly outlined. Male genitalia (Fig. 21). Uncus with medial structure, terminating in two pairs of short, pointed, heavily-sclerotised projections. Tegumen broad. Gnathos ring-shaped, thread-like and widest laterally. Mesal protrusions separated, trapezoidal, densely covered in setae. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section subrectangular, rounded at apex, with

long blade-like dorso-distal projection (as long as costal margin) pointed dorsad, and smaller projection positioned ventrad. Saccular section broad, triangular, tapering to long, robust, blade-like distal process, truncate at apex, three times as wide as distal costal process, and as long as sacculus; sacculus with triangular fold at apex, at base of distal process. Juxta semi-cylindrical, trapezoidal with two triangular processes dorsally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded and curved dorsad, carina heavily sclerotised and tapered; vesica with small patch of scobination.

Female (Fig. 12). Similar to male but antennae with shorter rami, forewings more elongate and more rounded outer margin. Forewing with vinaceous hue throughout; fasciae indistinct.

Diagnosis. This and the subsequent new species, *S. sarahae* **sp. n.** and *S. susanae* **sp. n.** are closely allied to *S. obtusa* but in the last, both sexes are smaller, the forewings are more elongate and the apex is pointed. In the males, the forewing ground colour of *S. obtusa* is more beige, the postmedial fascia is boldly marked, the submarginal fascia is weakly expressed and the obtuse subterminal fascia is well marked to the tornus where it is indicated by dark brown spots. The habitus of *S. lucindae* **sp. n.** is most similar to *S. sarahae* and *S. susanae*, but the ground colour is paler and the forewing more elongate. In the females, the markings are better defined in *S. obtusa*, while compared to *S. sarahae* and *S. susanae*, *S. lucindae* is darker. Since these species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance. In the male genitalia, *S. lucindae* is most similar to *S. obtusa* and *S. susanae* sharing with them the uncus that terminates in two pairs of pointed projections. However, *S. lucindae* differs from *S. obtusa* in the medial structure of the uncus being broader and both the inner and outer projections being nearly the same length (the outer is twice as long as the inner projection in *S. obtusa*), the smooth costal margin of the valve (clearly sinuate and irregular in *S. obtusa*) and a slightly broader distal saccular process. In *S. susanae*, the medial structure of the uncus is wider, the projections are stouter, and the outer projection is twice as long as the inner. Additionally, the distal process of the valve costa is half as long and much more robust in *S. susanae* compared to *S. lucindae*.

Molecular characterisation. The new species has been assigned the Barcode Index Numbers (BIN) BOLD:AFG4210 and BOLD:AFG3494. Intraspecific pairwise distances (PWD) were 0.0–2.4% (n=4) and diverged from its sibling *S. susanae* **sp. n.** by 5.0–5.9% (n=2).

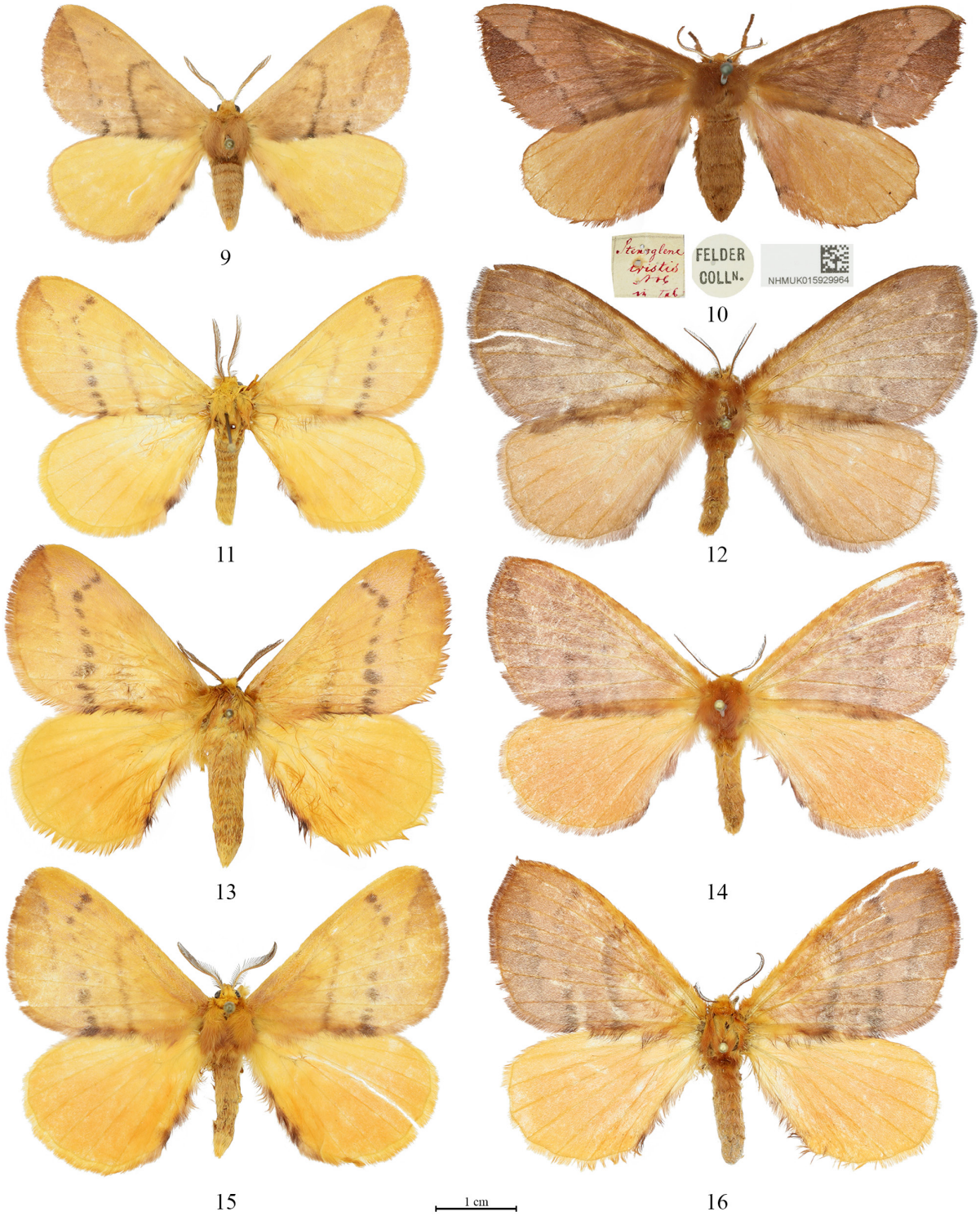
Derivatio nominis. It is with great pleasure that this new species is dedicated to Lucinda Sharp-Smith, a Trustee of ANHRT, who like her parents shares a great love of natural history and conservation, and in gratitude for her on-going support of the institution and its entomological activities. The matronym is a noun in the genitive case.

Distribution. This new species is known only from woodlands and forests of the Lake Victoria basin in Tanzania. It may be found more widely in similar habitats in Uganda and Tanzania.

***Stenoglene sarahae* sp. n.** (Figs 13–14, 22)
https://www.zoobank.org/urn:lsid:zoobank.org:act:7B322295-923D-4B02-9693-2E08FAF14D06

Holotype ♂ (Fig. 13):
"ZAMBIA 1437m | Kapishya Hot Springs, | Shiwa N'gandu Estate | 11°10'13"S, 31°36'00"E | 22.xi-8.xii.2015 Light Trap | Smith, R., Takano, H., | Oram, D. Leg | ANHRT:2017.15 || ANHRTUK | 00160315" (ANHRT).

Paratypes (24 ♂♂, 2 ♀♀):
ZAMBIA: same data as holotype (1 ♂ ANHRT); Chisimbe Falls, 1320 m, 10°06′31″S, 30°55′02″E, 27–29.iv.2013, leg. R. Smith, H. Takano & D. Oram (1 ♂ ANHRT); same locality as for preceding but 26.xi.2012, leg. R. Smith & H. Takano (1 ♂ ANHRT); Kalungu, 1280 m, 09°40′52″S, 32°42′50″, 22–24.xi.2016, leg. R. Smith, H. Takano & D. Oram (15 ♂♂, 2 ♀♀ ANHRT); same locality as for preceding but 05–08.iii.2017, leg. D. Oram, W. Miles & L. Smith (1 ♂ ANHRT); same locality as for preceding but 30.iv–01.v.2019, leg. V. Dérozier, G. László & W. Miles (1 ♂ ANHRT); Lukulu River, Lavushi Manda National Park, 1285 m, 12°15′05″S, 30°53′43″E, 27–29.xi.2012, leg. R. Smith & H. Takano (1 ♂ ANHRT); Mutinondo Wilderness Area, 1460 m, 12°27′06″S, 31°17′30″E, 26–29.xi.2016, leg. R. Smith, H. Takano & D. Oram (1 ♂ ANHRT); Ntumbachushi Falls, 1166 m, 09°51′12″S, 28°56′40″E, 03–04.xi.2014, leg. R. Smith, H. Takano & D. Oram (2 ♂♂ ANHRT).



Figures 9–16. *Stenoglene* Felder & Felder, 1874 species, habitus (in ANHRT unless otherwise stated). 9. *S. obtusa* (Walker, 1865), ♂, Mozambique, Maputo Special Reserve. 10. *Id*., holotype ♀ of *S. tristis* Felder & Felder, 1874 (NHMUK). 11. *S. lucindae* **sp. n.**, holotype ♂. 12. *Id*., paratype ♀. 13. *S. sarahae* **sp. n.**, holotype ♂. 14. *Id*., paratype ♀. 15. *S. susanae* **sp. n.**, holotype ♂. 16. *Id*., paratype ♀.

Description. Forewing lengths: male: 25–28 mm, female: 30 mm.

Male (Fig. 13). Upperside. Ground colour of head, abdomen and hindwing golden yellow, thorax and forewing deep orange-yellow. Antennae brown, bipectinate. Forewing elongate triangular, outer margin curved. Postmedial fascia dark brown, curved inwards at vein R_3, indistinct. Submarginal fascia dark brown, curved inwards at vein R_3 and displaced towards anal margin, indicated by spots between

the veins. Subterminal fascia dark brown, oblique, gently arcuate from apex, disappearing towards tornus. Area beyond subterminal fascia heavily irrorated with rich reddish-brown scales. Fringe brown. Hindwing rounded, paler towards costa. Anal margin with dark brown spots almost conjoined. Fringe yellow. Underside. Ground colour of body rich orangey-yellow. Wings uniform golden-yellow with only forewing postmedial and submarginal fasciae weakly outlined. Male genitalia (Fig. 22). Uncus with medial structure, terminating in two apically dentate and truncate, heavily-sclerotised projections. Tegumen broad. Gnathos ring-shaped, thin and thread-like. Mesal protrusions separated, trapezoidal, densely covered in setae. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section subrectangular, margin gently sinuate, rounded at apex, with long blade-like dorso-distal projection (two-thirds the length of costal margin) pointed dorsad, and smaller, triangular projection positioned ventrad. Saccular section broad, triangular, tapering to long, blade-like distal process, rounded at apex, twice as wide as distal costal process, and as long as sacculus; sacculus with triangular fold at apex, at base of distal process. Juxta semi-cylindrical, trapezoidal. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded and curved dorsad, carina heavily sclerotised and tapered; vesica with small patch of scobination.

Female (Fig. 14). Similar to male but antennae with shorter rami, forewings more elongate and with more rounded outer margin. Forewing with vinaceous hue throughout; fasciae indistinct.

Diagnosis. See under *S. lucindae* **sp. n.** for diagnostic comparisons. *Stenoglene sarahae* **sp. n.** is most similar to *S. susanae* **sp. n.** and is almost indistinguishable based on habitus but the male genitalia are highly diagnostic. The medial structure of the uncus in *S. sarahae* terminates in a pair of apically dentate and truncate projections, while in *S. susanae*, the uncus possesses two pairs of pointed projections.

Molecular characterisation. The new species has been assigned the BIN BOLD:AAO9401. Intraspecific PWDs were 0.0–0.6% (n=4) and diverged from its sibling *S. labda* by 2.0–2.7% (n=5).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Sarah Sharp-Smith FRES, a lifelong advocate of entomology, and who together with her husband Richard has travelled widely throughout Africa including Zambia from where the new species originates. The matronym is a noun in the genitive case.

Distribution. The new species is distributed in north-eastern Zambia.

***Stenoglene susanae* sp. n.** (Figs 15–16, 23)
https://www.zoobank.org/urn:lsid:zoobank.org:act:E412794D-5B44-477F-BC84-3BBDA7D15C7F

Holotype ♂ (Fig. 15):
"ZAMBIA 1440m | Kalene Hill | (Miombo woodland) | 11°11'11"S, 24°12'5"E | 27.xi-3.xii.2020 MV Light | Trap. Chizuwa, D., | Choongo, W. Leg. | ANHRT:2022.4 || ANHRTUK | 00270027" (ANHRT).

Paratypes (2 ♂♂, 1 ♀):
ZAMBIA: Jiwundu Swamp, 1340 m, 11°51′54″S, 25°33′20″E, 29.x–04.xi.2018, leg. M. Aristophanous, V. Dérozier, G. László & D. Oram (1 ♀ ANHRT); Kambishi, Jiwundu, 1340 m, 11°54′39″S, 25°29′05″E, 18–19.x.2014, leg. R. Smith, H. Takano & D. Oram (1 ♂ ANHRT); Ndanda, 1090 m, 15°04′44″S, 23°45′59″E, 10–11.xi.2013, leg. R. Smith, H. Takano & D. Oram (1 ♂ ANHRT).

Description. Forewing lengths: male: 23–26 mm, female: 30 mm.

Male (Fig. 15). Upperside. Ground colour of head, abdomen and hindwing golden yellow, thorax and forewing deep orange-yellow. Antennae brown, bipectinate. Forewing elongate triangular, outer margin curved. Postmedial and submarginal fasciae dark brown, curved inwards at vein R_3, the latter indicated by spots between the veins. Subterminal fascia dark brown, oblique, gently arcuate from apex, disappearing towards tornus. Area beyond subterminal fascia heavily irrorated with rich reddish-brown scales. Fringe brown. Hindwing rounded, paler towards costa. Anal margin with three dark brown spots almost conjoined. Fringe yellow. Underside. Ground colour of body rich orangey-yellow. Wings uniform golden-yellow with only forewing postmedial and submarginal fasciae weakly outlined. Male genitalia (Fig. 23). Uncus with medial structure, terminating in two pairs of short, stout, pointed, heavily-sclerotised projections, the outer projection twice as long as the inner. Tegumen broad. Gnathos ring-

shaped, thread-like and widest ventrally. Mesal protrusions separated, triangular, densely covered in setae. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section subrectangular, margin gently sinuate, rounded at apex, with long blade-like dorso-distal projection (half the length of costal margin) pointed dorsad, and smaller, triangular projection positioned ventrad. Saccular section broad, triangular, tapering to long, blade-like distal process, truncate at apex, twice as wide as distal costal process, and as long as sacculus; sacculus with narrow triangular fold at apex, at base of distal process. Juxta semi-cylindrical, trapezoidal with two rounded processes dorsally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded and curved dorsad, carina heavily sclerotised and tapered; vesica with small patch of scobination.

Female (Fig. 16). Similar to male but antennae with shorter rami, forewings broader with more rounded outer margin. Forewing with vinaceous hue throughout; fasciae broader and submarginal fascia with spots conjoined.

Diagnosis. See under *S. lucindae* **sp. n.** and *S. sarahae* **sp. n.** for comparative diagnoses.

Molecular characterisation. The new species has been assigned the BIN BOLD:AFI0043. The intraspecific PWD was 0.8% (n=2) and diverged from *S. lucindae* **sp. n.** by 5.0–5.9% (n=4).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Susan Smith, a Trustee of ANHRT and a passionate conservationist, in gratitude for her on-going support of the institution and its entomological activities. The matronym is a noun in the genitive case.

Distribution. The new species is known only from north-western Zambia.

***Cryptojana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:09DC16DA-E90E-4812-A66D-942EBBB3036D

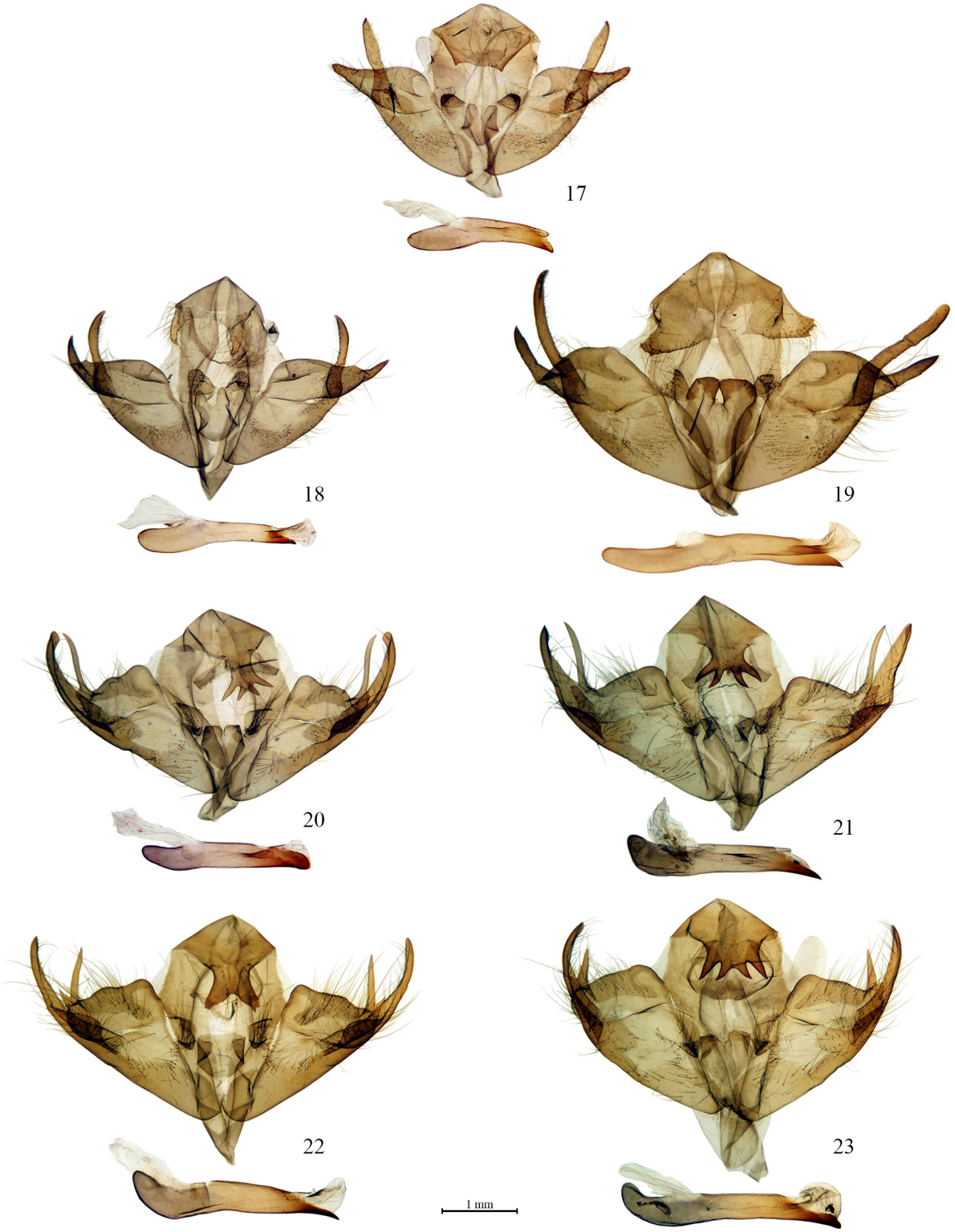
Type species: *Phasicnecus plagiatus* Aurivillius, 1911.

Description. Adults. Medium-sized moths. Antenna bipectinate, rami shorter in females. Ground colour ranging from grey and greenish beige through to dark brown. Forewing elongate with curved outer margin; $R_4 + R_5$ stalked. Discal marking present, sometimes weakly-expressed. Postmedial fascia curved and irregularly sinuate or crenulate, or straight and oblique; submarginal fascia strongly sinuate forming dark ovoid markings towards anal margin in some species. Hindwing ovoid, outer margin evenly arcuate to angled or produced tornus. Postmedial fascia arcuate, absent in some species; submarginal fascia arcuate, interrupted and indicated by dots in some species. Anal margin with dark markings at tornus and at termination points of postmedial/submarginal fasciae. Underside paler with markings often limited to postmedial and submarginal fasciae or indications thereof. Females (where known) similar to male but generally larger, forewing with produced apex and hindwing with produced, angled tornus. Male genitalia. Uncus produced into elongate medial structure bifurcate at apex or a pair of triangular medio-lateral processes and pair of hooked lateral projections. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, ovoid or trapezoid, densely covered in setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate, ovoid or triangular, margin strongly lobed giving the appearance of antlers. Saccular section triangular, half as wide as costal section, tapering to long distal process at least as long as sacculus. Juxta semi-cylindrical, inverse-triangular. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus slender, coecum rounded, slightly curved at entrance of ductus ejaculatoris, carina heavily sclerotised and tapered; vesica with a small patch of scobination.

Diagnosis. This genus is formed of a compact group of species with sombre colouration and all displaying a distinctively crenulate submarginal fascia of the forewing. It is reminiscent of *Lokiana* **gen. n.** and *Neojana* **gen. n.** but can easily be separated based on habitus and genital morphology – see diagnoses under these two genera for further comparisons. Some species of *Microjana* **gen. n.** have similar markings but all members are at least half the size of *Cryptojana* **gen. n.** species, with broader nearly transparent wings. The male genitalia of *Cryptojana* are highly distinctive, notably the uncus produced into either an elongate medial structure bifurcate at its apex or a pair of triangular medio-lateral processes, and a pair of hooked lateral processes, and the costal section of the valve with its lobed margin somewhat reminiscent of palmate cervid antlers.

Derivatio nominis. The compound genus name is derived from the Ancient Greek for 'concealed' (κρυπτός), in reference to the cryptic phenotype of its members, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Members of this genus are distributed in forest and forest-savanna mosaics, with the majority being found in the Guineo-Congolian rainforests.



Figures 17–23. *Stenoglene* Felder & Felder, 1874 species, ♂ genitalia. 17. *S. hilaris* Felder & Felder, 1874. 18. *S. rosea* (Druce, 1886). 19. *S. similis* (Rothschild, 1917). 20. *S. obtusa* (Walker, 1865). 21. *S. lucindae* **sp. n.**, holotype. 22. *S. sarahae* **sp. n.**, holotype. 23. *S. susanae* **sp. n.**, holotype.

Species content: 11 species are assigned to this genus, one of which is a new species.

***Cryptojana cinerea* (Holland, 1893) comb. n.**
Epijana cinerea Holland 1893: 515; Aurivillius 1901: 28; Gaede 1927: 306.
Stenoglene cinerea (Holland, 1893): Forbes 1955: 95, 129 (comb.); Dall'Asta & Poncin 1980: 481; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 21; Basquin & Faravel 2025: pl. 31.

Type locality: [Gabon] "Valley of the Ogové River".
Name-bearing type material: holotype ♀ in CMNH (not examined).

***Cryptojana clucki* (Bouyer, 2012) comb. n.**
Stenoglene clucki Bouyer 2012: 6, pl. 3, fig. 20; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138; Basquin 2024: 21.

Type locality: "South Africa, Maputoland, NE Ndumo, Tembe Elephant park".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Cryptojana decellei* (Dall'Asta & Poncin, 1980) comb. n.** (Figs 29, 35)
Stenoglene decellei decellei Dall'Asta & Poncin 1980: 483, figs 25–26, photo 2, fig. 18; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20; Basquin & Faravel 2025: pl. 24.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

Stenoglene decellei flava Dall'Asta & Poncin, 1980 **syn. n.**
Stenoglene decellei flava Dall'Asta & Poncin 1980: 484, photo 2, fig. 20; Vári *et al*. 2002: 150 (misidentification); Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138 (misidentification).

Type locality: [D.R. Congo] "Eala".
Name-bearing type material: holotype ♂ in RMCA.
Notes. Described as a 'pale' subspecies of *C. decellei* but with near identical genital morphology, *Stenoglene decellei flava* **syn. n.** is herein synonymised with the former.

***Cryptojana grandiplaga* (Holland, 1920) stat. rev., comb. n.** (Figs 25, 32)
Phasicnecus grandiplaga Holland 1920: 304, pl. 14, fig. 14.
Phasicnecus maculifera plagiatus auct. (nec Aurivillius): Gaede 1927: 307 (syn.).
Stenoglene maculifera plagiatus auct. (nec Aurivillius): Forbes 1955: 129 (comb.).
Stenoglene plagiatus auct. (nec Aurivillius): Dall'Asta & Poncin 1980: 481 (syn.); Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20; Basquin & Faravel 2025: pl. 30.

Type locality: [D.R. Congo] "Medje".
Name-bearing type material: holotype ♂ in AMNH (photograph examined).
Notes. Based on barcode divergences and differing genital configurations, *C. grandiplaga* is revived from synonymy with *C. plagiata*. Further work is needed on this group as publicly available barcode data suggests a complex of cryptic species.

***Cryptojana insignifica* (Rothschild, 1917) comb. n.** (Fig. 30)
Haplojana [sic] *insignifica* Rothschild 1917b: 489.
Hoplojana insignifica Rothschild, 1917: Gaede 1927: 304.

Rhodopteriana insignifica (Rothschild, 1917): Bouyer 2011: 213 (comb.); Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] "Yakusu, Upper Congo".
Name-bearing type material: holotype ♀ in NHMUK.

Notes. For the Eupterotidae chapter in Seitz' *Die Grossschmetterlinge der Erde*, Gaede (1927) studied the type specimen of *Hoplojana insignifica* adding a determination label of "*Phasicnecus* (*Epijana*) *maculifera* Strand" to it, but treated the taxon within *Hoplojana* (perhaps out of reverence to Rothschild), only alluding in the text that it was the female of *P. maculifera*. Based on publicly available barcodes of specimens from eastern D.R. Congo, it appears this taxon is not conspecific with *C. maculifera* or *C. cinerea*, although further investigations are required.

***Cryptojana maculifera* (Strand, 1912) comb. n.**
Epijana maculifera Strand 1912: 148.
Phasicnecus maculifera (Strand, 1912): Gaede 1927: 307, pl. 47 (comb.).
Stenoglene cinerea auct. (nec Holland): Forbes 1955: 95, 129 (comb. & syn.); Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20.
Stenoglene maculifera (Strand, 1912): Basquin & Faravel 2025: pl. 32 (stat.).

Type locality: [Equatorial Guinea] "Nkolentangan".
Name-bearing type material: 1 ♂ and 1 ♀ syntypes in MfN.

Notes. Based on barcode divergences, there are seemingly two near-identical species with the oblique forewing postmedial fascia found in Gabon. Basquin & Faravel (2025) resurrected *C. maculifera* without providing any justifications aside from the barcode differences, but considering *C. cinerea* was described from a female, further research is required to establish whether these names represent two valid or synonymous taxa, and in the case of the latter, a new species will need to be described.

***Cryptojana noellae* (Bouyer, 2012) comb. n.**
Stenoglene noellae Bouyer 2012: 7, pl. 2, fig. 14; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20; Basquin & Faravel 2025: pl. 33.

Type locality: "Cameroun, Obout".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Cryptojana plagiata* (Aurivillius, 1911) comb. n.** (Figs 24, 31)
Phasicnecus plagiatus Aurivillius 1911: 164.
Phasicnecus maculifera plagiatus (Aurivillius, 1911): Gaede 1927: 307 (stat.).
Stenoglene maculifera plagiata (Aurivillius, 1911): Forbes 1955: 129 (comb.).
Stenoglene plagiatus (Aurivillius, 1911): Dall'Asta & Poncin 1980: 481 (stat.), photo 2, fig. 22; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [Ghana] "Coomassie" [= Kumasi].
Name-bearing type material: holotype ♂ in NHMUK.

***Cryptojana preussi* (Aurivillius, 1893) comb. n.** (Fig. 26)
Jana preussi Aurivillius 1893: 209.
Phasicnecus preussi (Aurivillius, 1893): Aurivillius 1901: 29, pl. 4, fig. 3 (comb.); Gaede 1927: 307, pl. 47.
Stenoglene preussi (Aurivillius, 1893): Forbes 1955: 95, 129 (comb.); Dall'Asta & Poncin 1980: 480; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 21 (partim); Basquin & Faravel 2025: pl. 29 (partim).

Type locality: “Camerun interior”.
Name-bearing type material: holotype ♂ in MfN.

***Cryptojana shabae* (Dall’Asta & Poncin, 1980) comb. n.** (Figs 28, 33)
Stenoglene shabae Dall’Asta & Poncin 1980: 481, figs 21–22, photo 2, figs 21, 23; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] “Elisabethville” [=Lubumbashi].
Name-bearing type material: holotype ♂ in RMCA.

Description of new species

***Cryptojana volynkini* sp. n.** (Figs 27, 34)
https://www.zoobank.org/urn:lsid:zoobank.org:act:894FFC5B-945F-44AD-BF93-854C0003EA50

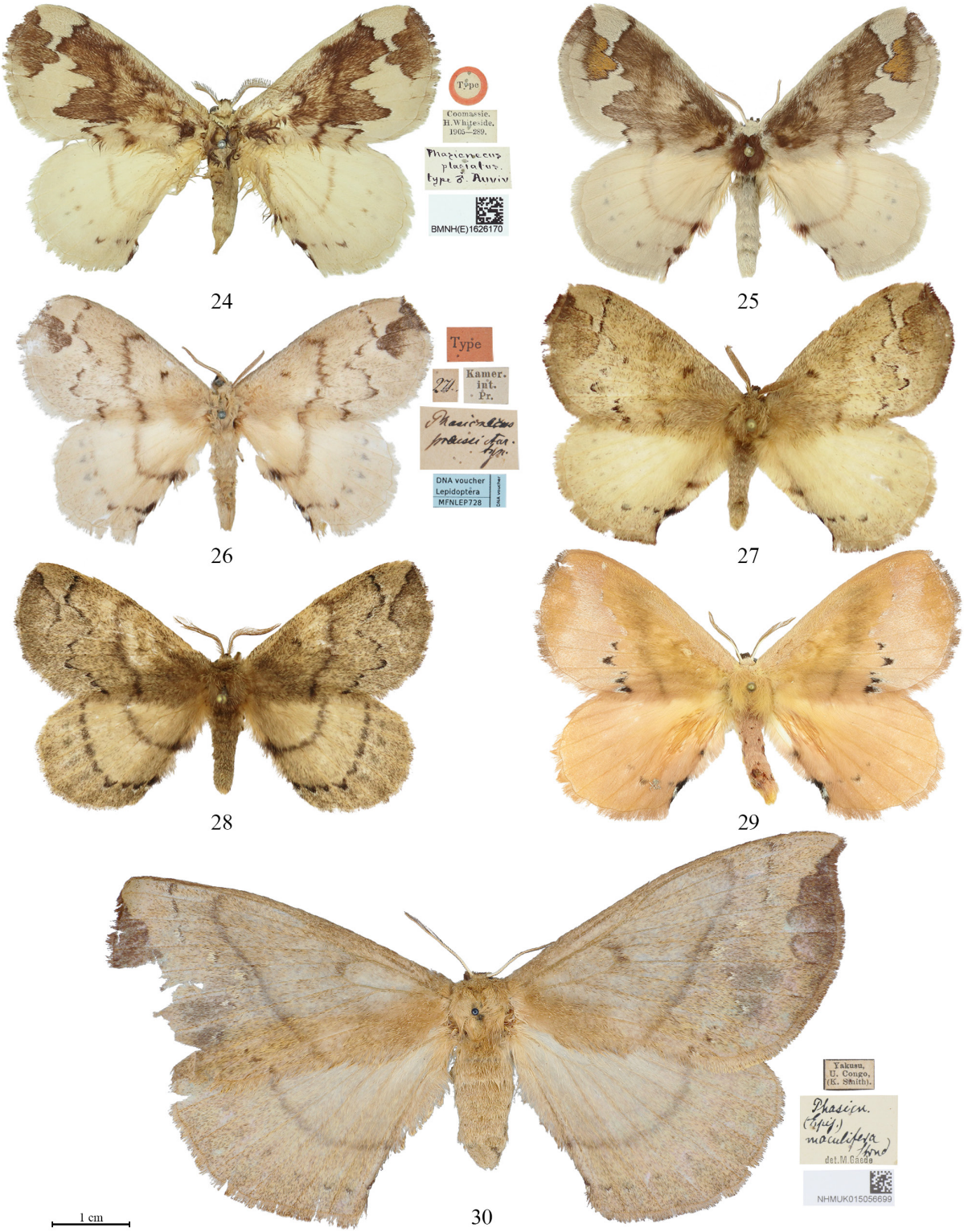
Holotype ♂ (Fig. 27):
“TANZANIE :Kagera Re- | gion, Minziro Forest, Kere | Hill, 1176 m., 01°06.087′S., | 031°31.168′ E., | 10-X-2012 (Ph. Darge) || Ex. Coll. Ph. Darge | ANHRT:2019.20 || ANHRTUK | 00276410” (ANHRT).

Description. Forewing length: 27 mm.

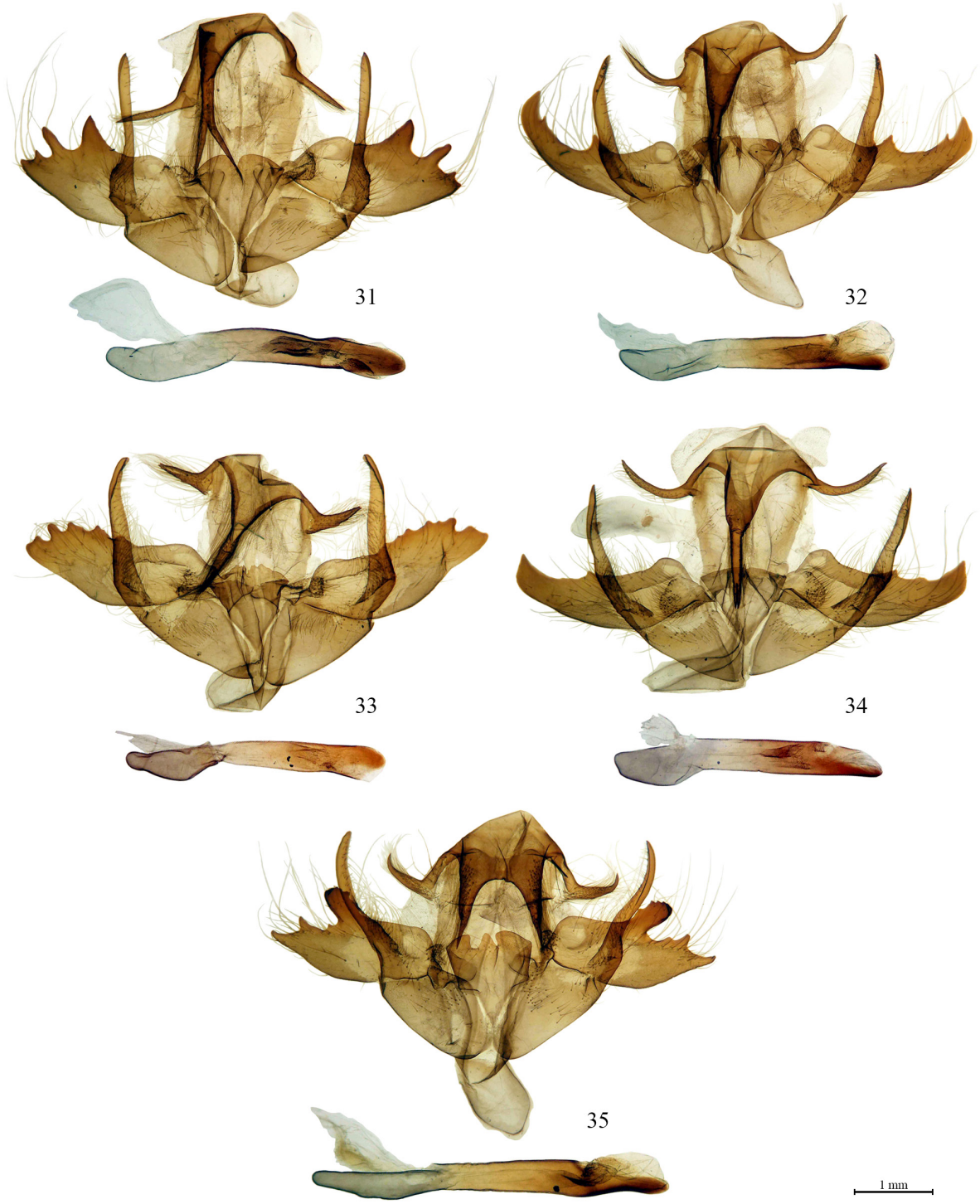
Male (Fig. 27). Upperside. Ground colour of body and forewing greyish-brown, hindwing beige. Antennae brown, bipectinate, with beige scaling along its entire length. Forewing elongate triangular, costal margin gently arcuate, outer margin arcuate to rounded tornus. Base heavily irrorated with brown scaling. Discal spot dark brown, small. Postmedial fascia brown, almost straight, crenulate, weakly defined, perpendicular to anal margin. Submarginal fascia dark brown edged with silvery-grey, irregularly sinuate, strongly displaced proximally in space M_2 and between veins Cu_2 and anal margin. Subapical marking brown, bordered on its inner edge by dark brown, crenulate subterminal fascia which terminates abruptly caudad of marking. Termen irrorated with dark brown scaling. Fringe dark brown. Hindwing rounded, sharply angled at tornus. Terminal region irrorated with dark brown scaling, heavy towards tornus. Postmedial fascia absent except for marking along anal margin. Submarginal fascia dark brown, curved, indicated by spots and weakly expressed except towards anal margin where it terminates in large dark brown spot. Tornus with dark brown spot along anal margin. Underside. Ground colour uniform greyish-brown. Wing markings reduced to dark brown submarginal fascia indicated by spots. Forewing subapical marking very weakly marked. Male genitalia (Fig. 34). Uncus produced into elongate medial structure tapering towards bifurcate apex, and pair of hooked, apically pointed lateral projections, each just over half the length of medial structure. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, ovoid, densely covered in setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate, triangular, truncate at apex and gently curved dorsad; margin weakly sinuate with small projections (non-symmetrical between left and right valves), terminating at a dorso-distal point. Saccular section triangular, half as wide as costal section, tapering to long blade-like distal process, pointed at apex, nearly twice as long as sacculus. Juxta semi-cylindrical, inverse-triangular, dorsal margin with medial emargination. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus slender, slightly curved ventrad at entrance of ductus ejaculatoris, carina heavily sclerotised and tapered; vesica with a small patch of scobination.

Female. Unknown.

Diagnosis. The new species is most similar in appearance to the *C. preussi* group to include *C. plagiata* and *C. grandiplaga* but in these three taxa, the postmedial fascia of both wings is well-defined and curved, most noticeable on the hindwing. The male genitalia of the new species is most similar to *C. grandiplaga* (N.B. the genitalia of *C. preussi* has not been examined) although in the latter, the costal section of the valve is more slender and curved with larger projections along the margin.



Figures 24–30. *Cryptojana* **gen. n.** species, habitus (in ANHRT unless otherwise stated). 24. *C. plagiata* (Aurivillius, 1911), holotype ♂ of *Phasicnecus plagiatus* Aurivillius (NHMUK). 25. *C. grandiplaga* (Holland, 1920), ♂, Republic of Congo, Nouabalé-Ndoki NP. 26. *C. preussi* (Aurivillius, 1893), holotype ♂ of *Phasicnecus preussi* Aurivillius (MfN). 27. *C. volynkini* **sp. n.**, holotype ♂. 28. *C. shabae* (Dall'Asta & Poncin, 1980), ♂, Zambia, Choma. 29. *C. decellei* (Dall'Asta & Poncin, 1980), ♂, Republic of Congo, Odzala-Kokoua NP. 30. *C. insignifica* (Rothschild, 1917), holotype ♀ of *Hoplojana insignifica* Rothschild (NHMUK).



Figures 31–35. *Cryptojana* **gen. n.** species, ♂ genitalia. 31. *C. plagiata* (Aurivillius, 1911). 32. *C. grandiplaga* (Holland, 1920). 33. *C. shabae* (Dall'Asta & Poncin, 1980). 34. *C. volynkini* **sp. n.**, holotype. 35. *C. decellei* (Dall'Asta & Poncin, 1980).

Molecular characterisation. The new species has been assigned the BIN BOLD:AAE1933 and the holotype diverged from *C. preussi* by 5.6% (n=1), *C. plagiata* by 7.3–7.5% (n=2) and *C. grandiplaga* by 7.7% (n=1).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Anton Volynkin, ANHRT, a specialist of the Old World Arctiinae, for helpful discussions in this and many other projects over the years, and for his outstanding contributions to the Lithosiini fauna of Africa. The patronym is a noun in the genitive case.

Distribution. Known only from the type locality on the western shores of Lake Victoria in Tanzania.

***Microjana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:297F2D6D-C317-46E6-BF9F-5374C8EC4F3B

Type species: *Chrysopoloma citrina* Druce, 1886.

Description. Adults. Small moths. Antenna bipectinate, rami shorter in females. Ground colour ranging from creamy-white through yellow to pink and ferruginous-brown; wings diaphanous with pavonine iridescence in most species. Forewing broad triangular with rounded apex, outer margin and tornus; R_4 + R_5 stalked. Hindwing outer margin arcuate. Postmedial and submarginal fasciae irregular, sinuate and often indicated by dots, the former absent in some species while the latter can be oblique; outer edge of submarginal fascia with dark ovoid markings towards anal margin most noticeable on forewing. Subterminal fascia indicated by dashes or dots following outer margin when present. Dark subapical spot or spots often present on forewing. Underside uniform, often paler, with bolder markings from upperside showing through. Male genitalia. Uncus medially reduced with pair of single or bifid lateral processes. Tegumen broad. Gnathos reduced, membranous. Mesal protrusions separated, ovoid or triangular, heavily reduced in some species. Valve elongate triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender, tapering towards bifurcate apex. Saccular section elongate triangular, broader than costal section, tapering to bifurcate or hooked distal process. Juxta semi-cylindrical, trapezoidal, sometimes with bilobed projection dorsally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus curved and constricted medially, tapering towards carina; coecum rounded; vesica globular with a small patch of scobination.

Diagnosis. This genus comprises a highly distinctive group of species, easily distinguishable from all other genera based on their small size, their delicate, almost diaphanous wings and distinctive markings. In the male genitalia, despite the extreme structural plasticity in the processes of the uncus (see additional figures in Dall'Asta & Poncin (1980) and Bouyer (2012)), the configuration of the valves, especially the costal section and the medially curved and constricted phallus is highly diagnostic.

Derivatio nominis. The compound genus name is derived from the Ancient Greek for 'small' (μικρός) in reference to the stature of its members, some of which are the smallest Eupterotidae in the world, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. With the exception of one species found in a forest mosaic, members of this genus are distributed throughout the Guineo-Congolian rainforests.

Notes. There is a great deal of intraspecific variability in the ground colour of the wings (see and dissections are required to confirm identification. There are likely to be a number of synonymies and a full revision of this group is required.

Species content: 25 species are assigned to this genus, three of which are new species.

***Microjana annae* (Giusti & László, 2025) comb. n.**
Stenoglene annae Giusti & László, *In*: László *et al*. 2025: 20, figs 10–11, 19–20.

Type locality: "Johan-Albrechts Höhe Station - Kamerun".
Name-bearing type material: holotype ♂ in NHMUK.

***Microjana brunneofasciata* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene brunneofasciata Dall'Asta & Poncin 1980: 471, figs 5–6, photo 1, fig. 8; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 23.

Type locality: [D.R. Congo] "Uele, Paulis".

Name-bearing type material: holotype ♂ in RMCA.

***Microjana citrina* (Druce, 1886) comb. n.** (Figs 36–37, 46)
Chrysopoloma citrina Druce 1886: 410, pl. 38, fig. 4.
Stenoglene citrina (Druce, 1886): Kirby 1892: 810 (comb.); Forbes 1955: 95, 129; Dall'Asta & Poncin 1980: 469; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 22.
Phasicnecus citrinus (Druce, 1886): Aurivillius 1901: 30 (comb.); Gaede 1927: 308, pl. 47.

Type locality: [Nigeria] "Old Calabar".
Name-bearing type material: holotype ♂ in NHMUK.

***Microjana citrinoides* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene citrinoides Dall'Asta & Poncin 1980: 471, figs 17–18, photo 1, fig. 5; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

***Microjana dallastai* (Bouyer, 2012) comb. n.**
Stenoglene dallastai Bouyer 2012: 11, pl. 1, fig. 4, pl. 2, fig. 11; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Guinée Conakry, forêt classée de Diéké".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Microjana dehanica* (Strand, 1911) comb. n.**
Phasicnecus dehanicus Strand 1911: 17; Gaede 1927: 307, pl. 41.
Stenoglene dehanica (Strand, 1911): Forbes 1955: 129 (comb.).
Stenoglene dehanicus (Strand, 1911): Dall'Asta & Poncin 1980: 477; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 18.

Type locality: "Dehane, Süd-Kamerun".
Name-bearing type material: holotype ♂ in MfN.

***Microjana dufranei* (Bouyer, 2012) comb. n.**
Stenoglene dufranei Bouyer 2012: 11, pl. 1, fig. 6, pl. 2, fig. 13; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "R.D. Congo, Kabunga, territoire Masisi, Kivu".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Microjana fontainei* (Dall'Asta & Poncin, 1980) comb. n.** (Figs 43–45, 50)
Stenoglene fontainei Dall'Asta & Poncin 1980: 469, figs 11–12, photo 1, fig. 14; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 24.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

***Microjana fouassini* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene fouassini Dall'Asta & Poncin 1980: 473, figs 15–16, photo 1, fig. 12; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 18.

Type locality: [D.R. Congo] "Zaïre, Kivu, Beni".
Name-bearing type material: holotype ♂ in RMCA.

***Microjana latimaculata* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene latimaculata Dall'Asta & Poncin 1980: 475, figs 9–10, photo 1, fig. 7; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type: holotype ♂ in RMCA.

***Microjana morettoi* (Bouyer, 2012) comb. n.**
Stenoglene morettoi Bouyer 2012: 10, pl. 1, fig. 5, pl. 2, fig. 9; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 17.

Type locality: "Centrafrique, route Mambele-Mbaéré".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Microjana novempunctata* (Bethune-Baker, 1927) stat. rev., comb. n.**
Phasicnecus novempunctatus Bethune-Baker 1927: 327.
Stenoglene novempunctata (Bethune-Baker, 1927): Forbes 1955: 129 (comb.).
Stenoglene pellucida auct. (nec Joicey & Talbot): Dall'Asta & Poncin 1980: 467 (syn.); Kitching & Rougerie *et al*. 2018: suppl. 1; László *et al*. 2025: 17, figs 2, 4, 12, 13.
Stenoglene sp. ♀: László *et al*. 2025: 17, fig 5.

Type locality: "Bitje, Cameroons, 2000 ft.".
Name-bearing type material: holotype ♂ in NHMUK.

Notes. László *et al.* (2025) concluded that there was not enough evidence to revive *Phasicnecus novempunctatus* from synonymy with *Stenoglene pellucida* due to a limited number of samples and the lack of comparative topotypical material of the two taxa. However, from the material they examined, two morphospecies could clearly be delimited based on the male genitalia, while the possible existence of a third taxon was discussed. Despite the conservative approach of László *et al.* (2025), it is believed there is morphological and biogeographic evidence to assign the aforementioned two names to the two morphospecies without the need for the molecular analyses called for by the authors, and *M. novempunctata* (**stat. rev.**, **comb. n.**) is herein revived from synonymy with *M. pellucida*.

László *et al.* (2025: 16) stated the two morphospecies were "externally indistinguishable" although critical examination of the habitus reveals congruent wing markings, notably the diffuse black spot in space R_3 which is always present in *M. novempunctata* (László *et al.* 2025: Figs 2 and 4, as well as the topotypical female specimen illustrated in Fig. 5 as *Stenoglene* sp.), albeit very faintly in some individuals, while it is completely absent in the holotype of *M. pellucida* and the male from Tshopo (László *et al.* 2025: Figs 1 and 6 respectively). Additionally, they stated that "no obvious geographic isolation" had been recognised and that there was no "clear allopatry", without providing any reasoning as to why two species with quite different genitalia could not be found in sympatry. In any case, there is no evidence thus far of their sympatry, the three *M. pellucida* specimens from Lesse, Isiro and Tshopo distributed in the Middle Congo Basin and more specifically the Aruwimi and Lindi basins, separated from the Uele basin (where *M. novempunctata* is found) and the broader northern watershed/basins of Sangha and Ubangi (Van Pul 2023). Similar distributional patterns to *M. pellucida* are observed in monkeys (Flügel *et al*. 2015) and catchment-driven biodiversification is known in other groups of insects with flight capability (e.g., Price *et al*. 2010).

A single male specimen from an isolated population in southern D.R. Congo was suggested as being a third taxon in the complex by László *et al.* (2025) and despite it being confusingly referred to as *Stenoglene* sp. 1 on p. 15 and *Stenoglene* sp. 2 on p. 16, this specimen possesses highly distinctive genitalia and is described below as *Microjana bouyeri* **sp. n.**

***Microjana opalina* (Druce, 1910) comb. n.** (Figs 38, 47)
Chrysopoloma opalina Druce 1910: 176.
Stenoglene opalina (Druce, 1910): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 477; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 20.

Type locality: "Bitje, Ja River, Cameroons".
Name-bearing type material: 1 ♂ and 1 ♀ syntypes in NHMUK.

Phasicnecus peropalinus Rothschild, 1917
Phasicnecus peropalinus Rothschild 1917b: 492; Gaede 1927: 308.
Stenoglene peropalina (Rothschild, 1917): Forbes 1955: 129 (comb.).
Stenoglene peropalinus (Rothschild, 1917): Dall'Asta & Poncin 1980: 477 (syn.).

Type locality: "Bitje, Ja River, Camaroons".
Name-bearing type material: holotype ♀ in NHMUK.

***Microjana parvula* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene parvula Dall'Asta & Poncin 1980: 472, figs 7–8, photo 1, fig. 6; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

***Microjana pellucida* (Joicey & Talbot, 1924) comb. n.**
Phasicnecus pellucida Joicey & Talbot 1924: 559; Gaede 1927: 308.
Stenoglene pellucida (Joicey & Talbot, 1924): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 467 (partim); Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; László *et al*. 2025: 17, figs 1, 21.
Stenoglene sp. 2: László *et al*. 2025: 17, figs 6, 7, 15, 16.

Type locality: [D.R. Congo] "Lesse, Ituri Forest".
Name-bearing type material: holotype ♀ in NHMUK.

***Microjana pujoli* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene pujoli Dall'Asta & Poncin 1980: 473, figs 19–20, photo 1, fig. 10; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 18; Basquin & Faravel 2025: pl. 19.

Type locality: "La Maboke, Rép. Centrafricaine".
Name-bearing type material: holotype ♂ in MNHN.

***Microjana sinjaevi* (Bouyer, 2012) comb. n.**
Stenoglene sinjaevi Bouyer 2012: 11, pl. 1, fig. 3, pl. 2, fig. 15; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin & Faravel 2025: pl. 19.

Type locality: "Cameroun, r[oute] Edea-Douala pk 20".

Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Microjana smithi* (László & Giusti, 2025) comb. n.**
Stenoglene smithi László & Giusti, *In*: László *et al*. 2025: 18, figs 8–9, 17–18.

Type locality: "Republic of Congo, Nouabale-Ndoki National Park, Mondika Camp".
Name-bearing type material: holotype ♂ in ANHRT.

***Microjana subiridescens* (Holland, 1893) comb. n.**
Stibolepis subiridescens Holland 1893: 490, pl. 18, fig. 11; Aurivillius 1901: 18.
Phiala subiridescens (Holland, 1893): Gaede 1927: 300 (comb.); Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin & Faravel 2025: pl. 14.
Stenoglene subiridescens (Holland, 1893): Forbes 1955: 95, 129 (comb.).

Type locality: [Gabon] "Valley of the Ogové River".
Name-bearing type material: syntype ♀ in CMNH (photograph examined).

Notes. Originally described in the genus *Stibolepis* Butler, 1878 (= *Euchera* Hübner, 1825, Striphnopteryginae) and treated as belonging to it by Aurivillius (1901), it was assigned to the genus *Phiala* Wallengren, 1860 by Gaede (1927). Forbes (1955: fig. 12) dissected and figured the genitalia of a male syntype and correctly assigned it to *Stenoglene*, but it was subsequently omitted from *Stenoglene* by Dall'Asta & Poncin (1980) and listed as a *Phiala* by Kitching & Rougerie *et al*. (2018). Basquin & Faravel (2025) suggested this taxon probably belonged to *Stenoglene* and a revision was necessary. It is likely that this taxon represents a senior synonym of one of the other ferruginous *Microjana* **gen. n.** species, and further research is required to clarify the taxonomy.

***Microjana sulphureoides* (Kühne, 2007) comb. n.**
Stenoglene sulphureoides Kühne 2007: 347, figs 11–12, 23–24; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Kenya, Western Prov[ince], Kakamega Forest N.R., Colobus trail".
Name-bearing type material: holotype ♂ in coll. Kühne (photograph examined).

***Microjana sulphureotincta* (Strand, 1912) comb. n.**
Phasicnecus sulphureotinctus Strand 1912: 150; Gaede 1927: 308, pl. 41.
Stenoglene sulphureotincta (Strand, 1912): Forbes 1955: 129 (comb.).
Stenoglene sulphureotictus (Strand, 1912): Dall'Asta & Poncin 1980: 473; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin & Faravel 2025: pl. 21.

Type locality: [Equatorial Guinea] "Nkolentangan".
Name-bearing type material: holotype ♂ in MfN.

***Microjana uelei* (Dall'Asta & Poncin, 1980) comb. n.**
Stenoglene uelei Dall'Asta & Poncin 1980: 477, figs 13–14, photo 1, fig. 9; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

Description of new species

***Microjana bouyeri* sp. n.** (see László *et al*. 2025: Figs 3, 14)
https://www.zoobank.org/urn:lsid:zoobank.org:act:78F1776E-83E6-4E22-94FF-451B975EE50C

Stenoglene pellucida auct. (nec Joicey & Talbot): Dall'Asta & Poncin 1980: 467 (partim), photo 1, fig. 13.
Stenoglene sp. 1: László *et al*. 2025: 15, 18, figs 3, 14.
Stenoglene sp. 2: László *et al*. 2025: 16.

Holotype ♂ (see László *et al*. 2025: Fig. 3):
"COLL. MUS. CONGO | Elisabethville | 22-1-1956 | Ch. Seydel [black border; partially handwritten] || G.Poncin dét.I9 | Genre Stenoglene | Esp. pellucida | Prép.N° 652 [black border; partially handwritten] || RMCA ENT | 000049943 [black border; QR code]" (RMCA).

Description. Forewing length: 17 mm.

Male (see László *et al.* 2025: Fig. 3). Upperside. Ground colour of body and wings creamy-white. Antennae brown, bipectinate, with creamy-white scaling along entire length. Forewing elongate triangular, costal margin gently arcuate, apex rounded, outer margin arcuate to rounded tornus; costal area irrorated with dark grey scaling to apex, continuing along terminal area where its inner edge is crenulate, to tornus. Submarginal fascia indicated by black spots in spaces R_4, M_3 and Cu_1, as well as a cluster of black scales at the termination point along the anal margin. Fringe creamy-white irrorated with dark grey. Hindwing ovoid, outer margin evenly curved. Submarginal fascia indicated by black spots in spaces M_3 and Cu_1, continuing thereafter in the form of an indistinct band towards anal margin. Terminal area irrorated with dark grey scaling, its inner edge crenulate. Underside. As on upperside but uniform, lacking markings except for spots showing through from upperside. Male genitalia (see László *et al.* 2025: Fig. 14). Uncus medially reduced with pair of bifurcate lateral processes each broad at base, the inner lobe apically pointed, curved outward, robust, nearly twice as wide and a third of the length of long, inwardly-curved, apically-pointed outer lobe. Tegumen broad. Gnathos reduced, membranous. Mesal protrusions separated, small, elongate ovoid and setose. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender, tapering towards bifurcate apex, each process evenly rounded. Saccular section elongate triangular, twice as broad as costal section at base, tapering to apically-pointed distal process, curved dorsad, with postmedial denticle along dorsal margin. Juxta semi-cylindrical, subtrapezoidal. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus strongly curved, almost right-angled, constricted medially, tapering towards pointed carina; coecum rounded twice as broad as carina.

Female. Unknown.

Diagnosis. The new species is most similar to *M. pellucida*, these two species differing from the allied *M. novempunctata* which possesses a spot in forewing space R_3 and a distally bifurcate juxta in the male genitalia. The habitus of *M. bouyeri* **sp. n.** and *M. pellucida* is virtually indistinguishable although the spot in space M_3 of both wings are smaller in the latter. The male genitalia are however highly diagnostic, the valves in *M. pellucida* with shorter apical processes of the costal section, and the absence of the postmedial denticle of the distal saccular process. Since these two species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Molecular characterisation. Unknown.

Derivatio nominis. It is with great pleasure that this new species is dedicated to Thierry Bouyer, for his important taxonomic contributions to the Afrotropical Eupterotidae fauna and for his extensive entomological research in the old Katanga Province of D.R. Congo from where this species originates. The patronym is a noun in the genitive case.

Distribution. The new species is known only from the type locality in the Haut-Katanga Province of southern D.R. Congo, and is the most southerly distributed species in the genus.

***Microjana hikaru* sp. n.** (Figs 39–40, 49)
https://www.zoobank.org/urn:lsid:zoobank.org:act:3C66D987-3B5A-430C-9EE1-8AC5D2FC0231

Holotype ♂ (Fig. 39):
"REPUBLIC OF CONGO 352m | Sangha Prov., Nouabale-Ndoki | National Park, Ndoki formation | (Secondary forest) | 02°12'47.7"N, 16°23'45.8"E | 29.ix–1.x.2022, MV Light Trap | Dérozier,V., Fouka,B., | Kirk-Spriggs,A., Takano,H. Leg. | ANHRT:2022.14 || ANHRTUK | 00290012" (ANHRT).

Paratypes (8 ♂♂, 2 ♀♀):
REPUBLIC OF CONGO: same data as holotype (2 ♂♂ ANHRT); Nouabalé-Ndoki National Park, Bomassa camp, 341 m, 02°12′36.9″N, 16°11′30.2″E, 16–23.ix.2022 (1 ♂ ANHRT); same locality as previous but 06–14.v.2023, leg. V. Dérozier, A. Kirk-Spriggs, G. László & S. Mvouende (1 ♂, 1 ♀ ANHRT); Nouabalé-Ndoki National Park, Makao camp, 377 m, 02°35′42.2″N, 17°10′08.3″E, 15–21.v.2023, leg. V. Dérozier, A. Kirk-Spriggs, G. László & S. Mvouende (1 ♂ ANHRT); Nouabalé-Ndoki National Park, Mbeli camp, 372 m, 02°14′23.8″N, 16°23′52.1″E, 01–10.x.2022 (1 ♂, 1 ♀ ANHRT); Nouabalé-Ndoki National Park, Mondika camp, 365 m, 02°21′50.63″N, 16°16′25.82″E, 07–14.ii.2023, leg. N.M. Bakala, V. Dérozier, A. Kirk-Spriggs & G. László (1 ♂ ANHRT); same locality as previous but 27.iv–06.v.2023, leg. V. Dérozier, A. Kirk-Spriggs, G. László & S. Mvouende (1 ♂ ANHRT).

Description. Forewing length: male: 18–20 mm, female: 23–24 mm.

Male (Fig. 39). Upperside. Ground colour of body and wings canary-yellow; vertex of head dark brown. Antennae brown, bipectinate, with yellow scaling at base. Forewing triangular, costal margin gently arcuate, apex rounded, outer margin arcuate to rounded tornus. Discal spot dark brown, minute. Postmedial fascia dark brown, indicated by spots, irregularly sinuate, and strongly displaced proximally in space Cu_2. Submarginal fascia dark brown, arcuate, indicated by spots. Subterminal fascia greyish-brown, arising at subapical spot, curved following outer margin, indicated by crenules and chevrons. Oblique band reddish-brown, straight, slightly diffuse, from apex terminating two-thirds of the way along anal margin; inner margin of band between veins M_3 and Cu_2 with two small ovoid markings, each silvery-grey and edged in greyish-brown. Fringe canary-yellow except at apex where it is reddish-brown in line with oblique band. Hindwing rounded, outer margin evenly curved. Postmedial fascia dark brown, gently arcuate, interrupted along veins. Submarginal fascia dark brown, curved, diffuse, fading into a band of small grey ovoid markings. Subterminal fascia greyish-brown, strongly interrupted, indicated by chevrons. Fringe yellow. Underside. Ground colour uniform canary-yellow. Wing markings absent except for oblique forewing band showing through from upperside. Male genitalia (Fig. 49). Uncus medially reduced with pair of lateral subrectangular processes, each with a long curved inner hook, half the length of the entire uncus, and a short outer tooth-like, apically-pointed projection. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, small, ovoid and setose. Valve elongate triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender, tapering towards bifurcate apex, each process rounded. Saccular section elongate triangular, as broad as costal section, tapering to bifurcate apex with rounded processes, and small dentate pad at based of dorsal process. Juxta semi-cylindrical, trapezoidal. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus curved and constricted medially, tapering towards carina; coecum rounded; vesica globular with a small patch of scobination.

Female (Fig. 40). Similar to male but rami of antenna slightly shorter and ground colour a darker yellow. Wing markings as in male but generally broader or larger and more diffuse; outer half of wings strongly irrorated with greyish-brown scales. Hindwing postmedial fasciae curved and sharply angled between vein Cu_2 and anal margin.

Diagnosis. Of the six canary-yellow *Microjana* species with an irregular forewing antemedial fascia (*M. brunneofasciata*, *M. citrina*, *M. citrinoides*, *M. parvula* and *M. tabithae* **sp. n.**), *M. hikaru* **sp. n.** shares the straight and gently arcuate hindwing antemedial fascia with *M. citrina* and *M. tabithae* which are virtually indistinguishable based on habitus alone. The oblique brown fascia of the forewing is perhaps widest in *M. hikaru*, especially near the apex in comparison to its congeners. The male genitalia are however highly distinct, differing from *M. citrina* and *M. tabithae* in the elongate lateral processes of the uncus (which are shorter, more robust and strongly curved in these congeners) and the

short bifurcate apex of the saccular section of the valve (the dorsal process at least three times longer in the others). The genitalia of *M. hikaru* is most similar to *M. parvula* (see Dall'Asta & Poncin 1980: fig. 7) but in the latter, the lateral processes of the uncus are more robust and shorter, the distal edge of the process is strongly serrate (smooth in *M. hikaru* with a sharp denticle pointed ventrad), and the bifurcate apices of the costal and saccular sections of the valve are more weakly developed.

Molecular characterisation. The new species has been assigned the BIN BOLD:AAM8653. Intraspecific pairwise distances were 0.0–0.9% (n=4) and diverged from *M. citrina* by 11.0–11.4% (n=3).

Derivatio nominis. It is with great pleasure that this new species is dedicated to the author's nephew, in the hope that it instils in him the importance of conserving and caring for the natural world. The patronym is a noun in apposition.

Distribution. The new species is known only from the type locality in northern Republic of Congo.

***Microjana tabithae* sp. n.** (Figs 41–42, 48)
https://www.zoobank.org/urn:lsid:zoobank.org:act:39EE7EE0-D635-4AD8-A94C-6900DEF9F268

Holotype ♂ (Fig. 41):
"REPUBLIC OF CONGO 352m | Nouabale-Ndoki National Park, | Mombongo camp | 02°10'30.7"N, 16°8'37.7"E | 02–07.ii.2023, MV light trap | Bakala N., M., Dérozier, V., | Kirk-Spriggs, A., László, G. leg. | ANHRT:2023.3 || ANHRTUK | 00329737" (ANHRT).

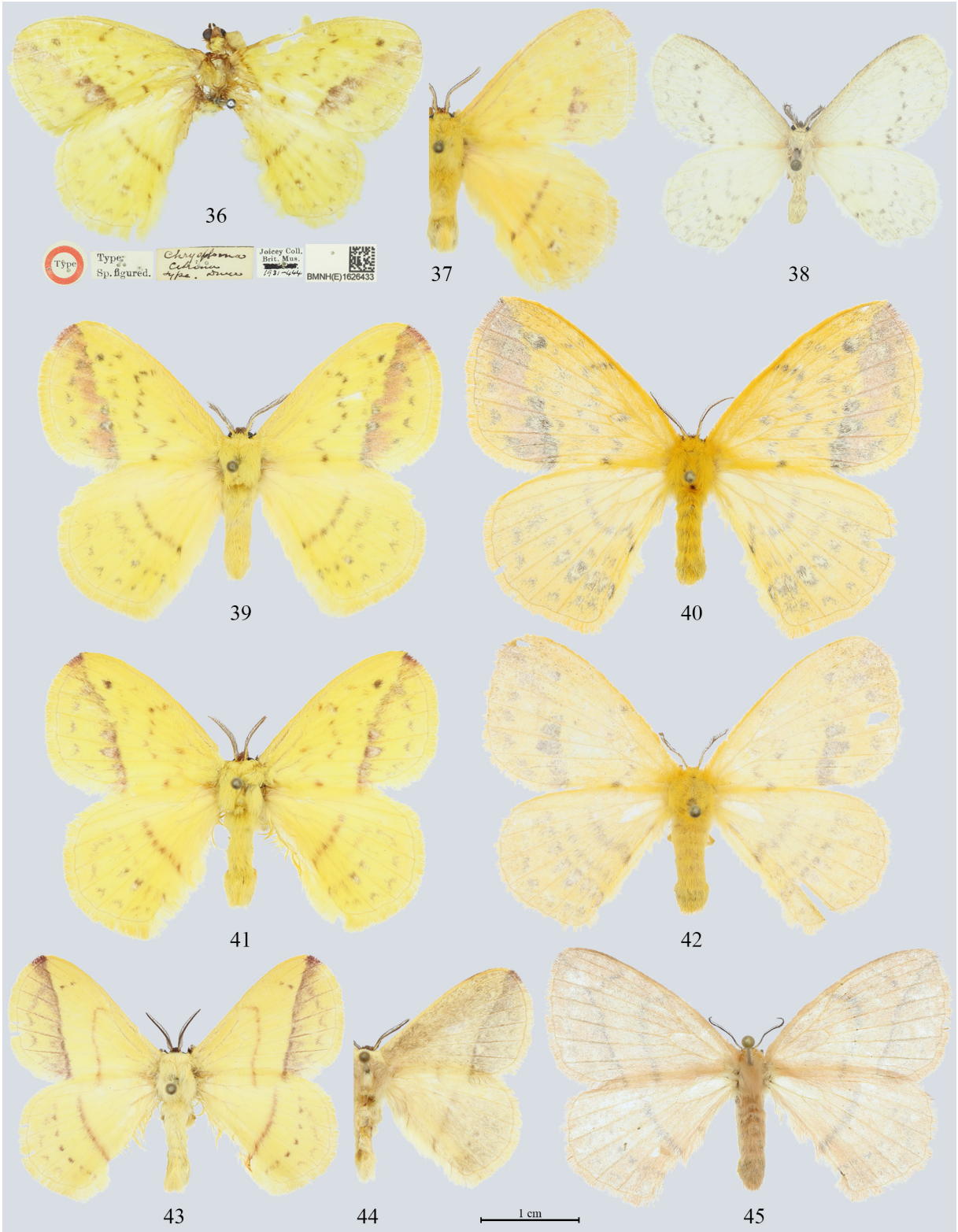
Paratypes (2 ♂♂, 1 ♀):
REPUBLIC OF CONGO: same data as holotype (1 ♀ ANHRT); Nouabalé-Ndoki National Park, Bomassa camp, 341 m, 02°12′36.9″N, 16°11′30.2″E, 06–14.v.2023, leg. V. Dérozier, A, Kirk-Spriggs, G. László & S. Mvouende (2 ♂♂ ANHRT).

Description. Forewing length: male: 18–20 mm, female: 21 mm.

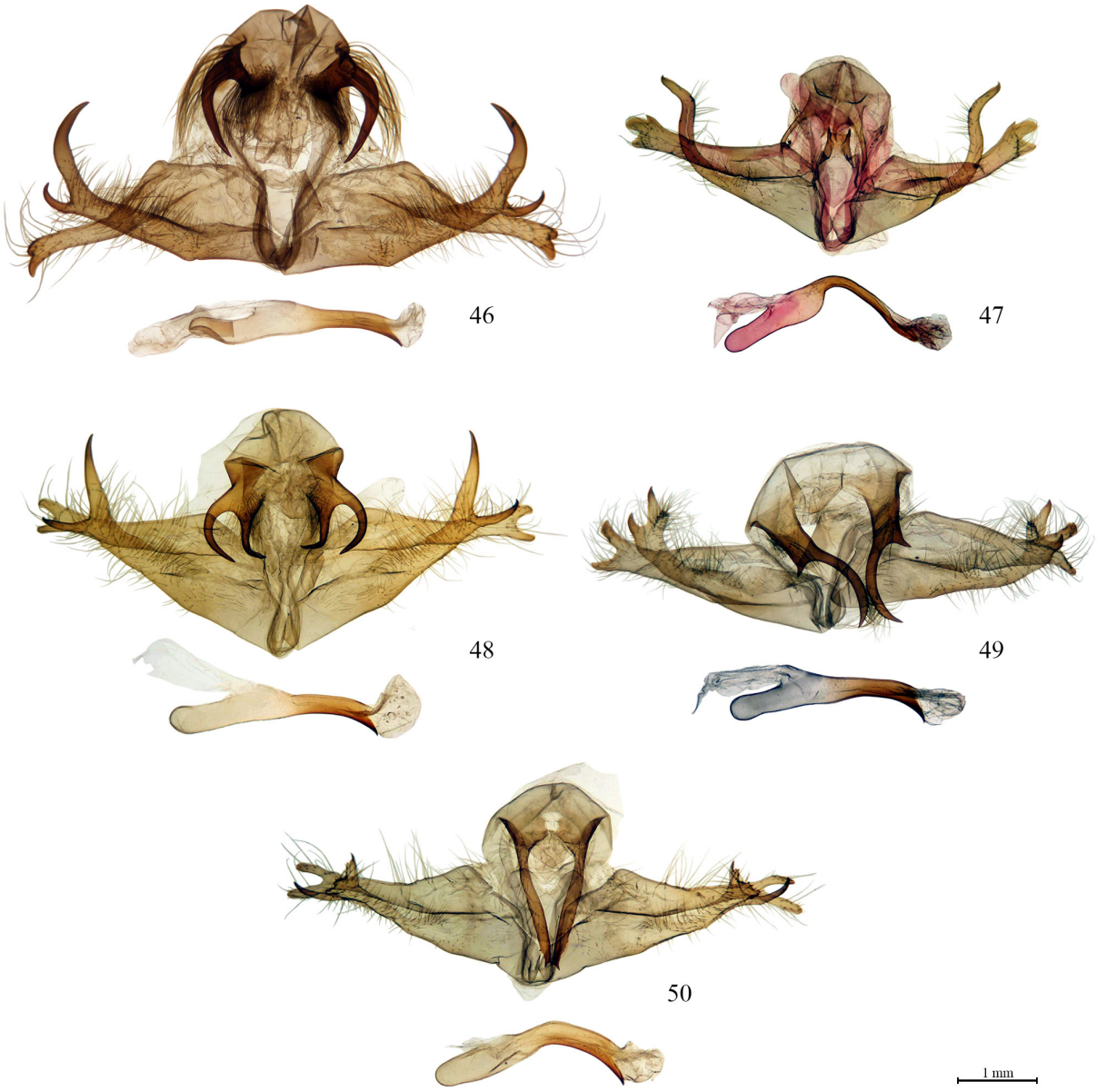
Male (Fig. 41). Upperside. Ground colour of body and wings canary-yellow; vertex of head mahogany. Antennae brown, bipectinate, with yellow scaling at base. Forewing triangular, costal margin gently arcuate, apex rounded, outer margin arcuate to rounded tornus. Discal spot mahogany, minute. Postmedial fascia mahogany, indicated by spots, irregularly sinuate, and strongly displaced proximally in space Cu_2. Submarginal fascia mahogany, arcuate, indicated by spots. Subterminal fascia dark-brown, arising at subapical spot, curved following outer margin, indicated by crenules and chevrons. Oblique band brown, gently arcuate, slightly diffuse, from apex terminating two-thirds of the way along anal margin; inner margin of band between veins M_3 and Cu_2 with two small ovoid markings, each creamy-white and edged in brown. Fringe canary-yellow except at apex where it is brown in line with oblique band. Hindwing rounded, outer margin evenly curved. Postmedial fascia mahogany, straight, interrupted along veins. Submarginal fascia mahogany, gently curved, diffuse, fading into a band of small dark brown ovoid markings. Subterminal fascia dark-brown, strongly interrupted, indicated by chevrons. Fringe yellow. Underside. Ground colour uniform canary-yellow. Wing markings absent except for oblique forewing band showing through from upperside. Male genitalia (Fig. 48). Uncus medially reduced with pair of bifurcate lateral processes, each broad at base with slight concavity along outer margin to two long apically-pointed processes, the inner slightly shorter and almost right-angled near apex, the outer evenly curved. Tegumen broad. Gnathos reduced, membranous. Mesal protrusions separated, elongate ovoid and heavily setose. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender, tapering towards bifurcate apex, each process rounded. Saccular section elongate triangular, twice as broad as costal section at base, tapering to bifurcate apex, each process sharply pointed, the dorsal process twice as long as the ventral process. Juxta semi-cylindrical, trapezoidal. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus curved and constricted medially, tapering towards carina; coecum rounded; vesica globular with a small patch of scobination.

Female (Fig. 42). Similar to male but rami of antenna slightly shorter and ground colour a darker yellow. Wing markings as in male but generally broader or larger and more diffuse; outer half of wings

strongly irrorated with greyish-brown scales. Forewing oblique band interrupted between R_5 and M_2. Hindwing postmedial fasciae gently curved and angled between vein Cu_2 and anal margin.



Figures 36–45. *Microjana* **gen. n.** species, habitus (in ANHRT unless otherwise stated). 36. *M. citrina* (Druce, 1886), holotype ♂ of *Chrysopoloma citrina* Druce (NHMUK). 37. *Id*., ♂, Gabon, Nyonié. 38. *M. opalina* (Druce, 1910), ♂, Republic of Congo, Nouabalé-Ndoki NP. 39. *M. hikaru* **sp. n.**, holotype ♂. 40. *Id*., paratype ♀. 41. *M. tabithae* **sp. n.**, holotype ♂. 42. *Id*., paratype ♀. 43. *M. fontainei* (Dall'Asta & Poncin, 1980), ♂, Republic of Congo, Nouabalé-Ndoki NP. 44. *Id*., ♂, Republic of Congo, Odzala-Kokoua NP. 45. *Id*., ♀.



Figures 46–50. *Microjana* **gen. n.** species, ♂ genitalia. 46. *M. citrina* (Druce, 1886). 47. *M. opalina* (Druce, 1910). 48. *M. tabithae* **sp. n.**, holotype. 49. *M. hikaru* **sp. n.**, holotype. 50. *M. fontainei* (Dall'Asta & Poncin, 1980).

Diagnosis. *Microjana tabithae* **sp. n.** is phenotypically identical to *M. citrina* but the male genitalia are highly distinct. In the latter, the lateral processes of the uncus comprise a single curved hook-like projection (two projections in the former) and the dorsal process of the bifurcate apex of the saccular section of the valve is longer and strongly recurved (straight in the former). See under *M. hikaru* **sp. n.** for further diagnoses.

Molecular characterisation. Unknown.

Derivatio nominis. It is with great pleasure that this new species is dedicated to Tabitha Taberer, ANHRT, a specialist of the Afrotropical Limacodidae, for her kind assistance during the preparation of this and numerous other manuscripts. The matronym is a noun in the genitive case.

Distribution. The new species is known only from the type locality in northern Republic of Congo.

***Artemisana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:219DC3C9-2A32-4A18-A5D0-21249743F92D

Type species: *Chrysopoloma thelda* Druce, 1887.

Description. Adults. Small moths. Antenna bipectinate, rami much shorter in females. Ground colour of body brown, heavily irrorated with pale-beige or grey scales; forewing beige, irrorated with dark brown scaling giving a speckled appearance; hindwing yellow; generally darker in females. Forewing triangular, weakly pointed at apex in males and produced in females, outer margin strongly curved and weakly crenulate; R_4 + R_5 stalked. Discocellular marking bar-shaped, sometimes absent. Postdiscal fascia dark brown, recurved, strongly crenulate arising at small triangular marking halfway along costal margin. Submarginal fasciae darker chocolate brown, interrupted and indicated by dashes and lunules with two or three spots near anal margin; arises at dark brown triangular marking along costal margin near apex, and follows outline of postdiscal fascia. Subterminal fascia brown running parallel to outer margin, with a protruding recurved chevron-shaped proximal expansion medially. Hindwing ovoid, outer margin rounded and strongly crenulate to a produced tornus. Postmedial and submarginal fasciae dark brown, crenulate, the former sometimes weakly expressed; curved band indicated by spots between veins, placed distad of submarginal fascia. Termination points of fasciae and band along anal margin indicated by dark brown markings. Fringe long. Underside. Ground colour warm yellow, slightly darker in females; wings with dark brown fasciae and markings. Forewing with narrow dark brown border along outer margin. Postmedial fascia becoming ill-defined towards anal margin; submarginal and subterminal fasciae visible only near costal. Hindwing markings as on upperside but well-defined. Additionally, basal region irrorated irregularly with brown scales and area between postmedial and submarginal fasciae and between submarginal fasciae and termen irrorated with brown scales, forming curved bands. Male genitalia. Uncus medially reduced with pair of long gently tapering lateral processes. Tegumen broad. Gnathos heavily reduced, membranous. Mesal protrusions separated, elongate ovoid, heavily setose. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, tapering towards bifurcate apex. Saccular section rectangular, twice as broad as costal section, terminating in two broad, hooked processes, the dorsal one longer than the ventral one. Juxta semi-cylindrical, inverse-triangular with rounded ventral margin. Vinculum narrow, V-shaped. Saccus rounded at apex. Phallus slender, coecum rounded, produced at the entrance of ductus ejaculatorius, carina hooked; vesica elongate with two patches of scale-like scobination.

Diagnosis. The distinct colouration and wing shapes of both sexes as well as the genitalia of the sole member of this genus is unique and cannot readily be confused with any other species of Afrotropical Eupterotidae

Derivatio nominis. The compound genus name is derived from the Greek goddess Artemis, and her Roman equivalent Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. The distribution of the single species in the genus is centred around the Guineo-Congolian rainforests but it is also found in areas of forest-savanna mosaics.

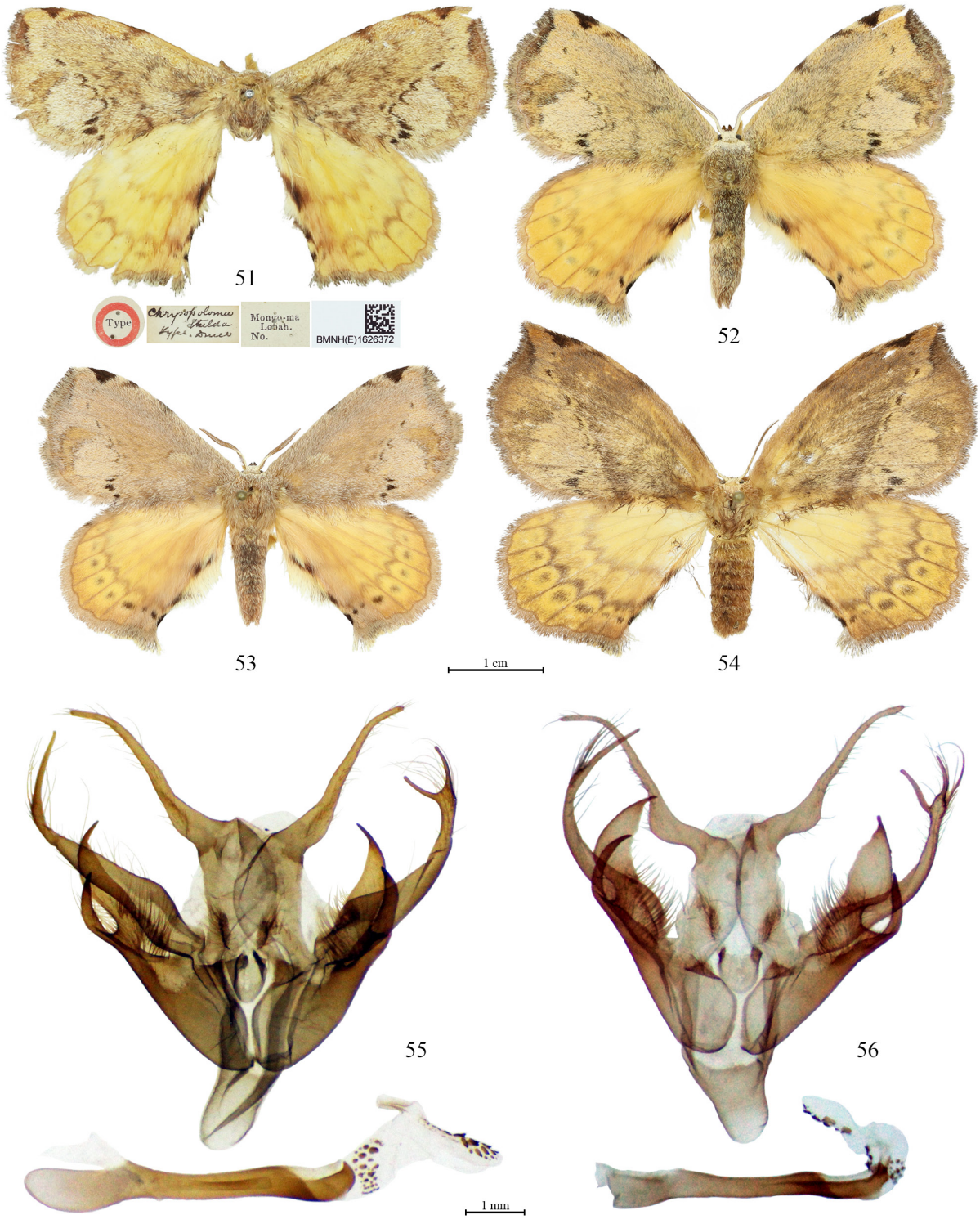
Species content: monotypic.

***Artemisana thelda* (Druce, 1887) comb. n.** (Figs 51–56)
Chrysopoloma thelda Druce 1887: 677.
Phasicnecus thelda (Druce, 1887): Aurivillius 1901: 30 (comb.); Gaede 1927: 309.
Stenoglene thelda (Druce, 1887): Kirby 1892: 810 (comb.); Dall'Asta & Poncin 1980: 480, photo 2, fig. 24; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 8, 19; Basquin & Faravel 2025: pl. 34.

Type locality: [Cameroon] "Mongo-ma Lubah".
Name-bearing type material: holotype ♂ in NHMUK.

Notes. Despite the large barcode divergences between two West African (Sierra Leone, Côte d'Ivoire) and one Central African (Republic of Congo) samples (PWDs of 5.8–6.1%), no differences

were observed in the male genitalia and it is herein considered to be a single widespread and variable species.



Figures 51–56. *Artemisana* **gen. n.** *thelda* (Druce, 1887) (in ANHRT unless otherwise stated). 51. Holotype ♂ of *Chrysopoloma thelda* Druce (NHMUK). 52. ♂, Republic of Congo, Nouabalé-Ndoki NP. 53. ♂, Liberia, Wologizi Mts. 54. ♀, *Id*. 55. ♂ genitalia (Republic of Congo). 56. ♂ genitalia (Liberia).

***Streptojana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:BCC6E4EB-ADAE-4770-BA67-5784F2792A33

Type species: *Phasicnecus nivalis* Rothschild, 1917.

Description. Adults. Small moths. Antenna bipectinate, rami shorter in females. Ground colour creamy-white or brown. Forewing triangular with rounded outer margin, gently angular in females; $R_4 + R_5$ stalked. Hindwing rounded. Postmedial, submarginal and subterminal fasciae silvery-grey or dark brown, strongly crenulate and forming marginal spots in most species; markings generally weakly-defined in females. Underside as on upperside but weakly marked and uniform. Male genitalia. Uncus medially reduced with pair of single lateral processes. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions either weakly fused medially or separated and ovoid; partly setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender triangular, tapering towards recurved distal process. Saccular section triangular, tapering to strongly curved or sigmoidal distal process. Juxta semi-cylindrical, trapezoidal, sometimes bilobed distally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus elongate, coecum rounded and tapering towards carina; vesica with small patch of scobination.

Diagnosis. The genus contains species with two distinctive phenotypes but are united by the male genitalia which all share a distinctive sigmoidal or twisted distal saccular process. The white species are somewhat reminiscent of *Pterocerota* species but can easily be distinguished based on the forewing venation, with $R_4 + R_5$ stalked in *Streptojana* **gen. n.** but unstalked in the latter. The genitalia of *Streptojana* species are most similar to the phenotypically dissimilar *Drepanojana* and differ in the presence of well-sclerotised lateral sclerites of the tegumen (reduced in the latter), the broader costal section of the valve, the presence of a projection on the outer edge at the base of the distal saccular process (a small projection is found on the inner edge at the first angle of the distal saccular process in the latter) and the carina of the phallus tapered to a point (in *Drepanojana*, the carina forms a large plate-like structure with a dentate margin).

Derivatio nominis. The compound genus name is derived from the Ancient Greek for 'twisted' (στρεπτός), in reference to the shape of the distal saccular process in the male genitalia, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Members of this genus are found in rainforest habitats from West Africa to the Albertine Rift.

Species content: five species are assigned to this genus, three of which are new species.

***Streptojana brunnea* (Bouyer, 2012) comb. n.** (Figs 63, 67)
Stenoglene brunneus Bouyer 2012: 6, pl. 3, fig. 23; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Cameroun, r. Edea-Douala pk 20".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Streptojana nivalis* (Rothschild, 1917) comb. n.** (Figs 59, 64)
Phasicnecus nivalis Rothschild 1917b: 492; Gaede 1927: 308, pl. 46.
Stenoglene nivalis (Rothschild, 1917): Forbes 1955: 95, 129 (comb.); Dall'Asta & Poncin 1980: 466; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Bingerville, Ivory Coast".
Name-bearing type material: holotype ♂ in NHMUK.

Description of new species

***Streptojana forbesi* sp. n.** (Figs 61–62, 68)
https://www.zoobank.org/urn:lsid:zoobank.org:act:55E210BE-B470-40B4-AC39-F0CCE8A6CEDD

Holotype ♂ (Fig. 61):
"LIBERIA 883m | Lofa County, Wologizi Mts., | Ridge Camp 2 | 8°7'20.79"N, 9°56'50.75"W | 22-30.xi.2018 Light Trap | Blended Bulb (250W) | Sáfián, Sz., Simonics, G. Leg. | ANHRT:2018.43 || ANHRTUK | 00230463"

Paratypes (27 ♂♂, 2 ♀♀):
LIBERIA: same data as holotype (2 ♂♂ ANHRT); Nimba Mountains, East Nimba Nature Reserve, Cellcom Road, 1000–1100 m, 07°32′45.88″N, 08°31′21.04″W, 16–28.xii.2018, leg. Sáfián & G. Simonics (1 ♂ ANHRT); Wologizi Mountains, Base Camp forest, 611 m, 08°07′17″N, 09°57′42″W, 20.xi–01.xii.2017, leg. M. Aristophanous, S. Sáfián, G. Simonics & L. Smith (2 ♂♂ ANHRT); Wologizi Mountains, Ridge Camp, 865 m, 08°07′10″N, 09°57′11″W, 24–29.xi.2017, leg. M. Aristophanous, S. Sáfián, G. Simonics & L. Smith (18 ♂♂, 2 ♀♀ ANHRT); Wologizi Mountains, Rosewood Camp, 585 m, 08°06′14.9″N, 09°58′27.3″W, 18.xi–01.xii.2018, leg. S. Sáfián & G. Simonics (2 ♂♂ ANHRT); Wetezu camp, Wonegizi Nature Reserve, 551 m, 08°04′57.11″N, 09°34′47.86″W, 19–27.iii.2019, leg. S. Safian & S. Koivogui (2 ♂♂ ANHRT).

Description. Forewing length: male: 18–20 mm, female: 26 mm.

Male (Fig. 61). Upperside. Ground colour of body and wings brown. Antennae brown, bipectinate. Forewing triangular, costal margin arcuate to angled apex, outer margin curved. Costa irrorated with greyish scaling. Discocellular weakly highlighted in dark brown. Postmedial fascia dark brown, diffuse, curved, irregularly crenulate and displaced proximad near anal margin. Submarginal fascia dark brown, diffuse, irregularly sinuose, displaced proximad between veins M_1 and Cu_1; outer edge highlighted with dark greyish-brown dots and chevrons tipped with beige scales. Subterminal fascia arising at apex, running at oblique angle and furthest from outer margin at vein M_2, and terminating near tornus; are beyond fascia to outer margin darker brown. Hindwing rounded, outer margin curved. Postmedial and submarginal fasciae as on forewing but curved, the latter slightly angled. Underside. Ground colour slightly paler than on upperside. Wings lack markings of upperside except for poorly-defined and diffuse dark brown postmedial fascia. Male genitalia (Fig. 68). Uncus medially reduced with pair of broad, digitate processes. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, ovoid and setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section triangular, tapering to bifurcate apex, the tines pointed dorsad. Saccular section triangular, twice as broad as costal section, tapering into long sigmoidal distal process, as long as sacculus, apically dilated and spatulate. Juxta semi-cylindrical, inverse-triangular. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus elongate, coecum rounded and sharply dilated at carina.

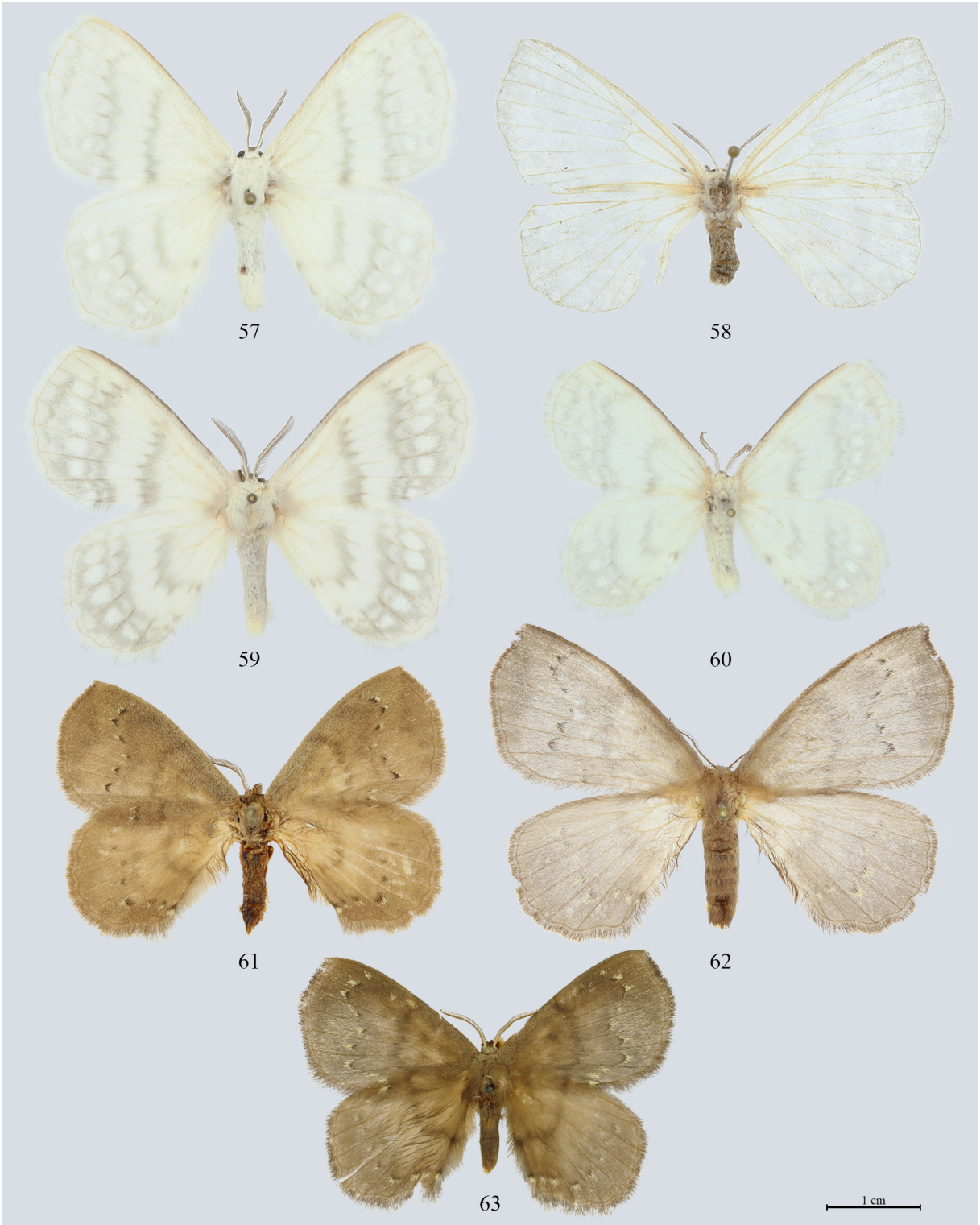
Female (Fig. 62). As in male but rami of antenna shorter; wings elongate and outer margin strongly curved. Forewing apex produced with subapical concavity along outer margin. Wings slightly diaphanous, darker in outer third and pattern weakly expressed.

Diagnosis. The new species is phenotypically indistinguishable from *S. brunnea*, however the male genitalia are diagnostic. In the male genitalia of *S. brunnea*, the processes of the uncus are shorter and broader, the costal section of the valve is shorter, more robust with a rounder margin, and the two apical processes are positioned closer together with the distal one half as small as in *S. forbesi* **sp. n.** Since these two species are seemingly found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Molecular characterisation. The new species has been assigned the BIN BOLD:AFF4367. Intraspecific pairwise distances were 0.0–0.5% (n=4) and diverged from the holotype of *S. brunnea* by 2.8–3.0%.

Derivatio nominis. The new species is named after William T.M. Forbes (1885–1968) whose invaluable work on the Eupterotidae in the mid-20th Century has formed the basis of all modern studies on this family of moths. The patronym is a noun in the genitive case.

Distribution. Known only from Liberia where it is distributed at higher altitudes and it is almost certainly endemic to the Upper Guinean forests of West Africa.



Figures 57–63. *Streptojana* **gen. n.** species, habitus (in ANHRT unless otherwise stated). 57. *S. molliae* **sp. n.**, holotype ♂. 58. *Id*., paratype ♀. 59. *S. nivalis* (Rothschild, 1917), ♂, Liberia, Wologizi Mts. 60. *S. laurenae* **sp. n.**, holotype ♂. 61. *S. forbesi* **sp. n.**, holotype ♂. 62. *Id*., paratype ♀. 63. *S. brunnea* (Bouyer, 2012), ♂, Cameroon, Johann-Albrechtshöhe (NHMUK).

***Streptojana laurenae* sp. n.** (Figs 60, 65)
https://www.zoobank.org/urn:lsid:zoobank.org:act:CE61B141-CB6D-4EA1-A713-C939BA09C7D9

Holotype ♂ (Fig. 60):
"DRC 950m | Mamove, Nord-Kivu | 0°49'N, 29°27'E | ix.2015 ex. A. Colley | ANHRT:2018.22 || ANHRTUK | 00158611" (ANHRT).

Description. Forewing length: 18 mm.

Male (Fig. 60). Upperside. Ground colour of body greyish white, wings snowy white. Antennae brown, bipectinate with white scaling along its entire length. Forewing elongate triangular, dark beige costal margin gently arcuate to rounded apex, outer margin curved to rounded tornus; hindwing elongate, outer margin curved. Postmedial fascia slivery-grey, curved, its outer edge strongly crenulate; forewing fascia recurved at anal margin. Submarginal and subterminal fasciae silvery-grey, broadly following outer margin; subterminal fascia weakly defined. Forewing fasciae more irregular. Space between submarginal and subterminal fasciae filled with reflective pearly-white spots edged with silvery-grey, between veins. Hindwing terminal region irrorated with silvery-grey scales. Underside. As on upperside. Male genitalia (Fig. 65). Uncus medially reduced with pair of long, apically rounded lateral processes. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions weakly fused medially, each lobe with small denticle distally and sparse setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender triangular, tapering to small recurved hook. Saccular section triangular, as broad as costal section, tapering to flattened, sigmoidal, apically-rounded distal process with small, rounded outer projection at its base. Juxta semi-cylindrical, trapezoidal. Vinculum narrow, V-shaped. Saccus very short, rounded at apex. Phallus elongate, coecum pointed and tube slightly upturned at carina; vesica with weak scobination.

Female. Unknown.

Diagnosis. The smallest member of the genus, and of the white species, it is the only one with a curved forewing postmedial fascia. In the male genitalia, it is most similar to *S. nivalis* and *S. molliae* **sp. n.** but differ from these two in the following: in *S. nivalis*, the lateral processes of the uncus are curved inwards, the mesal protrusions are more weakly sclerotised medially, the distal costal process is larger and more hooked, the distal saccular process is straighter and a little shorter, and the phallus is more robust; in *S. molliae*, the lateral processes of the uncus are broader, the mesal protrusions are narrower and greatly bilobed, the distal costal process is bifurcate, the distal saccular process is broader and shorter, and the phallus is more robust.

Molecular characterisation. The new species has been assigned the BIN BOLD:AFG4352 and diverged from its sibling *S. nivalis* by 10.5% (n=1).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Lauren Morgan, ANHRT, for her interest and kind assistance in re-curating the Eupterotidae collections. The matronym is a noun in the genitive case.

Distribution. Known only from the type locality on the western slopes of the Albertine Rift. It is possible that the specimens cited from northeastern D.R. Congo in Dall'Asta & Poncin (1980) refer to this species.

***Streptojana molliae* sp. n.** (Figs 57–58, 66)
https://www.zoobank.org/urn:lsid:zoobank.org:act:D079A930-2C9D-40DC-9FDE-9AFA386E15A2

Holotype ♂ (Fig. 57):
"REPUBLIC OF CONGO 480m | Odzala-Kokoua National Park, | Imbalanga Camp | 00°45'47"N, 15°15'39"E | 07–08.iv.2024, LepiLED light | trap. Bashford,M., László,G., | Talani,M., Yaba Ngouma,S. leg. | ANHRT:2024.7 || ANHRTUK | 00405099" (ANHRT).

Paratypes (62 ♂♂, 1 ♀ ANHRT):
REPUBLIC OF CONGO: same data as holotype (1 ♂ ANHRT); same data as for preceding but 05–09.iv.2024 (1 ♂ ANHRT); same data as for preceding but 05–11.iv.2024 (3 ♂♂ ANHRT); same locality as for preceding but 04–05.ix.2024, leg. M. Bashford, G. László, M. Talani & A. Volynkin (2 ♂♂

ANHRT); same data as for preceding but 13–20.ix.2024 (1 ♂ ANHRT); Odzala-Kokoua National Park, Camp Imbalanga, 490 m, 00°45′59″N, 15°15′20″E, 14–17.xi.2024, leg. M. Bashford, I. Elliott & A. Kirk-Spriggs (2 ♂♂ ANHRT); Odzala-Kokoua National Park, Bangassou Forest near Lobo, 400 m, 00°32′50″N, 14°51′47″E, 23–26.ix.2024, leg. M. Bashford, G. László, M. Talani, A. Volynkin & S. Yaba Ngouma (2 ♂♂ ANHRT); Odzala-Kokoua National Park, Kilo Forest, 500 m, 00°31′37″N, 14°52′20″E, 23–24.ix.2024, leg. M. Bashford, G. László & A. Volynkin (2 ♂♂ ANHRT); Odzala-Kokoua National Park, Kokoua base, 540 m, 01°28′39″N, 15°16′42″E, 05–13.ix.2024, leg. M. Bashford, G. László, M. Talani & A. Volynkin (16 ♂♂ ANHRT); same locality as for preceding but 11–13.ix.2024, leg. M. Bashford, G. László, M. Talani, A. Volynkin & S. Yaba Ngouma (5 ♂♂ ANHRT); Odzala-Kokoua National Park, Lango baï, 373 m, 00°36′03.83″S, 14°56′02.21″E, 24–25.ix.2024, leg. M. Bashford, G. László, M. Talani, A. Volynkin & S. Yaba Ngouma (10 ♂♂ ANHRT); Odzala-Kokoua National Park, Lekoli River near Mboko, 379 m, 00°37′04.79″N, 14°54′27.32″E, 21–23.ix.2024, leg. M. Bashford, G. László, M. Talani, A. Volynkin & S. Yaba Ngouma (1 ♂ ANHRT); same locality as for preceding but 23–24.xi.2024, leg. M. Bashford, I. Elliott & A. Kirk-Spriggs (1 ♂ ANHRT); Odzala-Kokoua National Park, Lobo Research Camp, 390 m, 00°35′04″N, 14°53′12″E, 20–27.ix.2024, leg. M. Bashford, G. László & A. Volynkin (1 ♂ ANHRT); Odzala-Kokoua National Park, Mboko, 379 m, 00°36′16.02″N, 14°53′57.49″E, 19–23.xi.2024, leg. M. Bashford, I. Elliott & A. Kirk-Spriggs (1 ♂ ANHRT); Odzala-Kokoua National Park, Mbomo Headquarters, 570 m, 00°26′13″N, 14°42′01″E, 28.ix–01.x.2024, leg. leg. M. Bashford, G. László & A. Volynkin (12 ♂♂, 1 ♀ ANHRT); Odzala-Kokoua National Park, Moba, 380 m, 00°48′52″N, 15°05′15″E, 11–12.iv.2024, leg. M. Bashford, G. László, M. Talani & S. Yaba Ngouma (1 ♂ ANHRT).

Description. Forewing lengths: male: 18–25 mm, female: 25 mm.

Male (Fig. 57). Upperside. Ground colour of body and wings snowy white. Antennae brown, bipectinate with white scaling along its entire length. Forewing triangular, costal margin almost straight to rounded apex, outer margin arcuate to rounded tornus; hindwing rounded, outer margin curved. Postmedial fascia silvery olive grey, almost straight and crenulate on forewing, curved on hindwing. Submarginal and subterminal fasciae silvery olive grey, crenulate, the former well-defined, the latter less so. Space between submarginal and subterminal fasciae filled with reflective pearly-white spots edged with silvery-grey, between veins; spots most well-defined in posterior half of both wings. Hindwing terminal region irrorated with silvery-grey scales. Underside. As on upperside but wings uniform snowy white, with markings weakly showing through from upperside. Male genitalia (Fig. 66). Uncus medially reduced with pair of long, apically rounded, spatulate lateral processes. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions fused medially, each lobe, elongate with some sparse setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender triangular, tapering to bifurcate apex, the tines pointing dorsad. Saccular section triangular, broader than costal section, tapering into flattened, curved, apically-rounded distal process with small, rounded outer projection at its base. Juxta semi-cylindrical, trapezoidal. Vinculum narrow, V-shaped, bifurcate distally. Saccus very short, rounded at apex. Phallus elongate, coecum pointed, tube tapering at carina towards barbed apex; vesica with weak scobination.

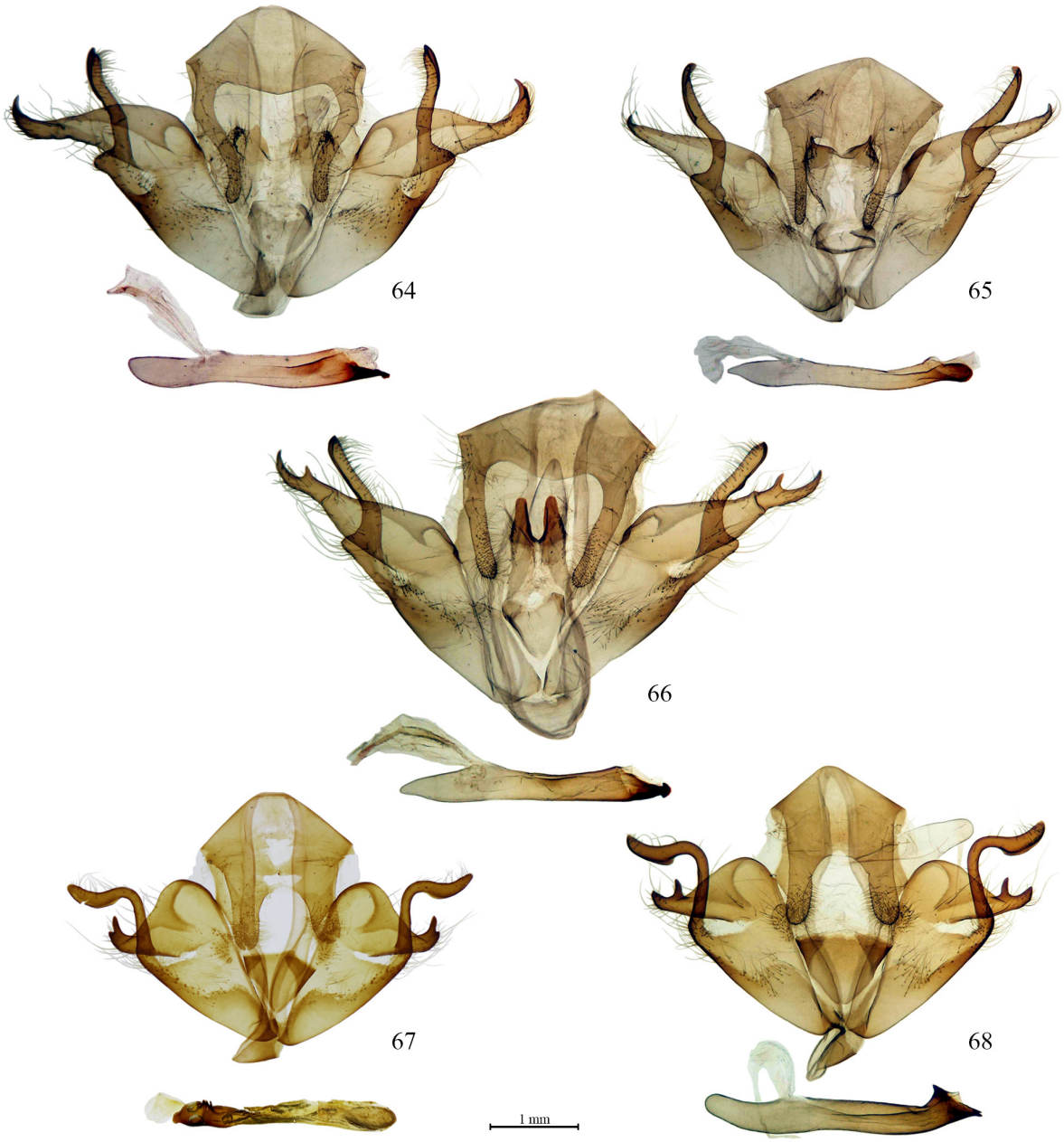
Female (Fig. 58). As in male but rami of antenna shorter; wings elongate and outer margin strongly curved. Pattern weakly expressed.

Diagnosis. The new species is most similar to *S. nivalis* but the wings are more elongate, fasciae are broader and the patterning generally heavier in the latter. In the male genitalia of S. nivalis, the lateral processes of the uncus are incurved (not straight), the mesal protrusions are weakly sclerotised with a small denticle on each lobe (as opposed to elongate, strongly-fused, well-sclerotised lobes), the distal costal process is a simple, single hook (bifurcate in *S. molliae* **sp. n.**) and the distal saccular process is straighter (as opposed to curved outwards). See under *S. laurenae* **sp. n.** for further diagnoses.

Molecular characterisation. Unknown.

Derivatio nominis. It is with great pleasure that this new species is dedicated to Molly Bashford, ANHRT, a member of the team that collected the majority of the type series, and for her organisation of several collaborative projects in sub-Saharan Africa that have yielded many new species including this one. The matronym is a noun in the genitive case.

Distribution. Known from the type series collected at Odzala-Kokoua National Park, it is also presumably found in Cameroon and Gabon (see Basquin & Faravel (2025) as *S. nivalis*) and is likely the Central African sibling of *S. nivalis*.



Figures 64–68. *Streptojana* **gen. n.** species, ♂ genitalia. 64. *S. nivalis* (Rothschild, 1917). 65. *S. laurenae* **sp. n.**, holotype. 66. *S. molliae* **sp. n.**, holotype. 67. *S. brunnea* (Bouyer, 2012). 68. *S. forbesi* **sp. n.**, holotype.

***Smithiana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:EB3AF68E-D182-4B81-951C-9481F7134726

Type species: *Camerunia bipartita* Rothschild, 1917.

Description. Adults. Medium-sized moths. Antenna brown, bipectinate. Ground colour of head and thorax dark chocolate brown; abdomen pale greyish-brown with a medial black dot on each abdominal segment except the eighth. Forewing elongate triangular, rounded at apex and produced at vein M_2, outer margin straight to rounded tornus; R_4 + R_5 stalked. Forewing divided obliquely from apex to near base of anal margin by pale yellowish fascia, the costal section dark chocolate brown and the remainder of

the wing pale greyish brown, darker subapically. Postmedial and subterminal fasciae dark greyish brown, straight, perpendicular to anal margin, indented proximally at anal margin; submarginal fascia dark greyish brown, bilineate, crenulate curved inwardly and almost perpendicular to oblique line. Hindwing subrhomboidal with rounded corners; ground colour fawn, darker toward anal margin. Fringe and anal margin with pale greyish-brown scaling. Postmedial and submarginal fasciae dark greyish brown, strongly curved, the latter slightly crenulate. Subterminal region irrorated with dark grey scales. Underside. Ground colour of body dark brown; forewing fawn, hindwing dark brown. Forewing oblique partition of upperside absent. Postmedia fascia curved inward towards costal margin; submarginal fasciae curved outward towards apex. Subapical and subterminal regions darker. Hindwing with postmedial and submarginal fasciae as on upperside. Female similar to male but rami of the antennae shorter, both wings more elongate, and forewing apex pointed and subapically concave. Male genitalia. Uncus broad at base with long, narrow, gently curved, apically pointed medial projection, and two short (one third of the length), membranous, setose processes, one on each side. Tegumen broad, lateral lobes setose. Mesal protrusions separated, well-developed, setose at apex. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section slender, longer than saccular section. Saccular section triangular, half as wide as costal section, tapering to two long, curved distal processes. Juxta semi-cylindrical with two sharp distal projections. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus slender, coecum rounded and curved dorsad, carina heavily sclerotised and tapered; vesica globular.

Diagnosis. Members of this genus display a unique phenotype within the Janinae with the highly distinctive obliquely split forewing, the costal section being a rich dark chocolate brown and the remainder a pale greyish brown, as well as a single row of spots running along the abdomen in both sexes and cannot readily be confused with any other genus. The male genitalia are also highly distinctive with a long, narrow uncus and two long, sharp distal projections of the juxta.

Derivatio nominis. The compound genus name is derived from Richard Smith, chairman of the board of trustees, ANHRT, after whom a new species is named below, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Members of this genus are restricted to the Guineo-Congolian rainforests.

Species content: two species are assigned to this genus, one of which is a new species.

***Smithiana bipartita* (Rothschild, 1917) comb. n.** (Figs 69–70, 72)
Camarunia [sic] *bipartita* Rothschild 1917b: 491.
Phasicnecus bipartita (Rothschild, 1917): Gaede 1927: 307, pl. 47 (comb.).
Stenoglene bipartita (Rothschild, 1917): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 467; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 8, 19; Basquin & Faravel 2025: pl. 27.

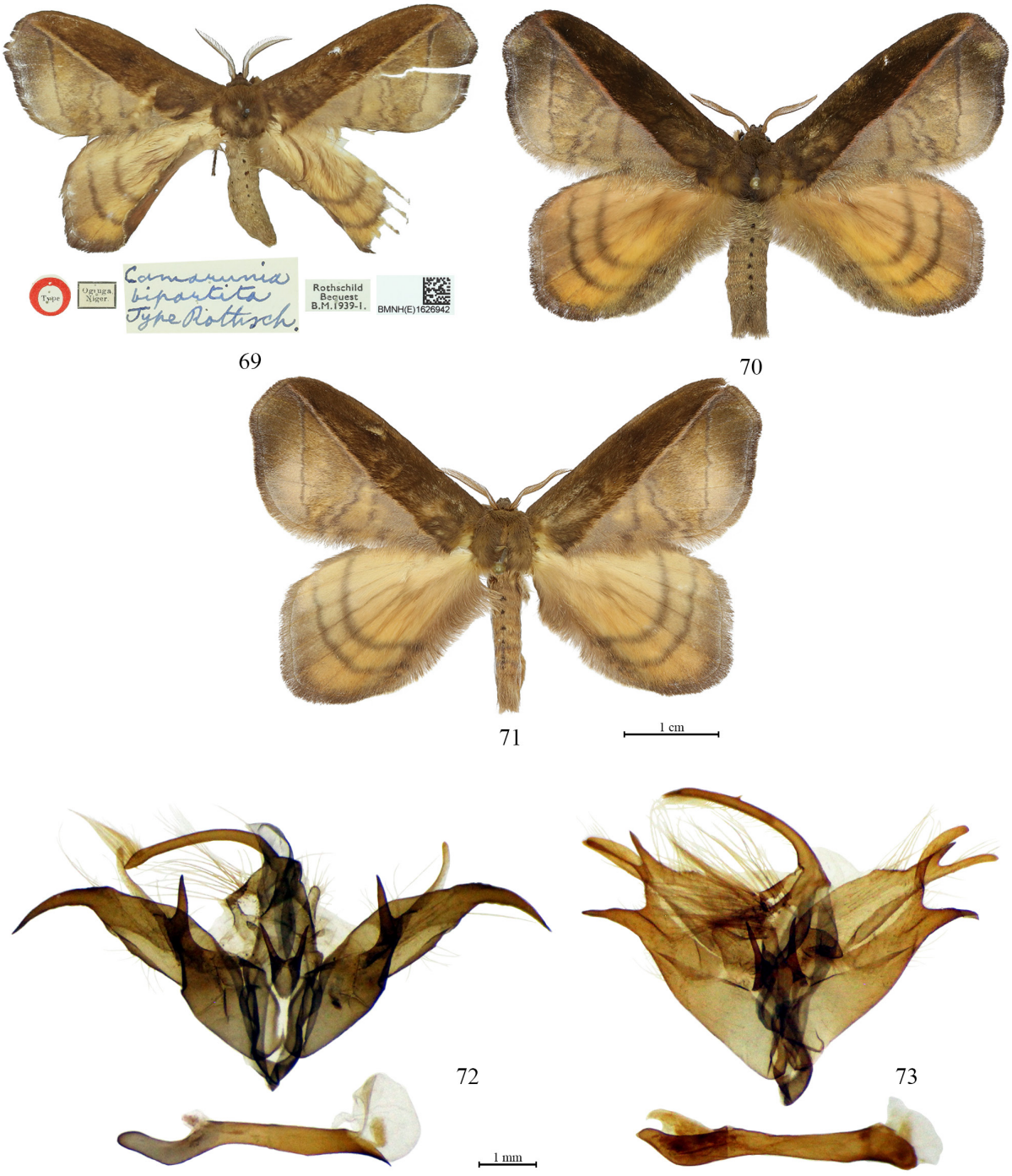
Type locality: [Nigeria] "Ogrugu, Niger" [= Ogurugu].
Name-bearing type material: holotype ♂ in NHMUK.

Description of new species

***Smithiana smithi* sp. n.** (Figs 71, 73)
https://www.zoobank.org/urn:lsid:zoobank.org:act:402B547B-8215-47FE-A5E7-B2C6B4E2F090

Holotype ♂ (Fig. 71):
"LIBERIA 103m | Sinoe County, 6.5km NW | of Jacksonville, Forest near | Solve Problem Village | 5°26'25" N, 9°7'93.9" W | 23-27.i.2018 MV Light Trap | Geiser, M., Sáfián, Sz., | Simonics, G. Leg. | ANHRT:2017.33 || ANHRTUK | 00405675" (ANHRT).

Paratypes (22 ♂♂):
CÔTE D'IVOIRE: Taï National Park, Taï Research Station, 174 m, 05°49′59.8″N, 07°20′32″W, 05–10.vii.2015, leg. M. Aristophanous, P. Moretto & E. Ruzzier (1 ♂ ANHRT); **GHANA**: Ankasa National



Figures 69–73. *Smithiana* **gen. n.** species (69–71 habitus (in ANHRT unless otherwise stated), 72–73 ♂ genitalia). 69. *S. bipartita* (Rothschild, 1917), holotype ♂ of *Camerunia bipartita* Rothschild (NHMUK). 70. *Id.*, ♂, Republic of Congo, Nouabalé-Ndoki NP. 71. *S. smithi* **sp. n.**, holotype ♂. 72. *S. bipartita*. 73. *S. smithi* **sp. n.**, holotype.

Park, 1 km southeast of Nkwanta camp, 103 m, 05°16′29.2″N, 02°38′46.0″W, 26.ii.2006, leg. M. Ochse (1 ♂ RCMO); **GUINEA**: Forêt Classée de Diécké, 27 km S Bounouma, 482 m, 07°27′52.20″N, 08°50′36.60″W, 08.vi.2013, leg. M. Ochse (1 ♂ RCMO); **LIBERIA**: same data as holotype (4 ♂♂ ANHRT); East Nimba Mountains, 3 km NW Geipa, 588 m, 07°27′50.24″N, 08°32′01.46″W, 30.x.2012, leg. M. Ochse (1 ♂ RCMO); East Nimba Mountains, 5 km N of Zortapa, 599 m, 07°24′47.92″N, 08°35′26.28″W, 29 x.2012, leg. M. Ochse (2 ♂♂ RCMO); Gola National Forest, 10 km NE Iseral, 288 m, 07°25′23.86″N, 10°48′42.87″W, 19.x.2012, leg. M. Ochse (1 ♂ RCMO); Gola National Forest, 232 m, 11km NE of Iseral, 07°25′25.9″N, 10°48′32.2″W, 21.x.2012, leg. M. Ochse (2 ♂♂ RCMO); Krahn-

Bassa Reserve, Juboe River, 7.5 km SW Pellokon Town, 140 m, 05°39′04″N, 08°39′04″W, 14–20.i.2018, leg. M. Geiser, S. Sáfián & G. Simonics (2 ♂♂ ANHRT); Nimba Mountains, East Nimba Nature Reserve, Grassfield FDA Field Station, 494 m, 07°29′32.45″N, 08°34′54.05″W, 22.xii.2018, leg. Sáfián & G. Simonics (1 ♂ ANHRT); Wologizi Mountains, Base Camp 2, 883 m, 08°07′20.79″N, 09°56′50.75″W, 22–31.xi.2018, leg. S. Sáfián & G. Simonics (1 ♂ ANHRT); Wologizi Mountains, Base Camp forest, 611 m, 08°07′17″N, 09°57′42″W, 20.xi–01.xii.2017, leg. M. Aristophanous, S. Sáfián, G. Simonics & L. Smith (1 ♂ ANHRT); Wologizi Mountains, Ridge Camp, 865 m, 08°07′10″N, 09°57′11″W, 24–29.xi.2017, leg. M. Aristophanous, S. Sáfián, G. Simonics & L. Smith (3 ♂♂ ANHRT); Wologizi Mountains, Rosewood Camp, 585 m, 08°06′14.9″N, 09°58′27.3″W, 18.xi–01.xii.2018, leg. S. Sáfián & G. Simonics (1 ♂ ANHRT).

Description and diagnosis. Forewing length: 24–28 mm.

Male (Fig. 71). As in the generic description above. Externally indistinguishable from *S. bipartita*, although the brown colouration is a little warmer in the new species. Despite similarities in habitus, the male genitalia (Fig. 73) are highly diagnostic: in the new species, the mesal protrusions are broader, the costal section of the valve is shorter and apically bifurcate (it is longer and terminates in a single, long hook pointed ventrad in *S. bipartita*), and the saccular section is broader (slightly truncate at base of distal processes in the congener). Since these two species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Female. Unknown.

Molecular characterisation. The new species has been assigned the BIN BOLD:AAP9523. Intraspecific PWDs were 0.0% (n=2) and diverged from its sibling *S. bipartita* by 3.8% (n=1).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Richard Smith, chairman of the board of trustees, ANHRT, in admiration of his extraordinary commitment to the field of African entomology and taxonomy through the institution he founded. The patronym is a noun in the genitive case.

Distribution. Known from the Upper Guinean forests of West Africa where it is likely to be endemic.

***Lokiana* gen. n.**

https://www.zoobank.org/urn:lsid:zoobank.org:act:8A01B156-AAB7-4F43-9110-B723B5B70D14

Type species: *Jana sigyna* Aurivillius, 1893.

Description. Adults. Medium-sized moths. Antenna bipectinate. Ground colour pale brown or beige. Wing fasciae dark brown or greyish-brown. Forewing elongate triangular with curved outer margin; R_4 + R_5 stalked. Discal marking chevron- or crescent-shaped. Postmedial and submarginal fasciae curved, irregularly crenulate, strongly displaced proximally in space M_2 and between veins Cu_2 and anal margin. Submarginal fascia highlighted on outer edge by silvery-grey spots and crenules ringed in dark brown, with the most noticeable being a large rounded marking near costa. Subapical and subterminal area dark brown or greyish brown, bordered on its inner edge by oblique, dark brown, subterminal fascia arising at apex and terminating near submarginal fascia. Fringe dark brown. Hindwing ovoid, outer margin evenly arcuate to produced tornus. Postmedial and submarginal fasciae curved, crenulate or sinuate, forming dark markings at termination points along anal margin; outer edge of submarginal fasciae near anal margin with silvery-grey markings as on forewing. Tornal area darker. Anal margin with dark markings at tornus and at termination points of postmedial/submarginal fasciae. Underside. Ground colour as on upperside. Wing markings reduced to poorly defined discal marking, arcuate, crenulate or sinuate, postmedial and submarginal fasciae, and dark subapical area as on upperside. Females generally larger, antenna rami shorter, forewing apex distinctly produced and hindwing with prominent produced tornus. Male genitalia. Uncus produced into elongate medial structure tridentate at apex and a pair of bifurcate lateral processes. Tegumen broad. Mesal protrusions separated, ovoid, setose. Valve elongate triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section rhomboidal, terminating in produced distal process with bifurcate apex. Saccular section triangular, as broad as costal section, tapering towards and terminating in produced, rounded, apically bifurcate,

recurved, plate. Juxta semi-cylindrical, inverse-triangular with V-shaped emargination distally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded, carina heavily sclerotised and tapered; vesica with small patch of scobination.

Diagnosis. Members of *Lokiana* **gen. n.** share some phenotypic similarities with *Cryptojana* **gen. n.** species but in the latter the postmedial fascia of the forewing is curved and does not follow the same irregular and displaced course as the submarginal fascia. The postmedial and submarginal fasciae are similar in *Neojana* **gen. n.** but neither (on both wings) are as heavily sinuate compared to *Lokiana*, and a forewing discal spot is always present in the latter, while the forewing is more uniform in the former (the outer region demarcated by the subterminal fascia in *Lokiana* is noticeably darker). The male genitalia of *Lokiana* species are very characteristic: the uncus is produced into a medial section with a tridentate apex and a pair of bifurcate lateral processes which in *Cryptojana* is either produced into a single long medial process bifurcate at apex or a pair of triangular medio-lateral processes, and a single pair of hooked lateral projections while in *Neojana* is medially reduced with a pair of lateral processes. Furthermore, the distal saccular process in *Lokiana* terminates in a bifurcated rounded plate which is always long and blade-like in *Cryptojana* and *Neojana*.

Derivatio nominis. The compound genus name is derived from Loki, a god in Norse mythology who is married to Sigyn presumably after whom Aurivillius named the type species of the genus, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Members of this genus are restricted to Guineo-Congolian rainforests.

Species content: two species are assigned to this genus, one of which is a new species.

***Lokiana sigyna* (Aurivillius, 1893) comb. n.** (Figs 74–75, 78)

Jana sigyna Aurivillius 1893: 209.

Phasicnecus sigyna (Aurivillius, 1893): Aurivillius 1901: 29, pl. 4, fig. 2 (comb.); Gaede 1927: 307, pl. 46.

Stenoglene sigyna (Aurivillius, 1893): Forbes 1955: 129 (comb.); Dall'Asta & Poncin 1980: 467; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 19; Basquin & Faravel 2025: pl. 26.

Phiala pseudatomaria auct. nec Strand, 1911: Vári *et al*. 2002: 150.

Type locality: "Camerun interior" [probably Barombi station (see Takano 2025)].
Name-bearing type material: holotype ♂ in MfN.

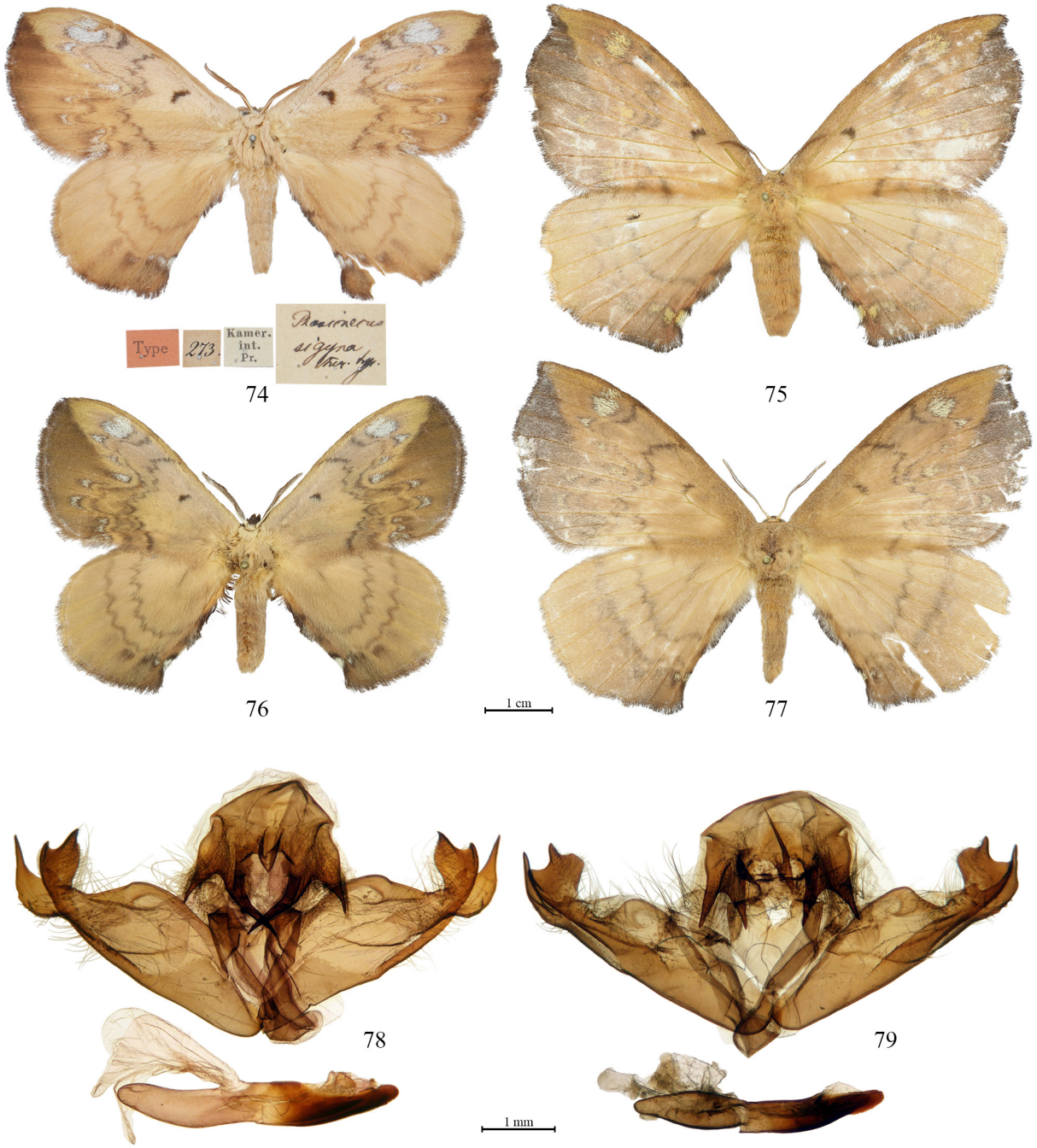
Description of new species

***Lokiana lydiae* sp. n.** (Figs 76–77, 79)
https://www.zoobank.org/urn:lsid:zoobank.org:act:58312BA4-E579-4461-9F85-E79A7531A410

Holotype ♂ (Fig. 76):
"IVORY COAST 60m | Parc National d'Azagny, entrée | Sonaye (Secondary forest) | 05°14'32"N, 04°48'05"W | 25-28.xi.2021 Act. Light Trap | Moretto, P., Mulvaney, L., | Takano, H. Leg. | ANHRT:2021.8 || ANHRTUK | 00405666" (ANHRT).

Paratypes (21 ♂♂, 2 ♀♀):
CÔTE D'IVOIRE: same data as holotype (3 ♂♂, 1 ♀ ANHRT); Taï National Park, Taï Research Station (SRET), 174 m, 05°50′00″N, 07°20′32″W, 25.iii–17.iv.2017, leg. A. Aristophanous, M. Aristophanous, M. Geiser & P. Moretto (1 ♂ ANHRT); **LIBERIA**: Grassfield, Nimba, ix–x.1963, leg. A. Forbes-Watson (1 ♂ NHMUK); Wetezu camp, Wonegizi Nature Reserve, 551 m, 08°04′57.11″N, 09°34′47.86″W, 19–27.iii.2019, leg. S. Safian & S. Koivogui (1 ♀ ANHRT); Wologizi Mts, Ridge Camp, 865 m, 08°07′10″N, 09°57′11″W, 24–29.xi.2017, leg. M. Aristophanous, S. Safian, G. Simonics & L. Mulvaney (4 ♂♂ ANHRT); same data as for preceding but Base Camp forest, 611 m, 08°07′17″N, 09°57′42″W, 20.xi–01.xii.2017 (6 ♂♂ ANHRT); same locality as for preceding but Ridge Camp 2, 883

m, 08°07′20.79″N, 09°56′50.75″W, 22–30.xi.2018, leg. S. Safian & G. Simonics (5 ♂♂ ANHRT); same data as for preceding but Rosewood Camp, 585 m, 08°06′14.9″N, 09°58′27.3″W, 18.xi–01.xii.2018 (1 ♂ ANHRT).



Figures 74–79. *Lokiana* **gen. n.** species (74–77 habitus (in ANHRT unless otherwise stated), 78–79 ♂ genitalia). 74. *L. sigyna* (Aurivillius, 1893), holotype ♂ of *Phasicnecus sigyna* Aurivillius (MfN). 75. *Id*., ♀. 76. *L. lydiae* **sp. n.**, holotype ♂. 77. *Id*., paratype ♀. 78. *L. sigyna*. 79. *L. lydiae* **sp. n.**, holotype.

Description and diagnosis. Forewing lengths: male: 32–38 mm, female: 40–42 mm.

Male (Fig. 76) and female (Fig. 77). As in the generic description above. Externally indistinguishable from *L. sigyna*, although the new species is perhaps not as boldly marked. Despite similarities in habitus, the male genitalia (Fig. 79) are highly diagnostic: in the new species, the central point of the tridentate apex of the medial section of the uncus is longer and more pointed, the bifurcate triangular lateral processes of the uncus are approximately the same size (the inner one is twice as long as the outer in *L. sigyna*), the distal section of the costa is weakly produced (it is strongly produced into

a rounded, plate-like apex in *L. sigyna*) and the dorsal projection of the distal saccular process is more rounded (triangular and pointed in *L. sigyna*). Since these two species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Molecular characterisation. The new species has been assigned the BIN BOLD:AAO6229. Intraspecific PWDs were 0.2–0.6% (n=4) and diverged from its sibling species by 4.1–5.0% (n=6).

Derivatio nominis. It is with great pleasure that this new species is dedicated to Lydia Mulvaney, ANHRT, a Lepidoptera and Neuroptera taxonomist as well as a seasoned fieldworker, who was part of the team that collected the holotype. The matronym is a noun in the genitive case.

Distribution. This species is restricted to the Upper Guinean forests of West Africa.

***Nepenthojana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:B9A29BEC-49E4-406B-9AB1-C4BB2540F55B

Type species: *Stenoglene uniformis* Dall'Asta & Poncin, 1980.

Description. Adults. Small moths. Ground colour brown. Antenna bipectinate. Forewing triangular with rounded apex, gently arcuate outer margin; $R_4 + R_5$ stalked. Hindwing outer margin arcuate and slightly angled at vein M_2. Discal marking chevron- or crescent-shaped, weakly defined. Postmedial fascia dark brown, curved, slightly irregular, crenulate or indicated by crescents. Area beyond fascia darker brown. Underside as on upperside but paler. Female similar to male but rami of antenna shorter and wing markings poorly-defined. Male genitalia. Uncus medially reduced with pair of apically pointed or twisted and truncate processes. Tegumen broad. Mesal protrusions separated, trapezoid, setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate, medially constricted and terminating in a dorso-distal pointed process. Saccular section elongate ovoid, slightly broader than costal section, with elongate distal process as long as sacculus, truncate at apex terminating with hooked projection, slightly longer than costal process. Juxta semi-cylindrical, trapezoidal, distally emarginate and with median rounded concavity. Vinculum narrow, V-shaped, bifurcate distally. Saccus very short, rounded at apex. Phallus curved, coecum bulbous, curved medially towards plate-like carina with serrate margin; vesica globular with small, dense patch of scobination.

Diagnosis. This genus comprises two plain, externally indistinguishable species that are highly distinct within the Janinae. There are no other genera that contain species with just a simple postmedial fascia on both wings in both sexes. The male genitalia are unique with the hooked projection at the apex of the distal saccular process, the medial concavity of the juxta, the serrated margin of the plate-like carina and dense scobination of the vesica.

Derivatio nominis. The compound genus name is derived from the carnivorous plant *Nepenthes* Linnaeus, 1753, in reference to the shape of the phallus which strongly resembles the pitcher, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Restricted to the equatorial rainforests of Central Africa from Cameroon and Gabon to D.R. Congo.

Species content: two species are assigned to this genus.

***Nepenthojana uniformis* (Dall'Asta & Poncin, 1980) comb. n.** (Figs 80, 83)
Stenoglene uniformis Dall'Asta & Poncin 1980: 467, figs 23–24, photo 2, fig. 19; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 22; Basquin & Faravel 2025: pl. 39.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

***Nepenthojana plenevauxae* (Bouyer, 2012) comb. n.**
Stenoglene plenevauxae Bouyer 2012: 10, pl. 1, fig. 2, pl. 3, fig. 21; Kitching & Rougerie *et al*. 2018: suppl. 1.

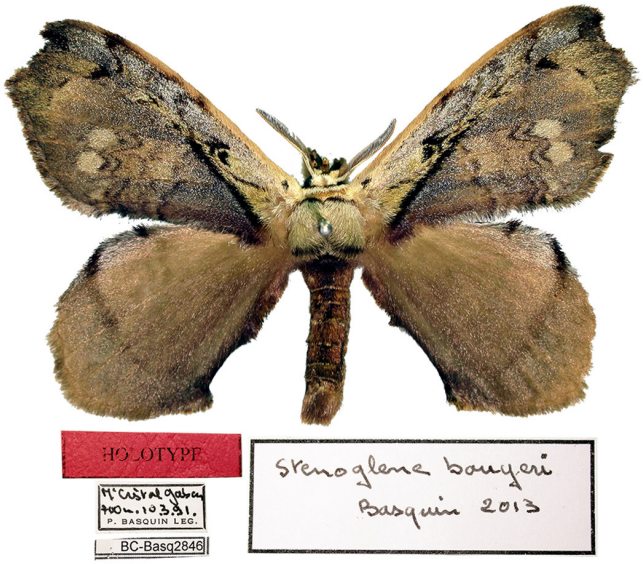
Type locality: "Cameroun, r. Edea-Douala pk 20".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).



80



81



82

1 cm

Figures 80–82. Janinae species, habitus (in ANHRT unless otherwise stated). 80. *Nepenthojana* **gen. n.** *uniformis* (Dall'Asta & Poncin, 1980), ♂, Republic of Congo, Nouabalé-Ndoki NP. 81. *Neojana* **gen. n.** *pseudosygina* (Bouyer & Basquin, 2012), ♂, Republic of Congo, Odzala-Kokoua NP. 82. *Atopojana* **gen. n.** *bouyeri* (Basquin, 2013), holotype ♂ of *Stenoglene bouyeri* Basquin (RCPB).

***Neojana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:CAED1E30-8069-4457-A2AF-5C5C7A8BADB8

Type species: *Stenoglene pseudosygina* Bouyer & Basquin, 2012.

Description. Adults. Medium-sized moths. Antenna bipectinate, covered in creamy-white scales. Ground colour of body and forewings pale brown or beige to rich mahogany; hindwing beige to brown. Wing fasciae dark brown or greyish-brown. Forewing elongate triangular with curved outer margin; R_4

+ R_5 stalked. Costal region, especially basally, irrorated with paler scales. Postmedial and submarginal fasciae curved, in the form of two chevrons or crescents joined along vein M_2. Submarginal fascia less well-defined and slightly interrupted, highlighted on outer edge by patches of silvery-grey scales, with a distinct spot near costa. Subapical and subterminal area darker irregularly irrorated in patches of silvery-grey scales, bordered on its inner edge by oblique, subterminal fascia arising at apex and terminating near submarginal fascia. Fringe dark brown. Hindwing ovoid, outer margin evenly arcuate to gently produced tornus. Postmedial and submarginal fasciae gently curved and slightly angled inwards near anal margin, the former more weakly-defined often disappearing towards costa. Tornal area darker. Anal margin with dark markings at tornus and irrorated throughout with silvery-grey scales. Underside. Ground colour uniform pale brown or beige; forewing margin darker brown. Wing markings limited to weakly-expressed dark brown curved postmedial and submarginal fasciae. The female is unknown. Male genitalia. Uncus medially reduced with pair of apically rounded lateral processes. Tegumen broad. Mesal protrusions separated, each lobe well-sclerotised, angled and weakly setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate, apically bifurcate, claw-like and terminating in two short points. Saccular section triangular, twice as broad basally as costal section, terminating in apically truncate, distal process half the length of sacculus. Juxta semi-cylindrical, trapezoidal, weakly tridentate distally. Vinculum narrow, U-shaped. Saccus short, rounded at apex. Phallus straight, coecum rounded, carina tapered; vesica with some scobination.

Diagnosis. The sole member of this genus is a distinctive moth but at first glance, may be confused with *Lokiana* **gen. n.** and *Cryptojana* **gen. n.** species (see under diagnosis section of the former for detailed comparisons). The colours and patterns are somewhat reminiscent of *Cryptojana* species but the forewing postmedial fascia is always curved and never displaced (like in *Neojana* **gen. n.**) and lack the curved postmedial and submarginal hindwing fasciae. In the male genitalia, the uncus of *Cryptojana* is either produced into a single long medial process bifurcate at apex or a pair of triangular medio-lateral processes, and a single pair of hooked lateral projections while in *Neojana* it is medially reduced with only a pair of lateral processes. Furthermore, in the barcode analyses, *Neojana* formed a well-differentiated and isolated clade and combined with the aforementioned morphological characters, is deemed a distinct genus.

Derivatio nominis. The compound genus name is derived from the Ancient Greek for 'new' (νέος), and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. The single species is known only from the refugial rainforests of Cameroon, Gabon and eastern Republic of Congo.

Species content: monotypic.

***Neojana pseudosygina* (Bouyer & Basquin, 2012) comb. n.** (Figs 81, 84)
Stenoglene pseudosygina Bouyer & Basquin, *In*: Bouyer 2012: 7, pl. 3, fig. 22; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 20; Basquin & Faravel 2025: pl. 37.

Type locality: "Cameroun, r. Edea-Douala pk 20".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Atopojana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:A89FD585-9E75-4049-B7BF-9EAEDD3EF1D4

Type species: *Stenoglene bouyeri* Basquin, 2013.

Description. Adults. Small moths. Ground colour of head creamy-white, thorax olive beige, abdomen brown. Antenna bipectinate, covered in creamy-white scales. Forewing elongate triangular, costa gently arcuate, apex pointed and subapically concave; outer margin produced and strongly angled at vein M_2, almost straight to rounded tornus; R_4 and R_5 not stalked. Forewing divided obliquely from sub-apex to near base of anal margin by gently arcuate fascia, highlighted in black along basal half; the ground colour



Figures 83–85. Janinae species, ♂ genitalia (in ANHRT unless otherwise stated). 83. *Nepenthojana* **gen. n.** *uniformis* (Dall'Asta & Poncin, 1980). 84. *Neojana* **gen. n.** *pseudosygina* (Bouyer & Basquin, 2012). 85. *Atopojana* **gen. n.** *bouyeri* (Basquin, 2013), Cameroon, Korup NP (RCMO).

of costal section basally olive beige, becoming a darker silvery-grey towards apex, and the remainder of the wing dark brown. Discal spot and discocellular marking black. Postmedial and submarginal fasciae charcoal, bilineate, irregular running almost perpendicular to costa, the former slightly broader and more diffuse, the latter more sinuate. Two olive spots present beyond submarginal fascia between veins M_3 and Cu_2. Subterminal fascia black, from spot at apex to tornus, displaced sharply proximad along vein R_5; region between subterminal fascia and margin irrorated with black scales. Fringe olive brown irrorated throughout with black scales. Hindwing subrhomboidal with rounded corners; anal margin with concavity near produced tornus; ground colour brown. Costal margin irrorated with silvery-grey scales and tufts of black scales at starting points of black postmedial and submarginal fasciae. Postmedial fascia gently arcuate, weakly-defined and disappearing towards anal margin. Submarginal fascia curved, diffuse, weaker towards anal margin before terminating in black marking along margin. Fringe olive brown irrorated infrequently with darker scaling. Underside. Ground colour of body beige, wings uniform greyish-brown lacking markings. Forewing costal and terminal regions darker. Hindwing postmedial and submarginal fasciae as on upperside but interrupted and indicated by dashes and

chevrons. The female is unknown. Male genitalia. Uncus produced into short truncate structure, weakly emarginate medially. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, ovoid, densely covered in setae. Valve triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular to pointed apex; margin weakly sinuose. Saccular section triangular, twice as wide as costal section at base, tapering to short curved and apically pointed distal process, one third the length of sacculus. Juxta semi-cylindrical, inverse-triangular, with two long apically-pointed and setose ensiform dorso-lateral projections, each as long as valve costa. Vinculum narrow, V-shaped. Saccus rounded at apex. Phallus slender, coecum rounded, slightly curved at entrance of ductus ejaculatoris, carina tapered.

Diagnosis. This peculiar genus is somewhat reminiscent of *Smithiana* **gen. n.** but the outer margin of the forewing is clearly produced at vein M_2 and angled in *Atopojana* **gen. n.** (rounded in *Smithiana*) and a concavity is present along the anal margin giving the impression of a prominent tornus. Recovered as sister to the phenotypically differing *Hecateana* **gen. n.** in every analyses, they do share the two prominent oval markings between veins M_3 and Cu_2 of the forewing which are not present in any other genera. The male genitalia display a highly distinctive juxta which easily distinguishes this genus from any other: the dorso-lateral projections of the juxta are as long as the valve costa and although the shape is not dissimilar in *Smithiana bipartita*, it is nearly an eighth of the length of the valve in the latter.

Derivatio nominis. The compound genus name is derived from the Ancient Greek for 'unusual' (ἄτοπος), in reference to its highly distinctive and unique phenotype, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Known only from the rainforests of Cameroon and Gabon.

Species content: monotypic.

***Atopojana bouyeri* (Basquin, 2013) comb. n.** (Figs 82, 85)
Stenoglene bouyeri Basquin 2013: 23, pl. 5, fig. 1; Basquin 2024: 8, 19; Basquin & Faravel 2025: pl. 35.

Type locality: "Gabon, Mont de Cristal".
Name-bearing type material: holotype ♂ in RCPB.

***Hecateana* gen. n.**
https://www.zoobank.org/urn:lsid:zoobank.org:act:FD72DD2B-3C05-44CE-B283-E0F2EC53A3F3

Type species: *Stenoglene paulisi* Dall'Asta & Poncin, 1980.

Description. Adults. Medium-sized moths. Antenna bipectinate, rami shorter in females. Ground colour of body and wings cream. Wing fasciae dark purplish-brown to dark greyish-brown. Forewing elongate triangular, rounded at apex; outer margin rounded, slightly angled between veins M_1 and M_2 in some species; termen crenulate; $R_4 + R_5$ stalked. Postmedial and submarginal fasciae curved, the former absent in some species. Two paler ovoid spots often visible in submarginal area between veins M_3 and Cu_2. Hindwing ovoid, outer margin rounded, termen weakly sinuate. Postmedial and submarginal fasciae only visible on anal half. Postmedial fasciae gently curved, indicated by dashes along veins. Submarginal fascia bilineate, curved, the inner zigzagged, the outer indicated by spots. Anal margin with dark markings at tornus and at termination points of fasciae. Underside. Ground colour as on upperside but uniform lacking all markings. Wing markings generally more weakly defined in females. Male genitalia. Uncus medially reduced with long lateral process; a mediolateral process is present in some species. Tegumen broad. Mesal protrusions separated, ovoid or trapezoid, densely covered in setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, terminating in distal process gently curved dorsad. Saccular section triangular, tapering to blade-like distal process. Juxta semi-cylindrical, trapezoid. Vinculum narrow, V-

shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded, carina heavily sclerotised and tapered.

Diagnosis. Members of this genus are highly distinctive moths with a cream ground-colour and dark fasciae on both wings. It may be confused with paler members of *Stenoglene sensu str*. but the hindwings lack all fasciae in the latter. *Hecateana* **gen. n.** is also somewhat reminiscent of *Cryptojana* **gen. n.** but in the male genitalia of the latter, the lateral processes of the uncus are always shorter than the medial processes, and the costal section of the valves lacks a distal process, while the costal margin is strongly lobed giving it the appearance of antlers.

Derivatio nominis. The compound genus name is derived from the Greek goddess Hecate who was closely associated with Diana and Artemis in the Roman era, and the Roman goddess Jana (or Diana) after whom the genus and subfamily of Eupterotidae is likely named. The gender is feminine.

Distribution. Members of this genus are distributed in the Guineo-Congolian forests with an isolated population in the Usambara Mountains.

Species content: four species are assigned to this genus, three of which are new species.

***Hecateana paulisi* (Dall'Asta & Poncin, 1980) comb. n.** (Figs 86, 90)
Stenoglene paulisi Dall'Asta & Poncin 1980: 479, figs 27–28, photo 1, fig. 11; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 21; Basquin & Faravel 2025: pl. 36.

Type locality: [D.R. Congo] "Uele, Paulis".
Name-bearing type material: holotype ♂ in RMCA.

Description of new species

***Hecateana dargei* sp. n.** (Figs 88, 92)
https://www.zoobank.org/urn:lsid:zoobank.org:act:CEAD9FB8-F3F4-4F3F-AC15-6901F42C1B05

Holotype ♂ (Fig. 88):
"TANZANIE : Kagera Re- | gion, KIYAMYULWA, 1157 | m.,02°08.628'S.,031°36.642' | E.,26-III-2010 (Ph. Darge) || Ex. Coll. Ph. Darge | ANHRT:2019.20 || ANHRTUK | 00276411" (ANHRT).

Description. Forewing length: male: 26 mm.

Male (Fig. 88). Ground colour of body and wings cream; vertex of head dark red. Antenna bipectinate, brown. Wing fasciae dark brown; veins highlighted on outer half of wings. Forewing elongate triangular, rounded at apex; outer margin rounded but slightly angled between veins M_1 and M_2; termen crenulate. Discocellular marking crescent shaped. Postmedial fascia evenly curved, strongly dentate. Submarginal fascia bilineate, curved but irregularly sinuate, the outer of the two indicated by spots and chevrons; two paler ovoid spots on outer edge between veins M_3 and Cu_2. Subapical and subterminal area irrorated with greyish brown scales, bordered on its inner edge by oblique, irregular, subterminal fascia arising at apex and terminating near submarginal fascia. Fringe greyish-brown turning dark cream towards tornus. Hindwing ovoid, outer margin rounded, termen weakly sinuate. Fasciae only visible on anal half. Postmedial fasciae gently curved, indicated by dashes along veins. Submarginal fascia bilineate, curved, the inner zigzagged and weakly-defined, the outer indicated by spots. Anal margin with three dark markings, one at tornus and at termination points of fasciae. Fringe cream. Underside. Ground colour as on upperside but uniform lacking all markings. Male genitalia (Fig. 92). Uncus medially reduced with two pairs of long, apically setose processes, a medio-lateral pair pointed at apex and a lateral pair spatulate at apex, the former slightly shorter than the latter. Tegumen broad. Mesal protrusions separated, ovoid densely covered in setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, terminating in distal process gently curved dorsad. Saccular section triangular, half as wide as costal section, tapering to blade-like distal process slightly shorter than sacculus. Juxta semi-cylindrical, trapezoid, and slightly

larger than mesal protrusions. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded, carina heavily sclerotised and tapered.

Female. Unknown.

Diagnosis. Externally indistinguishable from *H. paulisi* and *H. kitchingi* **sp. n.** although the new species is smaller and perhaps not as boldly marked. Despite similarities in habitus, the male genitalia are distinctive. Unlike its congeners, the medio-lateral process of the uncus is slightly shorter than the lateral process in *H. dargei* **sp. n.** and there is no suture between the spine and the process, i.e. it is one continuous pointed process.

Molecular characterisation. Unknown.

Derivatio nominis. It is with great pleasure that this new species is dedicated to the late Philippe Darge whose rich collection of Eupterotidae now in the ANHRT has shed new light on the systematics of these moths. The patronym is a noun in the genitive case.

Distribution. Known only from its type locality on the western shores of Lake Victoria.

***Hecateana giustii* sp. n.** (Figs 89, 93)
https://www.zoobank.org/urn:lsid:zoobank.org:act:DF0697F5-8E6D-43AB-B578-E517612B90A2

Holotype ♂ (Fig. 89):
"Afrique Orientale | Usambara,Pangani | L. Conradt | 1891 || Ex Oberthür Coll. | Brit. Mus. 1927–3. || NHMUK 010929820 [QR code]" (NHMUK).

Description. Forewing length: 25 mm.

Male (Fig. 89). Upperside. Ground colour of body and wings creamy white. Antenna brown, bipectinate, rami long. Forewing elongate triangular, costal margin gently arcuate to apex, outer margin rounded and crenulate, tornus rounded; hindwing outer margin rounded and crenulate. Wings lack patterning except irregular weakly defined dark brown submarginal fascia, indicated by spots between veins; spots on hindwing fascia becoming better defined towards anal margin and terminate in spot along margin. A second spot, half as small, is placed equidistant between it and the base. Underside. Ground colour of body slightly darker; wings uniform creamy-white with the patterning just discernible from upperside. Male genitalia (Fig. 93). Uncus medially reduced with pair of long apically pointed and setose lateral processes. Tegumen broad. Mesal protrusions weakly fused mesally, trapezoid, covered in setae. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, terminating in broad hook-like distal process gently curved dorsad; left process with subapical tooth. Saccular section triangular, half as wide as costal section, tapering to broad distal process hooked at apex. Juxta semi-cylindrical, inverse-triangular, and not much larger than mesal protrusions. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded, carina heavily sclerotised and tapered.

Female. Unknown.

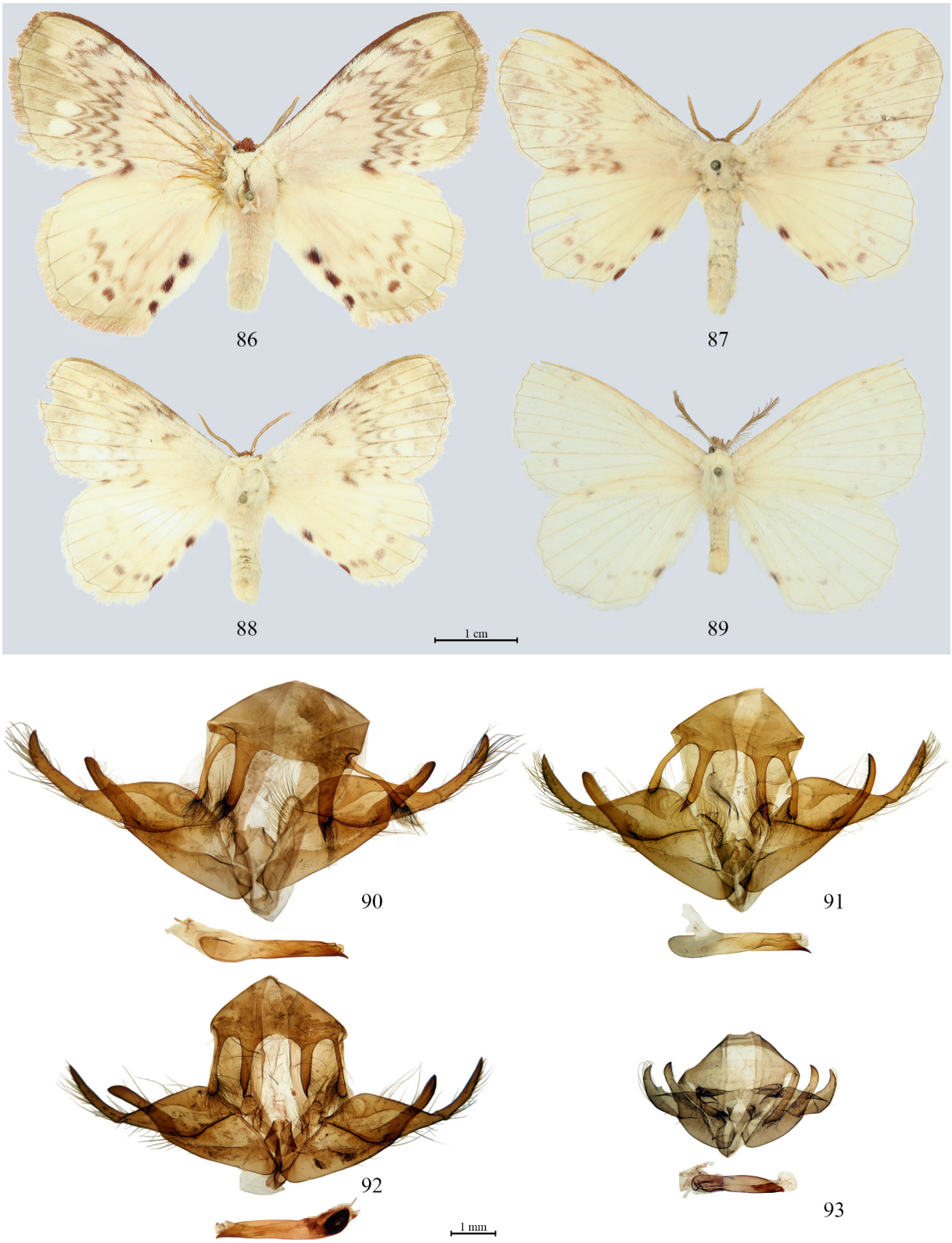
Diagnosis. The new species is most similar to *H. paulisi*, *H. dargei* **sp. n.** and *H. kitchingi* **sp. n.** sharing the crenulate wing margins and general ground colour but the forewing lacks all markings except for an irregular and weakly defined submarginal fascia. The holotype is not a worn or rubbed specimen and it seems this species is weakly marked. In the genitalia, the new species differs in the smaller size (nearly half the size of its congeners), the absence of the medial processes of the uncus and the broader, more robust processes of the costal and saccular sections of the valve.

Derivatio nominis. It is with great pleasure that the new species is dedicated to Alessandro Giusti, Senior Curator of Lepidoptera, NHMUK, without whose kind assistance this project could not have been completed. The patronym is a noun in the genitive case.

Distribution. This species is known only from the type locality of "Usambara, Pangani" in northeast Tanzania. The true locality is likely to be in the East Usambara Mountains, and refers either to the region around Derema where Leopold Conradt was based from May to December 1891 (Kolbe 1892) or the foothills of the mountains between the Derema and the Pangani River.

Notes. The new species was standing under *Pterocerota virginea* Hampson, 1905 in the NHMUK collections but based on wing shape and patterning, it does not belong in *Pterocerota* and is tentatively assigned to *Hecateana* **gen. n.** However, despite phenotypic similarities to *H. paulisi*, the

type species, there are noticeable differences particular in the genitalia and further investigations are required to establish whether they are congeneric.



Figures 86–93. *Hecateana* **gen. n.** species (86–89 habitus (in ANHRT unless otherwise stated), 90–93 ♂ genitalia). 86. *H. paulisi* (Dall'Asta & Poncin, 1980), ♂, Republic of Congo, Odzala-Kokoua NP. 87. *H. kitchingi* **sp. n.**, holotype ♂. 88. *H. dargei* **sp. n.** holotype ♂. 89. *H. giustii* **sp. n.**, holotype ♂ (NHMUK). 90. *H. paulisi*. 91. *H. kitchingi* **sp. n.**, holotype. 92. *H. dargei* **sp. n.** holotype. 93. *H. giustii* **sp. n.**, holotype

***Hecateana kitchingi* sp. n.** (Figs 87, 91)
https://www.zoobank.org/urn:lsid:zoobank.org:act:1355924B-0E38-491B-9883-AB44CAA69FBE

Holotype ♂ (Fig. 87):
"IVORY COAST, 174m | Taï NP, Taï Research Station (SRET), | 05°50'00"N, 07°20'32.0"W, | 25.III - 17.IV .2017, MV light, | Aristophanous,A., Aristophanous,M., | Geiser,M., Moretto,P., leg., | ANHRT:2017.25 || ANHRTUK | 00005444" (ANHRT).

Description and diagnosis. Forewing length: male: 28 mm.

Male (Fig. 87). As in the description of *H. dargei* **sp. n.** above. Externally indistinguishable from *H. paulisi* and *H. dargei* although the forewing of *H. kitchingi* **sp. n.** is perhaps most angular between veins M_1 and M_2. The male genitalia of *H. kitchingi* (Fig. 91) is most similar to *H. paulisi* but differs in the narrower medio-lateral process of the uncus and its sharp spike-like tip which is as wide as the process at its base in *H. kitchingi*, while it is half the width at its base in *H. paulisi* (also clearly visible in the figure of the holotype in Dall'Asta & Poncin (1980: Fig. 27)).

Molecular characterisation. The new species has been assigned the BIN BOLD:AA05978. The holotype diverged from Congolese *H. paulisi* specimens in BIN BOLD:AAM0172 by 3.2–4.1% (n=3).

Derivatio nominis. It is with great pleasure that the new species is dedicated to Ian Kitching, NHMUK, in gratitude for his kind and insightful discussions regarding Bombycoidea taxonomy over many years. The patronym is a noun in the genitive case.

Distribution. The new species is restricted to the Upper Guinean forests of West Africa.

***Epijana* Holland, 1893**

Epijana Holland 1893: 514; Aurivillius 1901: 21, 27; Gaede 1927: 306; Forbes 1955: 96, 129; Dall'Asta & Poncin 1980: 464; Fletcher & Nye 1982: 59; Nässig & Oberprieler 2008: 56, 71; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type species: *Epijana lanosa* Holland, 1893, by original designation.

Redescription. Adults. Medium- to large-sized moths. Antenna bipectinate in males, rami shorter in females. Ground colour ranging from a rich yellow through to dark brown. Forewing outer margin arcuate, apex produced in females of some species; R_4 and R_5 not stalked. Postmedial fascia dark, straight in some species, curved in others. Dark basal and discal markings present in some species. Hindwing uniform except for a straight oblique postmedial fascia in some species. Male genitalia. Uncus produced into elongate medial structure, bifurcate or trifurcate; some species with pair of lateral projections. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions setose, either fused medially, large and angled or separated and ovoid. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, bifurcate at apex or terminating in incurved hook. Saccular section triangular, shorter than costal section, tapering to long distal process shorter than sacculus. Juxta semi-cylindrical, inverse-triangular. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus slender, coecum rounded, slightly curved at entrance of ductus ejaculatoris, carina heavily sclerotised and tapered; vesica with a small patch of scobination.

Diagnosis. The genus *Epijana* has never been satisfactorily delimited and it is only through the integration of genetic and morphological characters that this has been achieved. The genus is easily diagnosed based on the combination of the curved or oblique forewing fascia and the unstalked forewing veins R4 and R5, the configuration of the veins shared only with the phenotypically different *Pterocerota* and *Urojana*. The male genitalia of the five species assigned to this genus differ considerably, especially the type species, and based solely on this character, the generic boundary would be difficult to define. It is thus unclear what Dall'Asta & Poncin (1980) were referring to when they stated that the only way to separate the two genera was through examination of the genitalia as no explanation was provided as to how they differ. Similarly, in the original description of the most recent *Epijana* species to have been

described, *E. vingerhoedti* Bouyer, 2012, no explanation was given as to why this taxon was placed in this genus (and not *Stenoglene*) except to say that it "seems to belong" in *Epijana* (Bouyer 2012: 4).

Distribution. Members of this genus are widespread in forest and forest-savanna mosaics from Sierra Leone to Uganda.

Species content: five species are assigned to this genus.

***Epijana cosima* (Plötz, 1880)** (Figs 94, 100)
Jana cosima Plötz 1880: 85; Kirby 1892: 801.
Epijana cosima (Plötz, 1880): Aurivillius 1901: 28, fig. 18 (comb.); Gaede 1927: 306, pl. 46; Forbes 1955: 96, figs 13, 40, 41; Dall'Asta & Poncin 1980: 464, photo 1, fig. 3; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 23; Basquin & Faravel 2025: pl. 43.

Type locality: [Cameroon] "Abo".
Name-bearing type material: holotype ♀ in ZIMG? (not examined).

Epijana lanosa Holland, 1893
Epijana lanosa Holland 1893: 514; Aurivillius 1901: 28 (syn.).

Type locality: [Gabon] "Kangwé" and [Equatorial Guinea] "Benita".
Name-bearing type material: syntypes (♂ and ♀) in CMNH (not examined), one ♂ syntype in NHMUK.

Stenoglene velutonia Druce, 1896
Stenoglene velutonia Druce 1896: 355;
Epijana velutonia (Druce, 1896): Aurivillius 1901: 28 (comb. & syn.).

Type locality: "Cameroons".
Name-bearing type material: holotype ♂ in NHMUK.

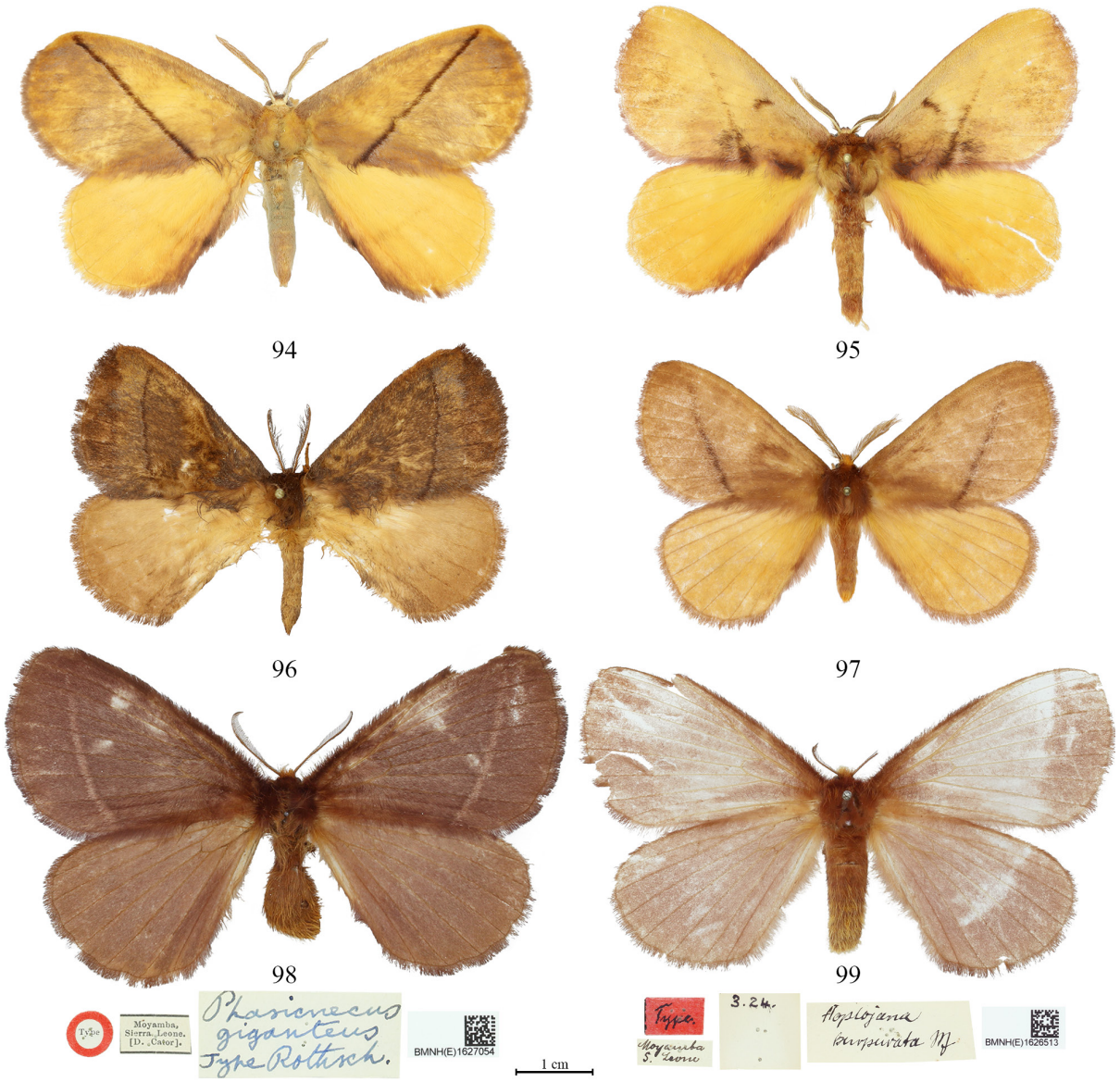
***Epijana gigantea* (Rothschild, 1917) comb. n.** (Figs 97–99, 102)
Phasicnecus giganteus Rothschild 1917b: 491; Gaede 1927: 307.
Stenoglene gigantea (Rothschild, 1917): Forbes 1955: 95, 129, fig. 1 (comb.).
Stenoglene giganteus (Rothschild, 1917): Dall'Asta & Poncin 1980: 484, photo 2, figs 16, 17; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 22.

Type locality: "Moyamba, Sierra Leone".
Name-bearing type material: 2 ♀ syntypes in NHMUK. The syntype figured (Fig. 96) is from the Rothschild Bequest, and the second specimen referred to in the original description as being in NHMUK is one from Sierra Leone presented to the museum by Messrs Watkins & Doncaster.

Hoplojana purpurata Wichgraf, 1921 **syn. n.** (Fig. 99)
Hoplojana purpurata Wichgraf 1921: 196; Gaede 1927: 305.
Stenoglene purpurata (Wichgraf, 1921): Kitching & Rougerie *et al*. 2018: suppl. 1 (comb.).

Type locality: "Moyamba (Sierra Leone)".
Name-bearing type material: holotype ♀ in NHMUK.

Notes. Based on the identical habitus and type locality of *Hoplojana purpurata* **syn. n.** is synonymised with *E. gigantea*. Excluded from *Hoplojana* Aurivillius, 1902 by Bouyer (2011), Kitching & Rougerie *et al*. (2018) assigned *H. purpurata* to *Stenoglene* and listed *Hoplojana zernyi* Gschwandner, 1923 as a junior synonym of it, but these two taxa are certainly not conspecific and based on the wingspan quoted in the original description, the latter likely belongs in *Jana* Herrich-Schäffer, [1854] as suggested by Gaede (1927).



Figures 94–99. *Epijana* Holland, 1893 species, habitus (in ANHRT unless otherwise stated). 94. *E. cosima* (Plötz, 1880), ♂, Cameroon, Nkongsamba. 95. *E. meridionalis* (Dall'Asta & Poncin, 1980), ♂, Zambia, Lukwakwa. 96. *E. vingerhoedti* Bouyer, 2012, ♂, Rwanda, Nyungwe. 97. *E. gigantea* (Rothschild, 1917), ♂, Sierra Leone, Gori Hills. 98. *Id*., syntype ♀ of *Phasicnecus giganteus* Rothschild (NHMUK). 99. *Id*., holotype ♀ of *Hoplojana purpurata* Wichgraf, 1921 (NHMUK).

***Epijana latifasciata* Dall'Asta & Poncin, 1980**
Epijana latifasciata Dall'Asta & Poncin 1980: 464, figs 1–2, photo 1, fig. 1.
Epijana latifascia [sic] Dall'Asta & Poncin, 1980: Kitching & Rougerie *et al*. 2018: suppl. 1.

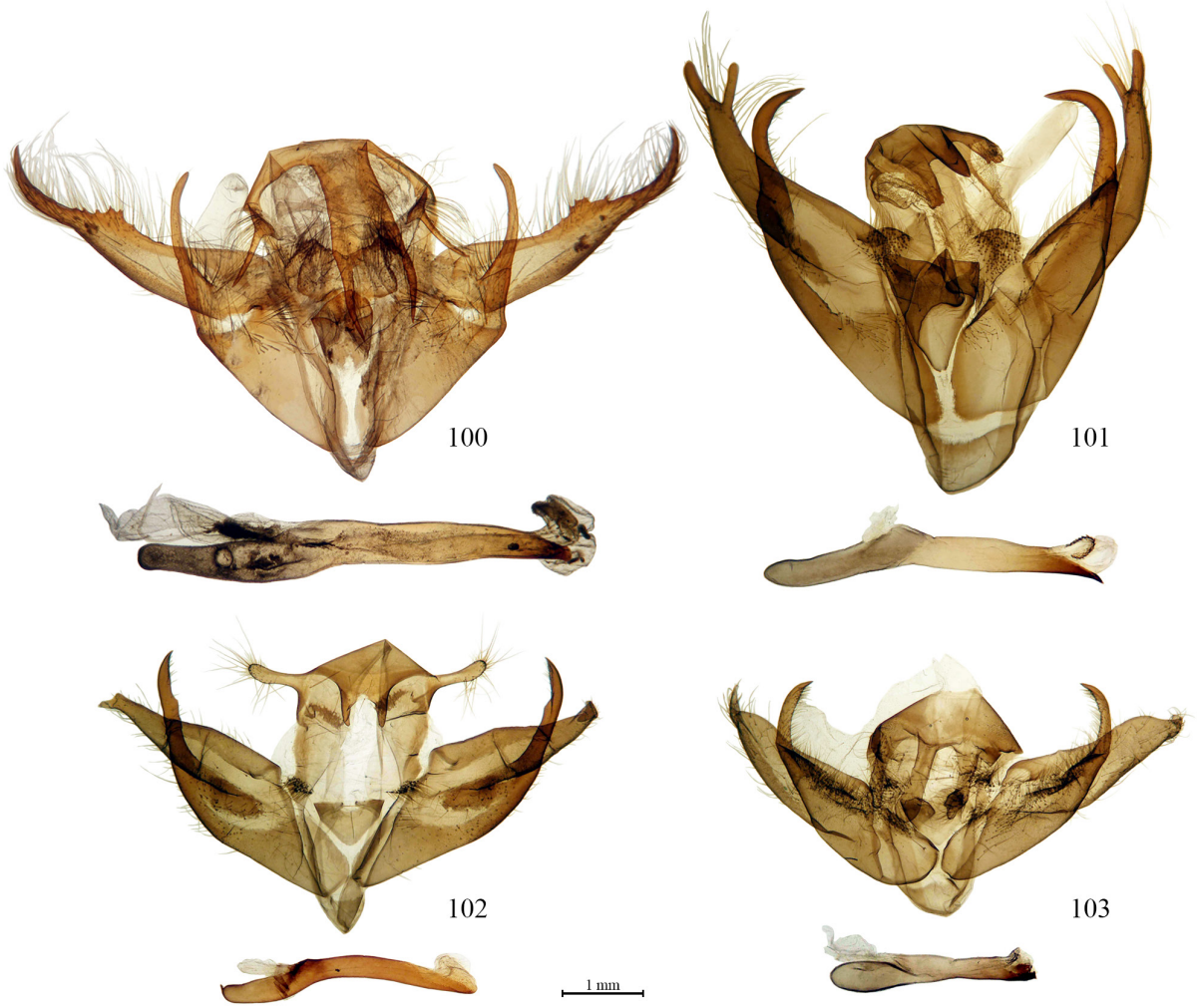
Type locality: [D.R. Congo] "Equateur, Flandria".
Name-bearing type material: holotype ♂ in RMCA.

***Epijana meridionalis* Dall'Asta & Poncin, 1980** (Figs 95, 101)
Epijana meridionalis Dall'Asta & Poncin 1980: 466, figs 3–4, photo 1, figs 2, 4; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 23; Basquin & Faravel 2025: pl. 42.

Type locality: [D.R. Congo] "Elisabethville" [= Lubumbashi].
Name-bearing type material: holotype ♂ in RMCA.

***Epijana vingerhoedti* Bouyer, 2012** (Figs 96, 103)
Epijana vingerhoedti Bouyer 2012: 4, pl. 3, fig. 19; Kitching & Rougerie *et al*. 2018: suppl. 1.
Stenoglene giganteus? auct. (nec Rothschild): Basquin 2024: 22.

Type locality: "Rwanda, P.N. Nyungwe, Pindura".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).



Figures 100–103. *Epijana* Holland, 1893 species, ♂ genitalia. 100. *E. cosima* (Plötz, 1880). 101. *E. meridionalis* (Dall'Asta & Poncin, 1980). 102. *E. gigantea* (Rothschild, 1917). 103. *E. vingerhoedti* Bouyer, 2012.

***Drepanojana* Aurivillius, 1893**
Drepanojana Aurivillius 1893: 212; Aurivillius 1901: 21, 30; Rothschild 1917a: 466; Gaede 1927: 309; Forbes 1955: 97, 129; Fletcher & Nye 1982: 54; Nässig & Oberprieler 2008: 56, 71; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type species: *Drepanojana fasciata* Aurivillius, 1893, by monotypy.

Redescription. Adults. Small moths. Antenna bipectinate, rami shorter in females. Ground colour brown. Male. Forewing triangular, costa gently arcuate subapically, outer margin straight, rounded at tornal angle; R_4 + R_5 stalked. Apical marking cream-coloured, small, with curved inner margin and straight outer margin, entirely absent in some individuals (Fig. 105). Hindwing almost triangular. Female. Forewing with arcuate costal margin to a well-produced apex and arcuate outer margin; broad subapical band gently curved, creamy white; one or two cream apical spots present. Hindwing triangular, outer margin very gently arcuate in males forming an acute angle with anal margin at tornus, while in

females the wings are broader with an arcuate margin. Male genitalia. Uncus medially reduced with pair of single lateral processes. Tegumen broad. Gnathos reduced membranous, weakly sclerotised. Mesal protrusions separated, ovoid, setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section rhomboidal, tapering towards recurved distal process. Saccular section triangular, tapering to long, sigmoidal distal process, with short projection on inner edge at the first angle. Juxta semi-cylindrical, trapezoidal, emarginate distally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus elongate, coecum rounded, produced at entrance of ductus ejaculatorius, tapering towards plate-like carina; vesica globular.

Diagnosis. The wing shape and markings of both sexes render this genus a highly distinctive one and cannot readily be confused with any other Eupterotidae. The females share phenotypic similarities with members of the butterfly genus *Bicyclus* Kirby, 1871. The male genitalia are similar to the phenotypically dissimilar *Streptojana* **gen. n.** (see under generic diagnosis section of the latter for further detailed comparisons).

Distribution. Members of this genus are restricted to Guineo-Congolian rainforests.

Species content: three species assigned to the genus, one of which is a new species.

***Drepanojana fasciata* Aurivillius, 1893** (Figs 104–106, 110)
Drepanojana fasciata Aurivillius 1893: 212; Aurivillius 1901: 30, fig. 21, pl. 5, fig. 2; Rothschild 1917a: 466; Gaede 1927: 309, pl. 47; Kitching & Rougerie *et al*. 2018: suppl. 1; Basquin 2024: 23; Basquin & Faravel 2025: pl. 41.

Type locality: “Sierra Leone”.
Name-bearing type material: holotype ♀ in MfN.

Drepanojana apicalis Aurivillius, 1915
Drepanojana apicalis Aurivillius 1915: 9; Rothschild 1917a: 466 (syn.).

Type locality: [Ghana] “Gold Coast, Bibianaha”.
Name-bearing type material: holotype ♂ in NHMUK.

***Drepanojana maeonia* (Druce, 1896) stat. rev.** (Figs 107–108, 111)
Melanothris [sic] *maeonia* Druce 1896: 353.
Drepanojana maeonia (Druce, 1896): Aurivillius 1901: 31 (comb.).
Drepanojana fasciata auct. (nec Aurivillius): Rothschild 1917a: 466 (syn.); Gaede 1927: 309; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: [Nigeria] “Old Calabar”.
Name-bearing type material: holotype ♀ in NHMUK.

Notes. Based on barcode divergences and differing genitalia, D. *maeonia* is revived from synonymy with *D. fasciata*.

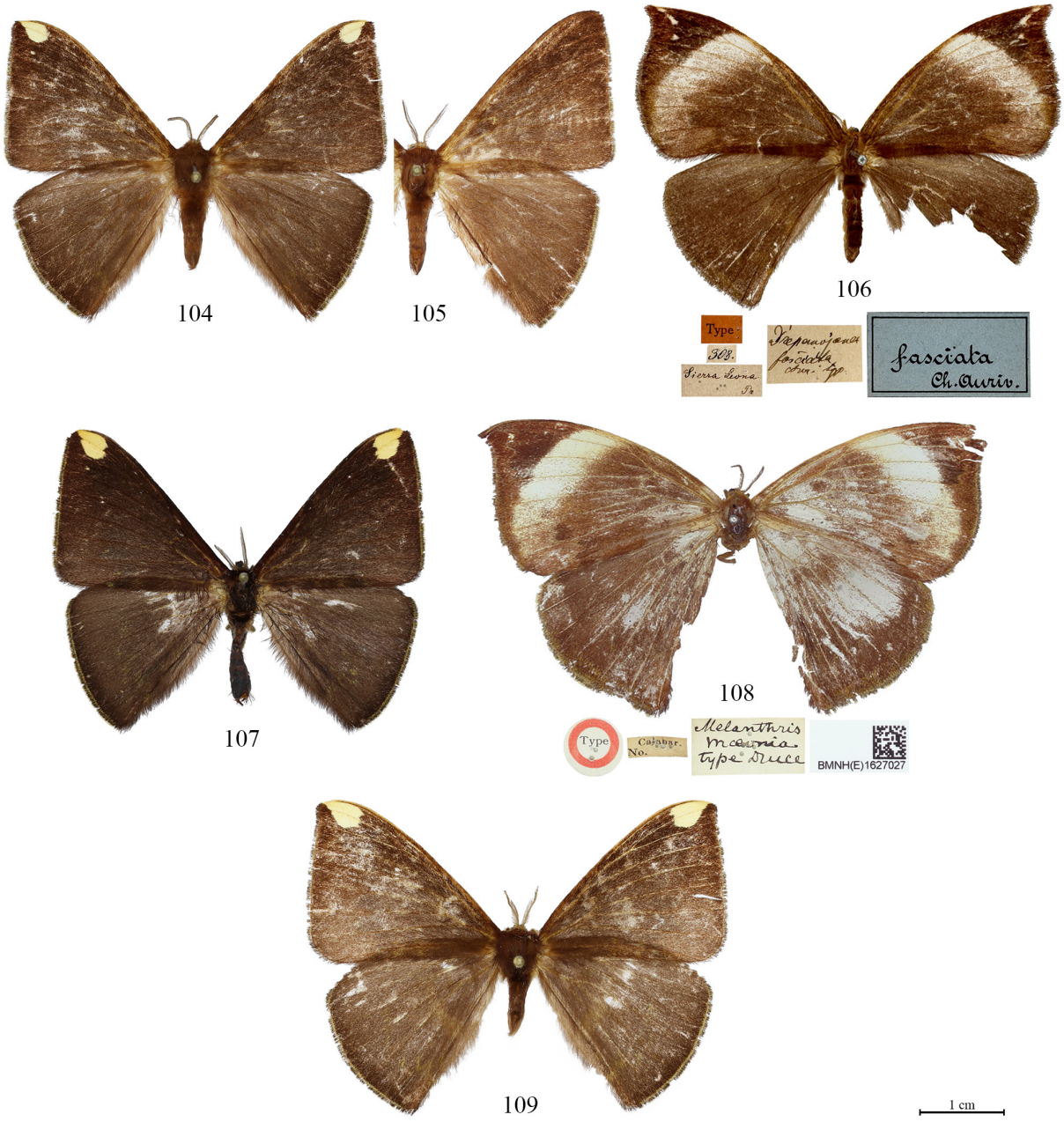
Description of new species

***Drepanojana hogani* sp. n.** (Figs 109, 112)
https://www.zoobank.org/urn:lsid:zoobank.org:act:F5999921-8954-4641-92E2-A455CFE481A8

Holotype ♂ (Fig. 109):
“TANZANIE : Kagera Re- | gion, Minziro Forest, Kere | Hills, 01°06.087′S.,031°31. | 168′E., 1176 m., 12-V-2011 | (Ph. Darge) || Ex. Coll. Ph. Darge | ANHRT:2019.20 || ANHRTUK | 00363959” (ANHRT).

Paratypes (5 ♂♂):

TANZANIA: near Kakindu Hill, 01°10′S, 31°30′E, 09.vii.1915, leg. G.D. Hale Carpenter (1 ♂ OUMNH); Minziro Forest, Kabale, 1156 m, 01°01.792′S, 31°34.446′E, 30.x.2011, leg. P. Darge (1 ♂ ANHRT); Minziro Forest Reserve, near Kakindo, 1149 m, 01°08.563′S, 31°30.516′E, 03.ix.2014, leg. P. Darge (3 ♂♂ ANHRT).



Figures 104–109. *Drepanojana* Aurivillius, 1893 species, habitus (in ANHRT unless otherwise stated). 104–105. *D. fasciata* Aurivillius, 1893, ♂, Liberia, Wologizi Mts. 106. *Id*., holotype ♀ (MfN). 107. *D. maeonia* (Druce, 1896), ♂, Republic of Congo, Odzala-Kokoua NP. 108. *Id*., holotype ♀ of *Melanothris* [sic] *maeonia* Druce (NHMUK). 109. *D. hogani* **sp. n.**, holotype ♂.

Description and diagnosis. Forewing length: 25 mm.

Male (Fig. 109). See generic description above. Phenotypically identical to its congener, but the new species possesses a slightly broader forewing with a larger apical spot. The ground plan of the genitalia (Fig. 112) is similar to its congeners but the apex of the valve costa is shorter (two-thirds the length of its congeners), less curved and terminates in a single denticle (rather than two in the others). Additionally, compared to *D. fasciata*, the processes of the uncus are shorter and the apical third of the

saccular process is thinner and twice as long, while compared to *D. maeonia*, the short projection along the inner edge of the distal saccular process is at the angle (rather than ventrad of it) and the apical two-thirds are almost right-angled. Since these species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Female. Unknown.

Molecular characterisation. Unknown.

Derivatio nominis. It is with great pleasure that the new species is dedicated to James Hogan, Collections Manager of Lepidoptera, OUMNH, who has kindly facilitated the author's numerous visits over the years to the collections under his care. The patronym is a noun in the genitive case.

Distribution. This new species seems to be restricted to the unique forests of the Sango Bay-Minziro complex in south-western Uganda and north-western Tanzania, an area of high floral and faunal diversity (Luke *et al*. 2024).



Figures 110–112. *Drepanojana* Aurivillius, 1893 species, ♂ genitalia (all in ANHRT). 110. *D. fasciata* Aurivillius, 1893. 111. *D. maeonia* (Druce, 1896). 112. *D. hogani* **sp. n.**, holotype.

Urojana Gaede, 1915

Urojana Gaede 1915: 73; 1927: 306; Forbes 1955: 97, 129; Fletcher & Nye 1982: 168; Nässig & Oberprieler 2008: 58, 71; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type species: *Urojana eborea* Gaede, 1915, by original designation.

Redescription. Adults. Medium-sized moths. Antenna bipectinate, rami shorter in females. Ground colour creamy-beige. Male. Forewing triangular, costa arcuate, apex rounded with slight subapical concavity; outer margin gently sinuate, strongly angled at vein M_2, and rounded at tornus; R_4 and R_5 not stalked. Discal marking dark brown, subrectangular. Postmedial fascia olive brown, arcuate, strongly angled inwards near costa. Submarginal fascia olive brown, sinuate, with several markings on outer edge: a ferruginous triangular marking edged dorsally with black at costa and several crescent shaped dark brown markings towards anal margin. Fringe colour the same as general ground colour with the exception of short black section at apex. Hindwing rhomboidal, outer margin produced and angled at

M_2. Postmedial and submarginal fasciae olive brown, curved, disappearing towards costa. Submarginal fascia with several crescent shaped dark brown or black markings edged distally with pale triangular markings towards anal margin. Terminal area at tornus heavily irrorated with dark brown scales. Anal margin with dark brown or black markings at termination points of fasciae. Underside. Ground colour as above. Wings uniformly coloured except for submarginal fascia indicated by dark brown spots; submarginal fascia on forewing arises at costal ferruginous triangular marking near apex. Female. Similar to male but wings broader. Forewing with purple hue in costal half; apical concavity of outer margin large. Forewing discal mark absent and fasciae of all wings indistinct. Male genitalia. Uncus produced medially into trilobed structure, the two lateral lobes wider than the dorsal one. Tegumen broad. Gnathos reduced, membranous, weakly sclerotised. Mesal protrusions separated, trapezoid, setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section elongate triangular, tapering towards bifurcate apex, the ventral process three times the length of digitate dorsal process; another shorter process is positioned proximad of dorsal process. Saccular section triangular, slightly broader than costal section, tapering towards bifurcate apex, the dorsal process five or six times longer than ventral process. Juxta semi-cylindrical, trapezoidal, distally emarginate. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus robust, coecum rounded, slightly curved at entrance of ductus ejaculatoris, carina heavily sclerotised and tapered; vesica with a small patch of scobination.

Diagnosis. Another highly unique and distinctive genus which can easily be identified based on the angular hindwing outer margin, beige colouration and pattern. The male is vaguely reminiscent of forest geometrids such as *Chorodnodes* Warren, 1897.

Distribution. Members of this genus are restricted to Guineo-Congolian rainforests.

Notes. One of only a few Janinae genera where the female was hitherto unknown (Basquin 2024), a recently collected barcode-confirmed specimen of *U. eborea* is illustrated in Fig. 115.

Species content: two species are assigned to the genus, one of which is a new species.

***Urojana eborea* Gaede, 1915** (Figs 113–115, 117)
Urojana eborea Gaede 1915: 73; Gaede 1927: 307, pl. 45; Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: “Elfenbeinküste” [= Côte d’Ivoire].
Name-bearing type material: holotype ♂ in MfN.

Description of new species

***Urojana basquini* sp. n.** (Figs 116, 118)
https://www.zoobank.org/urn:lsid:zoobank.org:act:25B65FB7-0F0D-4CAA-A76D-3E579F49E73E
Urojana eborea auct. nec Gaede: Basquin 2024: 24; Basquin & Faravel 2025: pl. 44.

Holotype ♂ (Fig. 116):
“Monts de Cristal | GABON-750m 16/x/93 | leg. P. Basquin [black border] || BC-Basq3064 [black border]” (RCPB).

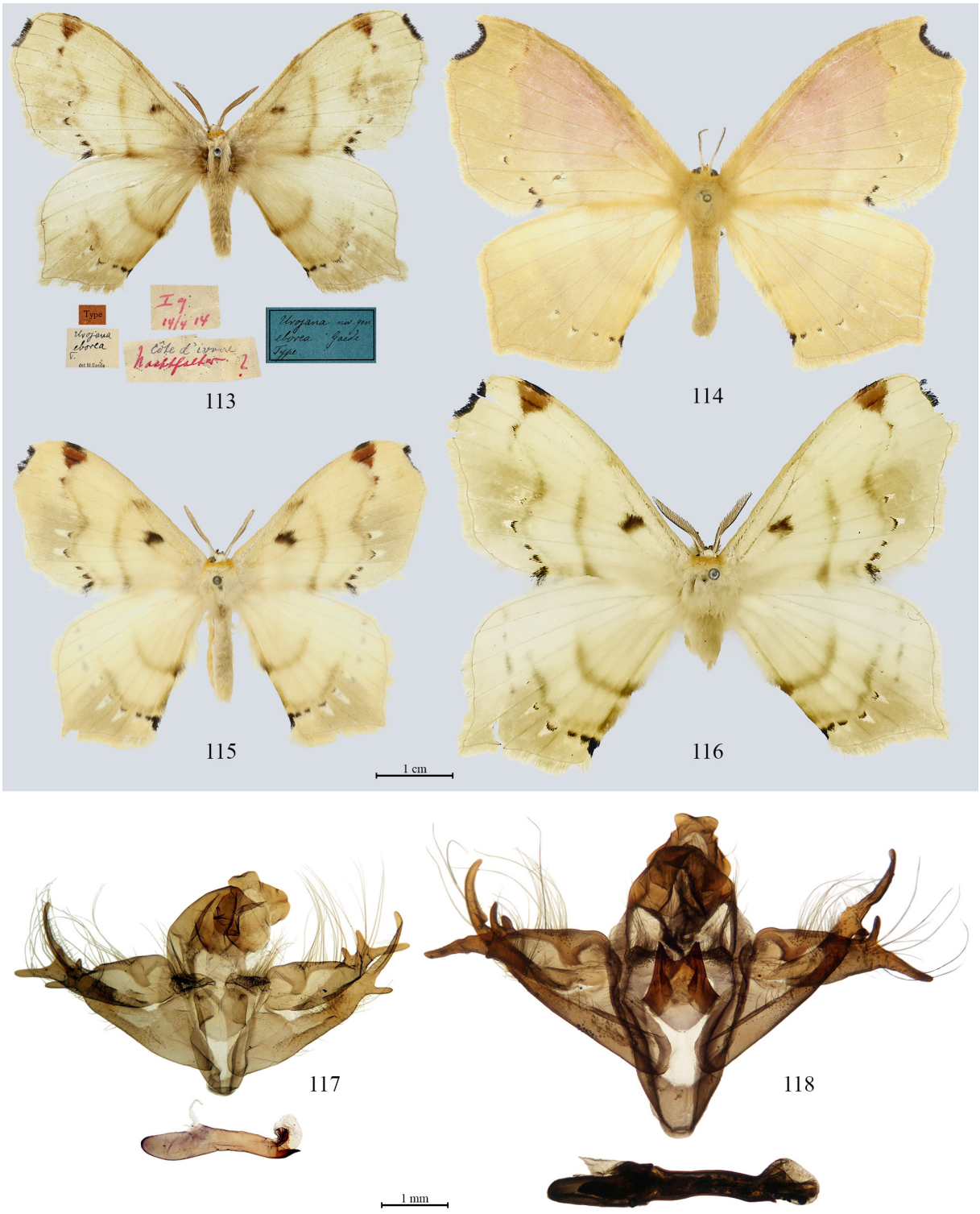
Paratypes (4 ♂♂):
GABON: same data as holotype but 700 m (1 ♂ RCPB); same locality as holotype but 10.xi.1991 (1 ♂ RCPB); same locality as holotype but 24.x.1992 (1 ♂ RCPB); Monts de Cristal, Kinguélé, 13.iv.1991, leg. P. Basquin (1 ♂ RCPB).

Description and diagnosis. Forewing length: 32–37 mm.
Male (Fig. 116). See generic description above. Phenotypically identical to its congener, but the new species is larger. The ground plan of the male genitalia (Fig. 118) is similar to *U. eborea* but the capsule is larger and with several distinctions: in the new species, the medial process of the uncus is dilated apically, the ventral process of the bifurcate saccular section of the valve is shorter, the juxta is twice as

long, distally bifurcate and the phallus is twice as long. Since these two species are found in allopatry, there is unlikely to be any confusion in specimens with good provenance.

Female. Unknown.

Molecular characterisation. The new species has been assigned the BIN BOLD:AAO9504. Intraspecific PWDs were 0.0% (n=2) and diverged from *U. eborea* by 1.5 % (n=2).



Figures 113–118. *Urojana* Gaede, 1915 species (113–116 habitus (in ANHRT unless otherwise stated), 117–118 ♂ genitalia). 113. *U. eborea* Gaede, 1915, holotype ♂ (MfN). 114. *Id*., ♀, Liberia, Wologizi Mts. 115. *Id*., ♂. 116. *U. basquini* **sp. n.**, holotype ♂ (RCPB). 117. *U. eborea*. 118. *U. basquini* **sp. n.**, holotype.

Derivatio nominis. It is with great pleasure that the new species is dedicated to Patrick Basquin, for his kindness in facilitating the author's numerous visits to his collection and whose research into the Lepidoptera fauna of Gabon has led to the discovery of many new species including this one. The patronym is a noun in the genitive case.

Distribution. This new species is known only from the type locality in northern Gabon.

***Pterocerota* Hampson, 1905**

Pterocerota Hampson 1905: 389; Janse 1917: 66; 1937: 4; Hering 1926: 124; Fletcher 1979: 178; Minet 1990: 353; 1994: 78; Lemaire & Minet 1998: 331; Vári *et al*. 2002: 125; Nässig & Oberprieler 2008: 57, 71; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138.

Type species: *Pterocerota virginea* Hampson, 1905, by monotypy.

Redescription. Adults. Small moths. Antenna bipectinate, rami long in males and shorter in females. Ground colour creamy- or pearly-white. Wings diaphanous. Forewing triangular, apex rounded; R_4 and R_5 not stalked. Hindwing rounded. Postmedial and submarginal fasciae grey or charcoal, curved, diffuse and weakly-defined sometimes indicated by spots. Underside as on upperside but weakly marked and uniform. Male genitalia. Uncus with pair of medial lobes and lateral digitate processes. Tegumen broad. Gnathos reduced, membranous. Mesal protrusions separated, miniscule and setose. Valve broadly triangular, divided into well-defined costal and saccular sections, fused at their base. Costal section triangular or ovoid, tapering towards pointed or truncate apex. Saccular section triangular, twice as broad as costal section at base, tapering to robust, pointed or hooked distal process half the length of sacculus. Juxta trapezoidal, semi-cylindrical, bilobed distally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus elongate or robust, coecum rounded and tapering towards carina.

Diagnosis. This genus is phenotypically most similar to the white *Streptojana* **gen. n.** species from which it can be instantly distinguished by the forewing venation. Veins R_4 + R_5 are stalked in *Streptojana* whilst they are unstalked in *Pterocerota*. The male genitalia of *Pterocerota* are highly diagnostic with a pair of medial lobes and digitate lateral lobes of the uncus which is unique in this group of Janinae.

Distribution. Members of this genus are found in lowland east and south-eastern Africa, seemingly in small disjunct populations.

Notes. Originally described in the family Pterothysanidae by Hampson (1905), *Pterocerota* was treated as belonging to this family (Janse 1917, Tessmann 1921, Hering 1926, Fletcher 1979) or the subfamily Pterothysaninae of Callidulidae (Janse 1937) until it was transferred to the Eupterotidae by Minet (1990). According to manuscript notes seen by Kitching (1984), R.H. Carcasson had earlier believed *Pterocerota* to be an eupterotid. In a later work, Minet (1994) specifically assigned it to the subfamily Janinae, and near *Stenoglene* and *Epijana*, based on wing venation similarities.

Species content: three species are assigned to this genus, one of which is a new species.

***Pterocerota perissinottoi* (Bouyer, 2012) comb. n.** (Figs 120, 124)

Stenoglene perissinottoi Bouyer 2012: 8, pl. 1, fig. 2, pl. 2, fig. 12; Kitching & Rougerie et al. 2018: suppl. 1.

Pterocerota virginea auct. nec Hampson: Takano & László 2022: 13, figs 48, 49; Basquin 2024: 23.

Type locality: "South Africa, False Bay".
Name-bearing type material: holotype ♂ in ZSM (photograph examined).

***Pterocerota virginea* Hampson, 1905** (Figs 119, 123)

Pterocerota virginea Hampson 1905: 389; Janse 1917: 66; 1937: 5, pl. 33, figs 1, 2; Hering 1926: 124; Fletcher 1979: 178; Vári *et al*. 2002: 125; Kitching & Rougerie *et al*. 2018: suppl. 1; Krüger 2020: 138.

Type locality: [South Africa] “Natal”.
Name-bearing type material: holotype ♂ in NHMUK.

Notes. There appears to be some discrepancies regarding the origin and type locality of this particularly uncommon species. Hampson (1905) stated “Natal” in the original description, following the label attached to the holotype that reads “Natal. | G.A.K.Marshall. | 96–67. | 22-I-96 [partially handwritten]”, but the entry in the NHMUK accession register reads “Transvaal”. Moreover, Marshall had been collecting in Mashonaland, Zimbabwe and spent Christmas and New Year’s 1895 in Gadzema (Butler 1898), before starting on his journey back to the UK for a period of leave which involved taking a coach to Beira, Mozambique (Anonymous 1958) via Umtali and Chimoio (i.e. not via South Africa) during January 1896 (Marshall 1897). It is unclear exactly when in January Marshall was in Beira and how long he spent there, or whether he stopped off in Natal on his return home but there is a distinct possibility that this species originated in Mozambique. Janse (1937) had not seen another specimen in the subsequent two decades since its description and no further specimens were located in the collections examined by the present author.

Description of new species

Pterocerota anima **sp. n.** (Figs 121–122, 125)
https://www.zoobank.org/urn:lsid:zoobank.org:act:D0488AF2-D320-4462-9B8B-2AE4DF08A096

Holotype ♂ (Fig. 121):
“Kimboza Forest | Morogoro T.T. | April 1954 | E. Pinhey || NHMUK015929962 [QR code]” (NHMUK).

Paratypes (1 ♀):
TANZANIA: same data as holotype (1 ♀ NHMUK).

Description. Forewing lengths: male: 28 mm, female: 27 mm.
Male (Fig. 121). Upperside. Ground colour of body creamy-white, wings pearly-white and diaphanous. Antenna brown, bipectinate with long rami. Forewing elongate triangular, beige costal margin gently arcuate to rounded apex, outer margin gently curved to rounded tornus; hindwing elongate, outer margin curved. Postmedial fascia grey, well defined along veins where scales are densely packed, curved, its outer edge strongly crenulate. Submarginal and subterminal fasciae dark grey, curved following outer margin, crenulate; crenules of subterminal fascia reaching termen along veins. Underside. As on upperside but abdomen irrorated with yellow scales. Markings showing through from upperside. Male genitalia (Fig. 125). Uncus with pair of medial lobes, fused medially, and pair of lateral, heavily setose, digitate processes midway along. Tegumen broad. Mesal protrusions separated, triangular, very small and setose. Valve broadly triangular, divided into costal and saccular sections, fused at their base. Costal section rectangular gently tapering towards truncate apex. Saccular section elongate triangular, tapering to robust, sigmoidal distal process. Juxta semi-cylindrical, trapezoidal, bilobed distally. Vinculum narrow, V-shaped. Saccus short, rounded at apex. Phallus elongate, coecum rounded and slightly curved, tapering towards carina; vesica globular.

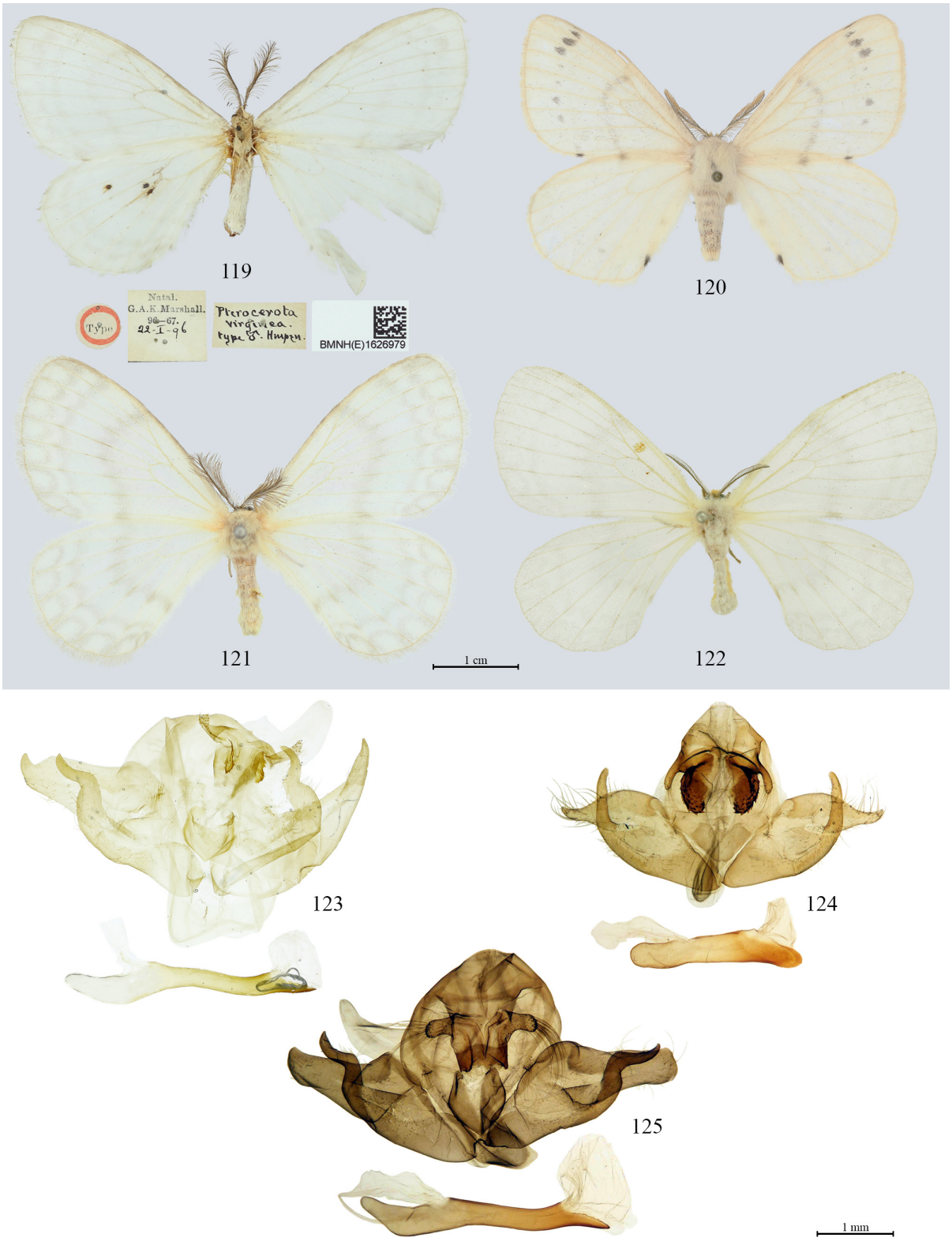
Female (Fig. 122). As in male but rami of antenna shorter, wings slightly more elongate and markings more weakly expressed.

Diagnosis. The new species is similar to *P. virginea* but the submarginal and subterminal fasciae are indistinct and weakly crenulate in the latter while they are strongly crenulate in *P. anima* **sp. n.**, with the crenules of the subterminal fascia reaching the termen. The male genitalia differ from both *P. virginea* and *P. perissinottoi* in the smooth medial lobes of the uncus (dentate in the other two) and the truncate apex of the costal section of the valve (pointed in the congeners).

Molecular characterisation. Unknown.

Derivatio nominis. The new species derives its name from the Latin for ‘spirit’ in reference to its ethereal appearance. The epithet is a noun in apposition.

Distribution. Known only from its type locality at the foothills of the Uluguru Mountains, Tanzania.



Figures 119–125. *Pterocerota* Hampson, 1905 species (119–122 habitus (in NHMUK unless otherwise stated), 123–125 ♂ genitalia). 119. *Pterocerota virginea* Hampson, 1905, holotype ♂. 120. *P. perissinottoi* (Bouyer, 2012) (ANHRT). 121. *P. anima* **sp. n.**, holotype ♂. 122. *Id*., paratype ♀. 123. *P. virginea*, holotype. 124. *P. perissinottoi*. 125. *P. anima* **sp. n.**, holotype.

Species excluded from *Stenoglene sensu lato*

The following two taxa were assigned to *Stenoglene* in the global Bombycoidea checklist of Kitching & Rougerie *et al*. (2018) but had previously been excluded from the genus and transferred to different families.

***Stenoglene bicolor* Distant, 1897**
Stenoglene bicolor Distant 1897: 206; Gaede 1927: 309; Kitching & Rougerie *et al*. 2018: suppl. 1.
Chrysopoloma bicolor (Distant, 1897): Distant 1903: 86, tab. 8, figs 2–3 (comb.); Janse 1917: 135; Hering 1928: 477, pl. 76, fig. b; Vári *et al*. 2002: 75; Krüger 2020: 62; Taberer & László 2022: 12, figs 38–39; Taberer 2023: 17, fig. 13.
Chrysopoloma similis f. *bicolor* (Distant, 1897): Hering 1937: 253, pl. 9, fig. 23 (stat.).

Type locality: [South Africa] "Transvaal, Pretoria, and Pienaars River".
Name-bearing type: 4 ♀ syntypes in NHMUK.

Notes. This taxon was transferred to *Chrysopoloma* Druce, 1888, a genus of Chrysopolomidae by Distant (1903).

***Stenoglene gemmatus* [sic] (Wichgraf, 1921)**
Phasicnecus gemmatus Wichgraf 1921: 19; Gaede 1927: 307.
Laeliopsis gemmatus (Wichgraf, 1921): Tams 1936: 96 (comb.).
Stenoglene gemmatus [sic] (Wichgraf, 1921): Dall'Asta & Poncin 1980: 484 (comb.); Kitching & Rougerie *et al*. 2018: suppl. 1.

Type locality: "Bailundo (Angola)".
Name-bearing type: holotype ♂ in NHMUK.

Notes. This taxon was transferred to *Laeliopsis* Aurivillius, 1911, a genus of Lasiocampidae by Tams (1936).

Summary of genera and their constituent species (listed alphabetically)

***Artemisana* gen. n.**
Artemisana thelda (Druce, 1887) **comb. n.**

***Atopojana* gen. n.**
Atopojana bouyeri (Basquin, 2013) **comb. n.**

***Cryptojana* gen. n.**
Cryptojana cinerea (Holland, 1893) **comb. n.**
Cryptojana clucki (Bouyer, 2012) **comb. n.**
Cryptojana decellei (Dall'Asta & Poncin, 1980) **comb. n.**
= *Stenoglene decellei flava* Dall'Asta & Poncin, 1980 **syn. n.**
Cryptojana grandiplaga (Holland, 1920) **stat. rev.**, **comb. n.**
Cryptojana insignifica (Rothschild, 1917) **comb. n.**
Cryptojana maculifera (Strand, 1912) **comb. n.**
Cryptojana noellae (Bouyer, 2012) **comb. n.**
Cryptojana plagiata (Aurivillius, 1911) **comb. n.**
Cryptojana preussi (Aurivillius, 1893) **comb. n.**
Cryptojana shabae (Dall'Asta & Poncin, 1980) **comb. n.**
Cryptojana volynkini **sp. n.**

***Drepanojana* Aurivillius, 1893**
Drepanojana fasciata Aurivillius, 1893

= *Drepanojana apicalis* Aurivillius, 1915
Drepanojana hogani **sp. n.**
Drepanojana maeonia (Druce, 1896) **stat. rev.**

***Epijana* Holland, 1893**
Epijana cosima (Plötz, 1880)
= *Epijana lanosa* Holland, 1893
= *Stenoglene velutonia* Druce, 1896
Epijana gigantea (Rothschild, 1917) **comb. n.**
= *Hoplojana purpurata* Wichgraf, 1921 **syn. n.**
Epijana latifasciata Dall'Asta & Poncin, 1980
Epijana meridionalis Dall'Asta & Poncin, 1980
Epijana vingerhoedti Bouyer, 2012

***Hecateana* gen. n.**
Hecateana dargei **sp. n.**
Hecateana giustii **sp. n.**
Hecateana kitchingi **sp. n.**
Hecateana paulisi (Dall'Asta & Poncin, 1980) **comb. n.**

***Lokiana* gen. n.**
Lokiana lydiae **sp. n.**
Lokiana sigyna (Aurivillius, 1893) **comb. n.**

***Microjana* gen. n.**
Microjana annae (Giusti & László, 2025) **comb. n.**
Microjana bouyeri **sp. n.**
Microjana brunneofasciata (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana citrina (Druce, 1886) **comb. n.**
Microjana citrinoides (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana dallastai (Bouyer, 2012) **comb. n.**
Microjana dehanica (Strand, 1911) **comb. n.**
Microjana dufranei (Bouyer, 2012) **comb. n.**
Microjana fontainei (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana fouassini (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana hikaru **sp. n.**
Microjana latimaculata (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana morettoi (Bouyer, 2012) **comb. n.**
Microjana novempunctata (Bethune-Baker, 1927) **stat. rev., comb. n.**
Microjana opalina (Druce, 1910) **comb. n.**
= *Phasicnecus peropalinus* Rothschild, 1917
Microjana parvula (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana pellucida (Joicey & Talbot, 1924) **comb. n.**
Microjana pujoli (Dall'Asta & Poncin, 1980) **comb. n.**
Microjana sinjaevi (Bouyer, 2012) **comb. n.**
Microjana smithi (László & Giusti, 2025) **comb. n.**
Microjana subiridescens (Holland, 1893) **comb. n.**
Microjana sulphureoides (Kühne, 2007) **comb. n.**
Microjana sulphureotincta (Strand, 1912) **comb. n.**
Microjana tabithae **sp. n.**
Microjana uelei (Dall'Asta & Poncin, 1980) **comb. n.**

***Neojana* gen. n.**
Neojana pseudosygina (Bouyer & Basquin, 2012) **comb. n.**

***Nepenthojana* gen. n.**
Nepenthojana uniformis (Dall'Asta & Poncin, 1980) **comb. n.**
Nepenthojana plenevauxae (Bouyer, 2012) **comb. n.**

***Pterocerota* Hampson, 1905**
Pterocerota anima **sp. n.**
Pterocerota perissinottoi (Bouyer, 2012) **comb. n.**
Pterocerota virginea Hampson, 1905

***Smithiana* gen. n.**
Smithiana bipartita (Rothschild, 1917) **comb. n.**
Smithiana smithi **sp. n.**

***Stenoglene* Felder & Felder, 1874**
Stenoglene basquini Bouyer, 2012
Stenoglene bipunctata (Aurivillius, 1909)
Stenoglene elgonae (Bethune-Baker, 1927) **stat. rev.**
Stenoglene gregorii (Butler, 1894) **stat. rev.**
Stenoglene hilaris Felder & Felder, 1874
= *Chrysopoloma bithynia* Druce, 1887
Stenoglene inversa (Gaede, 1927)
Stenoglene kenyensis Bouyer, 2012
Stenoglene labda (Druce, 1887) **stat. rev.**
= *Phasicnecus livingstonensis* (Strand, 1909) **syn. n.**
Stenoglene leonardi Bouyer, 2012
Stenoglene lucindae **sp. n.**
Stenoglene obtusa (Walker, 1865)
= *Stenoglene tristis* Felder & Felder, 1874
= *Phasicnecus evanescens* Distant, 1903
= *Phasicnecus subcroceus* Distant, 1903
Stenoglene pira Druce, 1896
= *Phasicnecus aequidistans* Strand, 1915 **syn. n.**
Stenoglene rosea (Druce, 1886)
= *Phasicnecus monteironis* Rothschild, 1917
Stenoglene sarahae **sp. n.**
Stenoglene similis (Rothschild, 1917) **stat. rev.**
= *Phasicnecus similis flavidior* Rothschild, 1917 **syn. rev.**
= *Phasicnecus pulverulentus* Joicey & Talbot, 1924 **syn. n.**
Stenoglene susanae **sp. n.**

***Streptojana* gen. n.**
Streptojana brunnea (Bouyer, 2012) **comb. n.**
Streptojana forbesi **sp. n.**
Streptojana laurenae **sp. n.**
Streptojana molliae **sp. n.**
Streptojana nivalis (Rothschild, 1917) **comb. n.**

***Urojana* Gaede, 1915**
Urojana basquini **sp. n.**
Urojana eborea Gaede, 1915

Discussion and conclusions

Since Gaede's (1927) revision of the Eupterotidae, *Stenoglene* has seemingly been treated as a "catch-all" genus, with members displaying great phenotypic disparity. In spite of Forbes' (1955) remark that the male genitalia were "so extremely plastic that groupings are not clear", the phenotypic and genitalic diversity has never been critically investigated, with all recent works on *Stenoglene* and *Epijana* focusing on descriptions of new species rather than defining generic boundaries.

This present review of *Stenoglene* and allied genera initially revealed a convoluted picture but some stability has been brought about through the examination and dissection of a large number of specimens, and the integration of morphological and molecular data. Phylogenetic inferences recovered *Stenoglene* as polyphyletic with regard to *Drepanojana*, *Urojana* and *Epijana*, and in order to maintain monophyly, two options were considered – either synonymising the allied genera under *Stenoglene*, or partitioning the genus into discrete diagnosable generic units. Following further morphological analyses of phenotype and genitalia, it was concluded that a single genus to encompass this speciose and heterogenous assemblage would be highly inadequate and to this end, the generic boundary of *Stenoglene sensu str.* was redefined and 10 new genera have been delimited. Concurrently, this review has uncovered a cryptic diversity of species and 18 new species have been described. Moreover, seven taxa have been revived from synonymy, 44 new combinations proposed, and six synonymies have been newly established or revived. It has been demonstrated that the differing phenotype, plasticity of the genitalia structures and uninformative venation reported by Forbes (1955) can be reconciled by the newly-implemented taxonomic rearrangement and the fact that the taxa he examined have now been assigned to numerous different genera. The male genital configurations of each genus formed neat compact groups with the exception of *Epijana*, which varied considerably but the genus was confidently circumscribed through a combination of venation and genetics. It was found that only three genera share the unstalked R_4 and R_5 veins of the forewing, and although *Epijana* and *Urojana* were consistently recovered as siblings, it is believed that *Pterocerota* does not form a clade with these two genera due to differing genitalia and origin of forewing vein M_1.

It must be stated that in some cases, the generic partitioning outlined above is not perfect and further investigations within a molecular and morphological framework are recommended to improve taxonomic clarity. From the molecular perspective, only barcodes were utilised in this study and analyses of other gene fragments will doubtless provide additional insights into relationships at the supraspecific level. Furthermore, although taxon sampling was fairly complete, important taxa such as *Pterocerota* were missing and their inclusion in future analyses would be a priority. From a morphological perspective, there are some genera described above that could perhaps be split even further (e.g., *Streptojana* **gen. n.**) based on phenotypic differences, but some caution has been exercised especially when not all the data has been available. Despite these limitations, all genera circumscribed above are monophyletic assemblages, each clearly diagnosable by a combination of characters and provide a much-needed framework for future revisionary work (e.g., *Microjana* **gen. n.**).

At the species level, the genitalia displayed highly diagnostic interspecific characters despite identical phenotypes (e.g., *Smithiana* **gen. n.**) and it is believed that there will be further new species that await description. In accordance with recent observations in other Afrotropical Janinae (Takano 2024a, 2025), barcode divergences alone were insufficient to distinguish between species, with conspecific populations in some species diverging by over 6% (e.g., *Artemisana thelda*), while morphologically distinguishable sibling taxa diverged in some cases by only 1.5% (e.g., *Urojana*).

The accurate delimitation of the genera has also shed further insights into the biogeography of this group of insects. Previously considered to be widely distributed throughout the African continent, the majority of the newly-defined genera are in fact restricted to the Guineo-Congolian rainforests (to include Kakamega Forest, Kenya in the east – see Wagner *et al*. (2008) for its biogeographical assignment), with the exception of five genera. *Stenoglene sensu str.* is widely distributed throughout sub-Saharan Africa where the majority of species are found in woodland and forest-savanna mosaics, with notable exceptions such as *S. bipunctata* and *S. leonardi* having successfully colonised afforested regions from more open habitats. In *Cryptojana* **gen. n.**, *Microjana* **gen. n.** and *Hecateana* **gen. n.**, the opposite is the case where the majority are Guineo-Congolian rainforest species. *Pterocerota* is the only genus with its constituent species distributed solely in east and south-east Africa. For species of the Guinean forests of West Africa, speciation has seemingly occurred across the xeric stretch of the

Dahomey Gap, a well-known barrier to gene flow, and in many cases, sibling species are found on either side (e.g., *Lokiana* **gen. n.**, *Urojana*, etc.). Perhaps of more interest are the assemblages of sibling taxa around the Sangha River Interval in Central Africa (e.g., *Microjana citrina* and *M. tabithae* **sp. n.**), where speciation has seemingly occurred in forest and fluvial refugia split by a xeric corridor during the late Holocene. The growing body of work where similar distributional patterns of sibling taxa have been observed in this region of Central Africa is summarised in Takano (2024b).

This present body of work dealing with *Stenoglene* and allied genera is a continuation of recent papers dealing with Janinae genera based on material housed in the ANHRT collections (Takano 2024a, 2025; Takano & Morgan 2025) and it is hoped that this taxonomic rearrangement not only enhances stability but facilitates future research on this group of moths.

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